Crystallization and Characterization of an Esteroproteolytic Enzyme from Porcine Pancreas*

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An earlier paper (1) from this laboratory described the presence of a proesterase in porcine pancreas which could be activated by trypsin. The activity of the esterase toward esters of o-hydroxybenzoic acid was studied. The impure enzyme appeared to be different from other pancreatic enzymes known to exist as zymogens: the chymotrypsins, the carboxypeptidases, trypsin from bovine pancreas, and elastase from porcine pancreas. This report describes the isolation and crystallization of the active enzyme and a preliminary study of some of its properties. Crystalline preparations of the enzyme have been found to possess proteolytic as well as esteratic activity. An estimate of the requirement for the degree of Doctor of Philosophy.

EXPERIMENTAL PROCEDURE

Materials—The enzyme substrate valeryl salicylate (o-(n-pentoyloxy)benzoic acid) was prepared as described earlier (1). N-Acetyl-L-tyrosine ethyl ester and poly-L-glutamic acid were obtained from Mann Research Laboratories, Inc. and Schwarz BioResearch, Inc., respectively. Diisopropyl fluorophosphate was purchased from Delta Chemical Works. DEAE-cellulose, 1.1 meq per g, was obtained from Brown Company. Crystalline preparations of chymotrypsin B and chymotrypsinogen B were a gift from Dr. M. Laskowski, Marquette University.

Enzymatic Assay Methods—The principle of the determination of esterase activity by means of esters of hydroxybenzoic acid has been described by Hofstee (2). The substrate used in this work was valeryl salicylate. For impure preparations of the enzyme the procedure described earlier (1) was used. For purified preparations of the enzyme the activity was determined by a continuous method as follows. One milliliter of the enzyme solution was placed in a 25-ml Erlenmeyer flask containing 3 ml of 0.4 M Tris buffer at pH 8.65, and 5 ml of water were added. The solution was equilibrated in a water bath at 37°. One milliliter of a freshly prepared solution of substrate, 0.15 M in 50% ethanol, was then added. An aliquot of the mixture was used for the determination of optical density at 300 mμ in a Beckman model DU spectrophotometer equipped with thermostatic constant temperature control at 37°. Readings were made at 3- to 5-minute intervals and corrections made for spontaneous hydrolysis as well as for absorption at 300 mμ due to substances other than reaction products. Excess at high enzyme concentrations, the rate of hydrolysis remained constant for 30 minutes.

Determinations of esterase activity with N-acetyl-L-tyrosine ethyl ester as substrate were done by the pH-Stat method (3). The equipment used was a Radiometer unit consisting of a Titirgraph (SBR2/SBU1) and a Type III automatic titrator. The substrate was dissolved in water and the final volume of the reaction mixture was 15 ml. No calcium salt was added. The titrating agent was 0.08 N NaOH. The pH of the reaction mixture was maintained between 8.05 and 8.10 at 37°. No corrections were made for spontaneous hydrolysis, which was negligible. The rate of hydrolysis was determined from the slope of the curve during the 1st minute of reaction.

Proteolytic activity was determined with casein and poly-L-glutamic acid as substrates. With casein as substrate the proteolytic activity was determined by the Kunitz method (4) as modified by Wu and Laskowski (5).

Crystalline as well as amorphous preparations of the esterase catalyze the hydrolysis of poly-L-glutamic acid.¹ The optimal pH for the hydrolysis of this substrate is between 4.6 and 5.6. The kinetics of hydrolysis was studied with the pH-Stat at pH 5.3 and at a temperature of 37° by titration with 0.1 N NaOH, and by paper chromatography with a mixture of butanol, acetic acid, and water as the solvent. The reaction mixture contained 20 μg of the poly-L-glutamic acid and 300 μg of enzyme protein in 5 ml. Aliquots of the hydrolysate were applied to paper strips by a striping technique. The paper strips were then developed by three successive passages of the solvent. The chromatograms after treatment with ninhydrin followed by Cu(NO₃)₂ were scanned with a Photovolt Varicord integrator to determine the relative amounts of components formed. Aliquots of the hydrolysate chromatographed at various times

¹ A detailed study of the enzymatic hydrolysis of poly-L-glutamic acid will be the subject of a later communication.
during the reaction always showed ten to twelve components, none of which was free glutamic acid. Both chromatographic and titration data showed that after 20 minutes under the above conditions, no further degradation occurred.

Electrophoresis—Electrophoretic analyses were done by the starch gel method of Smithies (6), and by the free boundary method.

