The Deoxyribonucleases of Escherichia coli

III. STUDIES ON THE NATURE OF THE INHIBITION OF ENZYMATIC INHIBITION OF ENZYMATIC ACTIVITY OF CELL-FREE EXTRACTS OF ESCHERICHIA COLI BY DEOXYRIBONUCLEASE BY RIBONUCLEIC ACID*

I. R. LEHMAN, G. G. ROUSSOS,† AND E. A. PRATT

From the Department of Biochemistry, Stanford University School of Medicine, Palo Alto, California

(Received for publication, August 2, 1961)

It was first observed by Kozloff (1) that the deoxyribonuclease activity of cell-free extracts of Escherichia coli could be increased by treatment with pancreatic ribonuclease. This stimulation was found to result from the destruction by the ribonuclease of an inhibitory ribonucleic acid. The phenomenon of cellular deoxyribonuclease inhibition is a widespread one, having been observed in yeast (2), mammalian tissues (3), and streptococci (4). The yeast and mammalian deoxyribonuclease inhibitors were identified as proteins; the streptococcal inhibitor was shown to have the properties of a ribonucleic acid.

Recent investigations of the nucleases of Escherichia coli (5, 6) have shown that the deoxyribonuclease activity in cell-free extracts of this organism is attributable to at least three different enzymes. Two of these have now been purified. It was, therefore, of interest to establish which of these enzymes is subject to inhibition by ribonucleic acid and what is the nature of the ribonucleic acid-enzyme interaction. The studies reported here show that the deoxyribonucleic acid-specific endonuclease described in the previous paper (6) is the only one of the known nucleases of E. coli which can be inhibited by ribonucleic acid. They further demonstrate that ribonucleic acids from a variety of sources, including E. coli, guinea pig liver, and tobacco mosaic virus, can serve as potent inhibitors of the purified endonuclease.

EXPERIMENTAL PROCEDURE

Materials and Methods

The E. coli endonuclease (carboxymethyl (CM) cellulose and Amberlite IRC-50 (XE-64 fractions) and DNA were prepared as described in the previous paper (6). Crystalline pancreatic RNase and DNase were purchased from the Worthington Biochemical Corporation. Crystalline horse heart cytochrome c was obtained from the Sigma Chemical Company. Crystalline bovine plasma albumin was obtained from the Armour Laboratories. Snake venom phosphodiesterase was prepared by the method of Sinshheimer and Koerner (7). E. coli phosphodiesterase was purified as described previously (5). Amino acid acceptor RNA and ribosomal RNA from E. coli (8, 9), amino acid acceptor RNA from guinea pig liver (10), and tobacco mosaic virus RNA (11) were gifts from Dr. Paul Berg. Polyadenylate and poly AGUC,1 both synthesized by means of poly-nucleotide phosphorylase from Azotobacter vinelandii (12), were kindly provided by Dr. Severo Ochoa. High molecular weight polyphosphate prepared by the method of Pfanstiel and Iler (13) was the gift of S. R. Kornberg.

Assay of Endonuclease Inhibitor—This assay measures the inhibition by RNA of the formation of acid-soluble products from P32-labeled DNA by the E. coli endonuclease. The reaction mixture (0.3 ml) contained 20 pmoles of Tris buffer, pH 7.5; 2 pmoles of Mgl2; 46 µg of cytochrome c; P32-labeled E. coli DNA, 20 mpmoles of P (1 µc per pmoles of P); 0.2 to 1.0 unit of RNA inhibitor; and 0.04 unit of Amberlite XE-64 fraction of endonuclease (6). As a rule, three levels of inhibitor were run for each assay. After incubation for 30 minutes at 37°, acid-soluble P32 was measured as previously described (6). Alternatively, residual nuclease activity could be determined by observing the development of hyperchromicity as described before (6). One unit of inhibitor is defined as that amount which produces a decrease in rate such that 1/V - 1/V0 is equal to 1, where V is the velocity of reaction in the presence of inhibitor and V0 is the velocity in its absence, when measured under the specified conditions of the assay.

