Flavin Mononucleotide: The Coenzyme of Reduced Diphosphopyridine Nucleotide Dehydrogenase*

N. Appaji Rao, S. P. Felton, F. M. Huennekens, and Bruce Mackler†

From the Departments of Biochemistry and Pediatrics, University of Washington, Seattle, Washington

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From the particulate reduced nicotinamide adenine dinucleotide (DPNH) oxidase of beef heart, Mackler (3) has obtained a soluble DPNH dehydrogenase which was estimated to be approximately 90 to 95% homogenous when examined electrophoretically. Similarly, when the enzyme was chromatographed on diethylaminoethyl cellulose with phosphate buffer, pH 7.5, in a gradient elution method (0.001 M to 0.01 M), only a single, symmetrical peak with respect to protein and flavin was observed in the elution profile (4). The enzyme contains bound flavin and nonheme iron in the ratio of 1:2. The minimal molar concentration of solubilization has been used. One of the principal disadvantages (molecular weight, pH optimum, turnover number, etc.) of the enzyme as described previously (5-11), it is difficult to compare the various DPNH dehydrogenases since, even when prepared from the same tissue, different methods of solubilization have been used. One of the principal discrepancies between the various DPNH dehydrogenases is the nature of the flavin component. It has been reported that only FAD is found in a soluble DPNH dehydrogenase (12). Earlier, Mahler et al. (5) had reported that the prosthetic group of the soluble DPNH-cytochrome c reductase from pig heart is a flavin dinucleotide not identical with FAD. On the other hand, we have reported in preliminary communications (1, 2) that FMN is the flavin component of the soluble DPNH dehydrogenase prepared from DPNH oxidase, and King (3) has reached a similar conclusion with a DPNH dehydrogenase prepared by a different procedure from beef heart (8).

The present paper extends our observations in support of the contention that FMN is the prosthetic group of the beef heart DPNH dehydrogenase. Several analytical techniques have been used to identify and quantitatively determine the flavin released from the enzyme by treatment with heat and acid. No evidence could be obtained to support the possibility that the enzyme contains bound FAD, which is broken down to FAD upon detachment from the protein. The enzyme has been resolved into an apoenzyme and FMN, and activity is restored to the apoenzyme by the addition of FMN, but not of FAD. Finally, various particulate precursors of the soluble DPNH dehydrogenase have been assayed for FAD and FMN and it has been shown that, during purification of the dehydrogenase, there is a progressive enrichment of FMN in the enzyme preparation.

EXPERIMENTAL PROCEDURE

Materials—Chemicals and enzymes were obtained from the following commercial sources: DPN, TPN, DPNH, TPNH, and nucleotide pyrophosphatase (Aspergillus oryzae) from California Corporation for Biochemical Research; riboflavin, FMN, FAD, lactic dehydrogenase, alkaline phosphatase, and Trypsin-300 from Sigma Chemical Company; DEAE-cellulose from Schleicher and Schuell Company; AMP from Nutritional Biochemicals Corporation; 1,2-naphthoquinone-4-sulfonate from Eastman Organic Chemicals; Florisil from Floridian Company; catalase from Worthington Biochemical Corporation; pig kidney acetone powder from Biobin Laboratories. Brewers’ yeast was kindly supplied by the Rainier Brewing Company. Nagarse mitochondrial (13), particulate DPNH-cytochrome Q reductase, and particulate DPNH-cytochrome c reductase were kindly supplied by Drs. Y. Hatefi and D. E. Green.

Methods—D-Amino acid oxidase and TPNH-cytochrome c reductase were prepared from pig kidney and brewers’ yeast, respectively, by the methods of Huennekens and Felton (14). For the assay of FAD by the D-amino acid oxidase, our previously described procedure (14) was adapted for spectrophotometry by coupling the reaction product, pyruvate, with excess lactic dehydrogenase according to the method of DeLuca, Weber, and Kaplan (15). The complete system contained in a cuvette of 1 cm light path: 0.4 μmole of DPNH, 100 μmoles of sodium phosphate buffer, pH 7.5, 0.1 mg of crystalline lactic dehydrogenase, 150 units of crystalline catalase, sample containing 0.1 to 1.0 μg of FAD, 0.1 ml of D-amino acid oxidase, and water to make 3.0 ml. The optical blank was identical except for the omission of DPNH. One hundred micromoles of DL-

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The abbreviation used is: FMN, flavin mononucleotide (riboflavin 5′-phosphate).