In the starch gel procedures 0.02 M borate buffer, pH 9.0, was used to prepare the starch gel. The pH of the finished slab was between 8.4 and 8.6. Preparations of protein were applied to a slit in the horizontal slab by means of filter paper strips. The voltage gradient was approximately 5 volts per cm, and the time of electrophoresis was 15 to 17 hours. The temperature during electrophoresis was maintained between 2 and 5°C by circulation of fluid from a low temperature ethanol-water bath through the bed of the starch slab.

Fractionation of pancreatic extracts on the DEAE-cellulose column was followed routinely by means of starch gel electrophoresis. In order to determine which of the components was active, assays were made on extracts from the starch slab. Sections in an unstained slab corresponding to protein sections in a stained slab were removed and homogenized in a Potter-Elvehjem homogenizer with Tris buffer, pH 8.6, at 0-5°C. The suspension was centrifuged at 10,000 × g for 20 minutes at 0°C, which removed almost all the starch, and the supernatant fluid was assayed using valeryl salicylate as substrate.

An American Instrument Company portable electrophoresis apparatus was used for free boundary electrophoresis studies. Acetate buffers were used at various pH values and all at an ionic strength of 0.1. The temperature was 1.5°C, and the potential gradient varied from 5.32 to 5.83 volts per cm. All preparations of the enzyme used for analysis had been crystallized. Because of the low solubility of the enzyme at pH values close to its isoelectric point (at pH 4.3, maximal solubility is 1.5 mg of protein per ml), the concentration of protein used in the determination of the isoelectric point was 0.15%.

Preparation of Enzyme—Porcine pancreas was placed on ice within 10 to 20 minutes after the animal had been killed and was subsequently held at -30°C for several weeks before the tissue was processed. The pancreas was minced and then homogenized in a Waring Blender at 0°C with 4 times its weight of water. The pH was adjusted to 3.6 to 3.8 and the mixture was then filtered at 0-5°C.

Preliminary work showed that, when the pH 3.8 extracts of porcine pancreas were fractionated on a DEAE-cellulose column with gradient elution from pH 8 to pH 1, essentially all enzymatic activity, with valeryl salicylate as substrate, was recovered in the last fraction emerging from the column, at a pH between 5 and 4. This behavior of the enzyme on the column permitted the development of a preparative procedure for large amounts of the pancreatic extract. Since preliminary work showed some loss of activity when the time of fractionation was protracted and the pH of the system was high, the following procedure described for the preparative column fractionation was completed within 24 hours and the pH of the system was lowered as soon as possible.

DEAE-cellulose, 66 g, was equilibrated in a column with potassium phosphate buffer at pH 6.0 and ionic strength 0.005. A single mixing chamber with a volume of 500 ml was used, and all operations were performed at 0-5°C. The pH 3.6 extract from 700 g of porcine pancreas was adjusted to pH 6.0 and centrifuged. The supernatant liquid, 2550 ml, was then added to the column. The buffers and their sequence were 1 liter of potassium phosphate buffer at pH 6.0 and ionic strength 0.005, and the following sodium acetate buffers (all at ionic strength 0.1): 4 liters at pH 5.6, 3 liters at pH 4.8, 0.5 liter at pH 4.5, and 3 liters at pH 3.9. The flow rate was 10 to 20 ml per minute until the effluent pH had decreased to 4.5, at which time the collection of 18-ml samples was begun, and the flow rate was decreased to 5 to 10 ml per minute. With the above sequence of buffers, the enzyme emerges from the column as the pH of the effluent decreases below 4.5. The location of the enzyme in the effluent fractions was determined simply by optical density readings at 280 μm. The fractions containing the enzyme were combined and the solution was saturated with (NH₄)₃SO₄ or NaCl. The resulting precipitate was suspended in 20 to 25 ml of water in a Visking cellulose sac and dialyzed against water at 0-5°C to remove the salt.

In order to investigate the active fraction further, it was again chromatographed on a DEAE-cellulose column. The dimensions of the column were 45 × 1.2 cm; it contained 4.0 to 4.1 of DEAE-cellulose. The column was equilibrated first with a pH 6.0 phosphate buffer, ionic strength 0.005. The active fraction (55 ml, pH 6.0) containing 28.64 mg of N was then added. The effluent pattern was developed with a series of acetate buffers and which contained 5 to 10 mg of protein per ml, ionic strength 0.005. The fractionation was performed as follows. A suspension of the enzyme, which had been dialyzed exhaustively against water, and which contained 5 to 10 mg of protein per ml, is brought into solution by adjusting the pH to 5.8 to 6.2 by the addition of a few drops of borate buffer (pH 7.85, 0.11 M). The solution is filtered and then dialyzed against acetate buffer, pH 4.2 and ionic strength 0.005. The dialyzate after a few hours yields a heavy opalescent suspension which, after several weeks at 0°C, precipitated as a heavy white powder. The precipitate was removed by centrifugation at 10,000 × g for 20 minutes at 0-5°C.}