RESULTS

Addition to the purified endonuclease of a crude extract of E. coli which had been heated at 100° for 20 minutes resulted in complete inhibition of nuclease activity. This inactivation was specifically attributable to the RNA in the boiled extract, since it could be alleviated by pretreatment of the extract with pancreatic RNAase (Table I).

There are two major RNA components in E. coli, the lower molecular weight amino acid acceptor RNA, and a larger RNA component found in the ribosomes (8, 9). Both were found to be potent inhibitors of the endonuclease.

Kinetics of Inhibition by Amino Acid Acceptor RNA—A detailed examination of the kinetics of inhibition by amino acid acceptor RNA showed it to be a competitive inhibitor of the endonuclease. The dissociation constant of the enzyme-inhibitor complex (Kd) calculated by the method of Lineweaver and Burk (14) (Fig. 1) was 9 X 10^-9 M (expressed as moles of

* This work was supported in part by a grant from the United States Public Health Service.
† Present address, Radiisotope and Research Laboratory, Veterans Administration Hospital, Kansas City, Missouri.
‡ The abbreviation used is: poly AGUC, copolymer composed of adenylie, guanylic, cytidylic, and uridylic acids.
**Table I**

**Demonstration of inhibitory RNA in boiled extract of *E. coli***

The incubation mixtures were set up as described under "Materials and Methods." Boiled extract was prepared by heating a crude extract at 100° for 20 minutes. After chilling at 0°, precipitated protein was removed by centrifugation for 10 minutes at 10,000 X g. The supernatant fluid was diluted 50-fold, and 0.02 ml, containing 0.11 optical density unit at 260 mp, was added as indicated. RNase treatment was carried out in an incubation mixture (0.2 ml) containing 0.02 ml of boiled extract (5.5 optical density units at 260 mp); 15 μmoles of Tris buffer, pH 7.5; and 50 μg of RNase. After incubation for 30 minutes at 37°, the solution was diluted 5-fold, and 0.02 ml was tested for inhibition in the standard assay. The CM-cellulose fraction of endonuclease was used in these experiments.

<table>
<thead>
<tr>
<th>System</th>
<th>DNA-P made acid-soluble (μmoles)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Endonuclease</td>
<td>3.18</td>
</tr>
<tr>
<td>Endonuclease + boiled extract</td>
<td>&lt;0.10</td>
</tr>
<tr>
<td>Endonuclease + RNase-treated boiled extract</td>
<td>3.21</td>
</tr>
</tbody>
</table>

**Fig. 1.** Competitive inhibition of *E. coli* endonuclease by amino acid acceptor RNA from *E. coli*. The numbers in parentheses give the concentration of amino acid acceptor RNA in micromicro-moles of P per ml. 1/V is the reciprocal of the millimicromoles of DNA-P made acid-soluble in 30 minutes.

RNA nucleotide. This is to be compared with a value of 2.5 × 10⁻⁴ M, which is the *K*ₗ₀ observed for DNA (expressed as mol of DNA nucleotide). A plot of the reciprocal of the reaction velocity against inhibitor concentration according to the method of Dixon (15) gave a value of 1.0 × 10⁻⁴ M for the *K*₀ which is in good agreement with that calculated from the Lineweaver-Burk expression (Fig. 2). Moreover, this type of plot provided a means of assaying quantitatively the inhibitor activity. When 1/V - 1/Vₒ is plotted against concentration of Inhibitor I, a linear relationship is obtained (Fig. 3).

Magnesium ion is required for inhibition of the endonuclease by amino acid acceptor RNA. Thus, when the magnesium ion concentration normally present in the incubation mixture is reduced by a factor of 100 and cytochrome c is omitted, inhibition is no longer observed. It can be partially restored by the addition of cytochrome c (Table II).