2 T. E. King, private communication.
alanine were added to each cuvette in order to start the reaction and the change in absorbancy at 340 mμ was recorded over a 5-minute period. FMN was measured by the reactivation of the TPNH-cytochrome c reductase as described previously (14). These enzymatic assays are very specific for FAD and FMN, and no inhibition is observed when the inactive nucleotide (FMN in the FAD assay, or FAD in the FMN assay) is present in a 10-fold excess. Microbiological assays for riboflavin with Lactobacillus casei were carried out by the Wisconsin Alumni Research Foundation. Total flavin in a sample was determined by the reduction in absorbancy at 490 mμ upon the addition of sodium hydrosulfitc; a value of Δε = 11.3 × 10³ M⁻¹ cm⁻¹ per mole was used for the difference in extinction coefficient between the oxidized and reduced flavin (16). AMP was measured enzymatically with adenylic deaminase according to the method of Nikiforuk and Colowick (17). Adenine-containing compounds were detected by a chemical method in which the sample was hydrolyzed for 6 hours with 6 N HCl in a sealed tube, and the degradation product, 4-amino-5-imidazole carboxamide, was chromatographed on paper (RF = 0.69 in n-butanol-acetic acid-water, 4:1:5) and made visible by spraying with Folin’s reagent (18).

The reaction mixture for the hydrolysis of flavin coenzymes by nucleotide pyrophosphatase contained 100 mμmoles of flavin, 100 mμmoles of sodium phosphate buffer, pH 8.3, and 6 μg of enzyme in a total volume of 1.0 ml. After incubation of the mixture at 30° for 10 minutes, the fluorescence of a suitable diluted aliquot was measured in a Farrand spectrofluorometer with the appropriate riboflavin filters.

The hydrolysis of flavins by alkaline phosphatase was carried out in the following reaction mixture: 100 mμmoles of Tris buffer, pH 8.5, 3.0 mμg of alkaline phosphatase, and 1.0 mμg of flavin in a total volume of 1.0 ml. The reaction mixtures were incubated at 30° and aliquotes were withdrawn at various time intervals and assayed for residual FMN with the TPNH-cytochrome c reductase system.

Isolation and Assay of DPNH Dehydrogenase. The enzyme was solubilized by incubating the DPNH oxidase at 44° for 15 minutes in the presence of 9% ethanol, as described earlier (3). The enzyme was lyophilized to remove ethanol, taken up in water, and fractionated with saturated ammonium sulfate, pH 7.0; the 0 to 60% fraction was used in all experiments reported in this paper. DPNH oxidizing activity with cytochrome c and dichlorophenolindophenol were measured spectrophotometrically, as described previously (3). For assay with ferricyanide as the acceptor, the cuvettes contained: 400 mμmoles of potassium buffer, pH 7.5, 1.3 mμmoles of sodium ferricyanide, 10 mμmoles of sodium azide, and 0.8 mμg of DPNH. The blank was identical except for the omission of ferricyanide. The nonenzymatic reduction of ferricyanide was observed by the change in absorbancy at 410 mμ for 1 minute after the addition of DPNH, whereupon the enzyme was added and the reaction rate followed at 15-second intervals.

Release of Enzyme-bound Flavin—The flavin of the DPNH dehydrogenase was completely released by heating the enzyme, approximately 6 mg per ml, in a boiling water bath for 6 minutes, followed by cooling to 5° and acidification with perchloric acid to a final concentration of 10%. After centrifugation, the clear yellow supernatant fluid was adjusted to pH 7.5 to 8.0 with 6 N KOH and potassium perchlorate was removed by centrifugation. Alternatively, particulate preparations, 50 mμg of protein, were incubated with 10 mμg of trypsin at 37° for 90 minutes (19) before the above heat and acid treatment.