![Fig. 1. Chromatographic effluent diagram from a DEAE-cellulose column of the active fraction obtained from the preparative column. (See text.) Left ordinate, absorbancy at 280 μm. Right ordinate, pH of effluent. Abcissas, volume in liters and fractions combined for analysis (8 see Table 1).](image-url)
and frequent changes of the buffer, gradually changes into large crystals (Fig. 2). Crystallization was also obtained in the presence of ammonium sulfate at pH 4.3; needles were formed. However, in a number of attempts to repeat this type of crystallization, only occasionally were we successful.

Different preparations of the enzyme crystallized under identical conditions have yielded crystals as depicted in Fig. 2. The electrophoretic properties, as well as activities toward the various substrates, appear to be the same for all the crystalline preparations.

RESULTS AND DISCUSSION

An effluent diagram obtained by the chromatography of the active fraction is shown in Fig. 1. Inspection of this figure shows that practically all the protein is eluted within a pH range of 4.6 to 4.3. The main component (Fractions V and VI) accounts for 50% of the total nitrogen added to the column. This component has been collected in several experiments and crystallized (Fig. 2). The specific activities, as well as recoveries of activities in the fractions indicated in Fig. 1, are shown in Table I. The data indicate that no further separation of the enzymatic activities, present in the active fraction, was obtained by a re-chromatography; this suggests that the activity toward these substrates is associated with the same protein or proteins. These data show the possibility that the enzyme may exist in several forms. Fig. 3 shows the effluent diagram obtained by chromatography of the esterase which had been subjected to three crystallizations. The column and elution procedures were the same as those in Fig. 1 and are described in the text above. The pattern suggests some heterogeneity, which may be due to anomalous effects of the column. This result is in contrast to the electrophoretic analysis (see below). In the latter case both starch gel and free boundary electrophoreses indicate the preparation to be homogeneous after three crystallizations. The experimental work to date leads us to believe that this heterogeneity may be due to slightly different forms of the same enzyme. The heterogeneity could be caused by the autoproteolytic effects of the enzyme. Such effects are known to occur with e-chymotrypsin, giving rise to several forms of the same enzyme (7).

### Table I

<table>
<thead>
<tr>
<th>Substrate</th>
<th>Active fraction</th>
<th>I</th>
<th>II</th>
<th>III</th>
<th>IV</th>
<th>V</th>
<th>VI</th>
<th>VII</th>
<th>VIII</th>
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<tr>
<td>Valeryl salicylate X 10^-4</td>
<td>9.9</td>
<td>0.9</td>
<td>1.7</td>
<td>10.4</td>
<td>15.6</td>
<td>14.9</td>
<td>13.9</td>
<td>13.4</td>
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</tr>
<tr>
<td>N-Acetyl-L-tyrosine ethyl ester X 10^-4</td>
<td>3.6</td>
<td>0.8</td>
<td>4.5</td>
<td>5.0</td>
<td>5.4</td>
<td>4.7</td>
<td>5.4</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Casein X 10^8</td>
<td>13.7</td>
<td>2.6</td>
<td>13.3</td>
<td>15.8</td>
<td>17.5</td>
<td>15.8</td>
<td>19.0</td>
<td></td>
<td></td>
</tr>
<tr>
<td>% activity recovered</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Valeryl salicylate</td>
<td>100</td>
<td>0</td>
<td>10</td>
<td>10</td>
<td>8</td>
<td>32</td>
<td>7</td>
<td>11</td>
<td></td>
</tr>
<tr>
<td>N-Acetyl-L-tyrosine ethyl ester</td>
<td>100</td>
<td>0</td>
<td>12</td>
<td>34</td>
<td>31</td>
<td>7</td>
<td>12</td>
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<tr>
<td>Casein</td>
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<td>10</td>
<td>28</td>
<td>27</td>
<td>6</td>
<td>10</td>
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* Expressed as moles hydrolyzed per liter per 30 minutes per mg of N per ml.
† Expressed according to Kunitz, per µg of N.