**Susceptibility of Inhibitor to RNase and Snake Venom Dipeptidase—**Pretreatment of purified amino acid acceptor RNA with RNase results in a loss of its activity as an inhibitor of the endonuclease (Fig. 4). At a concentration of RNase (3.7 × 10⁻⁴ μg per ml) at which 50% of the RNA was converted to acid-soluble products, essentially all inhibitor activity was lost.

The inhibitor is also destroyed by the action of snake venom phosphodiesterase, another enzyme which attacks RNA. In this case, when 23% of the RNA had been converted to acid-soluble fragments by the action of this enzyme, a 34% decrease in inhibitory activity was observed. When 40% of the RNA had become acid-soluble, only 25% of the inhibitor activity remained. It is noteworthy that the endonuclease inhibitor activity of this RNA is considerably less sensitive to hydrolysis by these two enzymes than is its amino acid-binding activity. For example, venom phosphodiesterase digestion of the RNA results in essentially complete loss of amino acid acceptor activity when as few as 5% of the nucleotides have been cleaved off (16). Since this enzyme carries out an exonucleolytic attack beginning at the 3'-hydroxyl end of the RNA chain (17), the very rapid inactivation of acceptor activity is consistent with the known requirement for an adenosine residue at this end of the chain (18). The

**Fig. 2.** Determination of *K*_ᵢ for amino acid acceptor RNA from *E. coli*. The numbers in parentheses are the concentrations of DNA in millimicromoles of P per ml. 1/V is the reciprocal of the millimicromoles of DNA-P made acid-soluble in 30 minutes.
presence of a specific residue at this end of the RNA molecule is clearly not essential for its activity as an inhibitor of the E. coli endonuclease.

Inhibitor Activity of Other RNA's—Examination of a variety of RNA preparations showed the amino acid acceptor RNA from E. coli to be the most active of those tested; however, all the RNA's examined were to some extent active (Table III). Amino acid acceptor RNA from guinea pig liver showed approximately one-third as much activity as the corresponding RNA from E. coli. High molecular weight RNA from E. coli ribosomes or tobacco mosaic virus was also approximately one-third as active as the E. coli acceptor RNA when compared per mole of RNA rather than per mole of nucleotide. In contrast, the RNA-like polymers synthesized by means of polynucleotide phosphorylase were relatively inert. Poly AGUC showed only slight inhibitor activity, and polyadenylate, even at very high concentrations, was completely inactive. Examination of both of these polymers in the ultracentrifuge showed them to be polydisperse ma-

Table II

<table>
<thead>
<tr>
<th>Magnesium concentration</th>
<th>Acid-soluble nucleotide formed</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>-Inhibitor</td>
</tr>
<tr>
<td>moles/liter</td>
<td></td>
</tr>
<tr>
<td>7 × 10⁻⁴</td>
<td>2.0</td>
</tr>
<tr>
<td>7 × 10⁻⁵</td>
<td>0.9</td>
</tr>
<tr>
<td>7 × 10⁻⁶</td>
<td>0.7</td>
</tr>
</tbody>
</table>

* Cytochrome c (40 μg) was added to the incubation mixture.

We are grateful to Dr. Ross Inman for performing these studies.