![Fig. 2](image-url) Crystals of the esterase after three crystallizations. A, crystals from active fraction from preparative column. Magnification, 70 X. B, crystals obtained after a chromatography of the same active fraction (Fractions V and VI, see Fig. 1). Magnification, 42 X.
In Table II the recovery and purification data of a fractionation on the preparative column are shown, beginning with the homogenate of pancreas and terminating with the preparation which had been crystallized three times. The results are typical and consistent with those of a large number of fractionation experiments.

Table III shows the specific activity of the active fraction, and of the enzyme after three crystallizations, toward the four substrates used throughout this investigation. The data indicate clearly that the specific activity after crystallization increases proportionately for each of the substrates. These results support the conclusions derived from the data in the rechromatography experiment described above; namely, that the activity toward these substrates is associated with the same enzyme protein.

Fig. 4 shows the electrophoretic behavior of a preparation of the esterase which had been crystallized three times, in free boundary and in starch gel electrophoresis. Both methods of electrophoresis indicate a homogeneous preparation. The low isoelectric point, 4.1 to 4.2, estimated from a plot of pH against mobility (Fig. 5) agrees well with the elution behavior of the enzyme on the DEAE-cellulose column.

The isoelectric point of the esterase is well below that of the known proteolytic enzymes. Trypsin, α-chymotrypsin, and chymotrypsin B, all from bovine pancreas, have isoelectric points of 10.8 (8), 8.3 (9), and 4.7 (9), respectively. Elastase from porcine pancreas has an isoelectric point of 9.5 (10, 11). Starch gel electrophoresis of trypsin and α-chymotrypsin at pH 8.4, with or without previous incubation with diisopropyl fluorophosphate, showed trypsin to be cationic and α-chymotrypsin to be essentially neutral. Chymotrypsin B from bovine pancreas, with a known isoelectric point of 4.7 (9), was studied in detail and compared with the esterase enzyme. Mixtures of the two enzymes subjected to electrophoresis at pH 8.5 yielded two well separated protein zones, with the esterase having the greater mobility. Electrophoresis after incubation with diisopropyl fluorophosphate yielded the same results.
Fig. 6 shows the specific activity of the esterase, α-chymotrypsin, and chymotrypsin B toward the substrate, valeryl salicylate. The data for chymotrypsin B were obtained with crystalline chymotrypsin B and crystalline chymotrypsinogen B. The latter was activated as described by Laskowski. The protein concentrations of chymotrypsin B and chymotrypsinogen B were determined from optical density data in conjunction with conversion factors recorded in the literature. Although both α-chymotrypsin and chymotrypsin B show some activity, it is of such a low order of magnitude that the $K_m$ could not be calculated with any degree of certainty from the Lineweaver-Burk plot. $K_m$ for the esterase, at pH 8.63 in 5% ethanol and at 37°, was calculated to be $2.7 \times 10^{-3}$ (Table IV).

N-Acetyl-L-tyrosine ethyl ester, a substrate which has been used extensively in kinetic studies of the chymotrypsins, is hydrolyzed by the esterase. Table IV shows the $K_m$ values of α-chymotrypsin and the esterase obtained with this substrate using the pH-Stat method. Experiments at different concentrations of substrate and enzymes have shown α-chymotrypsin to have a specific activity 4 to 5 times that of the esterase.

The esterase has proteolytic activity as determined by the Kunitz method with casein as the substrate. The proteolytic activity of the esterase toward casein is less than that of α-chymotrypsin (Table IV).

An interesting feature of the enzyme is its ability to hydrolyze rapidly poly-L-glutamic acid. This is in marked contrast to α-chymotrypsin and trypsin, both from bovine pancreas, which do not attack this substrate to any significant extent. The degradation products are peptides and no detectable amount of free glutamic acid is formed. The enzyme carboxypeptidase does hydrolyze poly-L-glutamic acid; however, the only product formed is glutamic acid. Work with crude pancreatic extracts has suggested the presence of an enzyme other than carboxypeptidase which does hydrolyze poly-L-glutamic acid. The pH optimum for the hydrolysis of poly-L-glutamic acid by the esterase is close to 5.3, which is well below the optimum for the hydrolysis of acetyl-L-tyrosine ethyl ester, pH 7.5, and valeryl salicylate, pH 8 to 9. A number of other poly-L-amino acids were studied as substrates for the enzyme; in all cases the hydrolysis of N-acetyl-L-tyrosine ethyl ester and casein. Its activity toward each of these substrates is less than that of α-chymotrypsin.

The enzyme catalyzes a rapid hydrolysis of poly-L-glutamic acid.
acid at pH 5.3, a substrate that is not attacked significantly by trypsin or α-chymotrypsin.

REFERENCES
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