Table III

<table>
<thead>
<tr>
<th>Inhibitor</th>
<th>Approximate molecular weight</th>
<th>Inhibitor units per mpmole of RNA</th>
<th>Inhibitor units per mpmole of RNA</th>
</tr>
</thead>
<tbody>
<tr>
<td>Amino acid acceptor RNA (E. coli)</td>
<td>3 × 10⁴ (8)</td>
<td>30</td>
<td>3000</td>
</tr>
<tr>
<td>Ribosomal RNA (E. coli)</td>
<td>1 × 10⁸ (9)</td>
<td>0.4</td>
<td>1200</td>
</tr>
<tr>
<td>Tobacco mosaic virus RNA</td>
<td>2 × 10⁸ (11)</td>
<td>0.2</td>
<td>1200</td>
</tr>
<tr>
<td>Amino acid acceptor RNA (guinea pig liver)</td>
<td>3 × 10⁴ (10)</td>
<td>14</td>
<td>1400</td>
</tr>
<tr>
<td>Poly AGUC</td>
<td>1 × 10⁴</td>
<td>0.1</td>
<td>30</td>
</tr>
<tr>
<td>Polyadenylate</td>
<td>1 × 10⁴</td>
<td>&lt;0.002</td>
<td>&lt;0.5</td>
</tr>
<tr>
<td>Polyphosphate</td>
<td>&gt;1 × 10⁴</td>
<td>&lt;0.002</td>
<td>&lt;0.5</td>
</tr>
</tbody>
</table>

Fig. 4. Destruction of inhibitor activity of amino acid acceptor RNA from E. coli by pancreatic RNases. The incubation mixtures (0.3 ml) contained 10 μmoles of Tris buffer, pH 7.5; 0.5 mg of crystalline bovine plasma albumin; 60 mmoles of RNA-P, and the indicated amounts of RNase (diluted in albumin, 1 mg per ml). After 30 minutes at 37°C, 0.02 ml aliquots were removed for assay of inhibitor activity. To 0.2 ml of the remainder were added 0.01 ml of a solution of albumin (50 mg per ml), 0.29 ml of water, and 0.5 ml of 1 N perchloric acid. The tubes were kept at 0°C for 5 minutes, then centrifuged at 10,000 × g for 5 minutes. The optical density of the supernatant fluids at 260 nm was determined. In calculating the percentage of the total optical density made acid-soluble, no correction was made for the hyperchromic effect resulting from the hydrolysis of the RNA.
terials with approximate weight average molecular weights of about $1 \times 10^4$. Polyphosphate, another high molecular weight polymer, was found to be inert as an inhibitor of the endonuclease. On the contrary, this compound caused a 3-fold stimulation in endonuclease rate when added in concentrations as high as 20 μmoles of P per ml.

**Specificity of RNA for Endonuclease**—Other enzymes which attack DNA or polydeoxynucleotides are unaffected by amino acid acceptor RNA from *E. coli*. Incubation of pancreatic DNase (1 unit) or *E. coli* phosphodiesterase (1 unit) each with 10 μmoles of amino acid acceptor RNA-P did not diminish their nuclease activity. Snake venom phosphodiesterase was similarly unaffected by the presence of RNA when assayed for its activity on a mixture of deoxyribonucleotides.

Inhibition of the *E. coli* endonuclease, which is a basic protein (6), by amino acid acceptor RNA is not affected by the presence in the incubation mixture of a very large excess of another basic protein, cytochrome c, which is routinely added to stabilize the endonuclease. Thus, the same inhibition in the activity of 0.01 μg of enzyme protein (approximately 60%) by amino acid acceptor RNA was observed in the presence or absence of 40 μg of cytochrome c.

**DISCUSSION**

Of the deoxyribonucleases now known to be present in *E. coli*, the endonuclease described here and in the previous paper (6) is the only one which is sensitive to inhibition by RNA. Although many RNA's are capable of inhibiting the purified *E. coli* endonuclease, the inhibition exhibits certain specific features. Thus, the phenomenon appears to be confined to an interaction between the *E. coli* endonuclease and naturally occurring RNA. On the one hand, other nucleases which attack DNA, such as pancreatic DNase and the *E. coli* and snake venom phosphodiesterases, are unaffected by RNA. Moreover, proteins, such as cytochrome c, which have dimensions and ionic properties similar to the *E. coli* endonuclease do not interfere with the inhibition of the endonuclease by RNA, even when they are present in very great excess. On the other hand, the enzymatically synthesized polymers which resemble RNA in a number of respects (12) appear to lack features characteristic of the naturally occurring RNA's which permit the natural RNA's to act as inhibitors.

It should be pointed out that, although the amino acid acceptor and ribosomal RNA's from *E. coli* are both potent inhibitors of the purified *E. coli* endonuclease, the possibility cannot be excluded that there exist within the cell other, more potent, RNA's whose specific function is, indeed, the immobilization of this nuclease.

In this regard, the relevance of the RNA inhibition of the purified endonuclease to the deoxyribonuclease activity of extracts of phage-infected bacteria deserves some comment. It was first demonstrated by Pardee and Kunkee (19) that extracts of *E. coli* infected with bacteriophage T2 exhibit a 2- to 3-fold greater deoxyribonuclease activity than uninfected cell extracts. This increase in total deoxyribonuclease activity was attributed by Kozloff (1) to a partial destruction of an inhibitory RNA, which he discovered as a result of these studies. The experiments reported here would suggest that the increase in deoxyribonuclease activity following phage infection is not entirely attributable to the destruction of a specific RNA. Thus, it is conceivable that if the RNA which normally binds the endonuclease within the cell were in some way destroyed as a result of the infection, the amino acid acceptor or ribosomal RNA's, both potent inhibitors of the endonuclease, could immediately bind the endonuclease, rendering it inactive.

Recent studies of the enzymatic activities of phage-infected cells (21) have demonstrated the induction, as a result of the infective process, of new enzymes specific to the infected cell. In addition, the increased activity of enzymes already present at the time of infection has been observed (21, 22). It is, therefore, not inconceivable that the increase in over-all deoxyribonuclease activity found in phage-infected cells may in fact be due to an increased synthesis of existing nuclease, or perhaps to synthesis of deoxyribonuclease unique to the phage-infected bacterium.

**SUMMARY**

The *Escherichia coli* endonuclease is found in extracts bound to an inhibitory ribonucleic acid.

1. When purified, this enzyme is free of inhibitor and can be inhibited by a variety of ribonucleic acids from *Escherichia coli*, guinea pig liver, and tobacco mosaic virus. Of those tested, the amino acid acceptor ribonucleic acid from *Escherichia coli* is the most active. The kinetics of the enzyme inhibited by the amino acid acceptor ribonucleic acid obey the equations for a competitive inhibition, and the calculated Kᵢ is of the order of $1 \times 10^{-4}$ M (ribonucleic acid nucleotide).

2. The ribonucleic acid-like polynucleotides synthesized by polynucleotide phosphorylase are essentially inactive as inhibitors, as is high molecular weight polyphosphate.

3. The activity of other deoxyribonucleic acid-cleaving enzymes, including pancreatic deoxyribonuclease and the *E. coli* and snake venom phosphodiesterases, is not affected by ribonucleic acid.

4. Other basic proteins such as cytochrome c, when present in great excess, do not prevent the inhibition of the endonuclease (also a basic protein) by amino acid acceptor ribonucleic acid.

The relevance of these observations to the changes in over-all deoxyribonuclease activity following phage infection of *Escherichia coli* is discussed.

**REFERENCES**


---

4 A unit of pancreatic DNase activity is defined as the amount which will produce 0.1 μmole of acid-soluble DNA nucleotide in 30 minutes under the conditions used for assay of the *E. coli* endonuclease.

5 The level of RNA in *E. coli* following infection with the T-even bacteriophages remains relatively unchanged (20).
The Deoxyribonucleases of *Escherichia coli* : III. STUDIES ON THE NATURE OF THE INHIBITION OF ENDONUCLEASE BY RIBONUCLEIC ACID

I. R. Lehman, G. G. Roussos and E. A. Pratt


Access the most updated version of this article at http://www.jbc.org/content/237/3/829.citation

Alerts:

- When this article is cited
- When a correction for this article is posted

Click here to choose from all of JBC's e-mail alerts

This article cites 0 references, 0 of which can be accessed free at http://www.jbc.org/content/237/3/829.citation.full.html#ref-list-1