Reversible Resolution of Enzyme—A rapid resolution of the enzyme into apoenzyme and flavin was achieved by passing 2.0 ml of the preparation, 10 mg of protein per ml, through a 1-× 7 cm Florisil column in the cold. The flavin was retained at the top of the column, and after the column had been washed with 3.0 ml of water, the pale yellow, protein-containing eluate was recycled through the same column to yield approximately 7.0 ml of a colorless eluate representing the DPNH apodehydrogenase, 2.5 mg of protein per ml.

Column Chromatography of Flavins—Commercial DEAE-cellulose was washed repeatedly with 5.0 m K₂HPO₄, pH 8.5, until a clear supernatant fluid was obtained after the suspension had settled. The material was then washed with water until the filtrate had a pH of approximately 7.0 and showed essentially no absorbancy at 260 mμ. A thin slurry of purified DEAE-cellulose was packed under gravity into a 2-× 15-cm glass column to a height of 14 cm and washed with 500 ml of water. The flavin, approximately 0.1 to 10.0 mg in 1 to 2 ml, was added carefully to the top of the column, and water was used initially to elute riboflavin. The solvent was then changed to 0.05 m K₂HPO₄, pH 5.5, whereupon FMN and FAD were eluted consecutively.

Paper Chromatography of Flavins—Descending paper chromatography of flavins was performed on 28- X 42-cm sheets of Whatman No. 1 or No. 3 paper, with 5% Na₂HPO₄, n-butanol-acetic acid-water (4:1:5, volume for volume), or isobutyric acid-ammonia-water (66:1:33) as the solvent systems (20). In addition, circular paper chromatography was also used (21). The latter method involves application of the flavins to the circumference of a small circle with a radius of approximately 2 cm, the center of which coincides with the center of a circular filter paper 24 cm in diameter.

RESULTS AND DISCUSSION

Identification of Dehydrogenase Flavin as FMN

Total Flavin—The flavin of the DPNH dehydrogenase was completely released by heating the enzyme at 100° for 6 minutes followed by acidification with perchloric acid to a final concentration of 10%. No additional flavin was released when the heat- and acid-denatured protein was subjected to tryptic digestion. The flavin content of both the intact dehydrogenase and the supernatant fluid obtained from the denatured enzyme was measured by means of a microbiological assay with L. casei and also by the change in absorbancy at 450 mμ after treatment with hydrosulfite. Data presented earlier (1, 2) demonstrated an average content of 9.8 mμmoles of flavin per mg of protein. Comparably, 10 mμmoles of flavin per mg of protein were found in the DPNH-cytochrome c reductase from pig heart (16), whereas when the DPNH dehydrogenase is obtained by treatment of beef heart electron transport particle with snake venom (10), only 1 mμg of flavin per mg of protein is present (22). Thus, the latter preparation may actually be an apodehydrogenase with respect to flavin. Further evidence on this point will be discussed in a subsequent section of this paper.

Enzymatic Assay—Assay systems which respond specifically to FMN and FAD revealed that essentially all of the flavin released from the DPNH dehydrogenase by the above treatment could be accounted for as FMN (see Huennecens et al. (1),
only FMN was detected in all instances by the specific enzymatic activity. The dehydrogenase flavin, when admixed with FMN before chromatography, migrated with an RF value which more nearly resembled that of FAD but was not treated with acid. Both samples were subjected to circular paper chromatography with the n-butanol-acetic acid-water solvent system. From the data in Fig. 1, it is evident that shorter periods, 30 seconds to 2.5 minutes, of heat denaturation unaccompanied by acid treatment resulted in the flavin migrating with an RF value which more nearly resembled that of FAD than FMN. Even when the acid treatment was included with samples heated for less than 2 minutes, an RF value intermediate between those of FAD and FMN was obtained. With increasing periods of heat denaturation, however, the RF value increased progressively and reached that of FMN. This anomalous chromatographic behavior could be misleading in elucidating the identity of the dehydrogenase flavin, but when the above samples were analyzed before chromatography, or when the flavin spots were eluted from the paper and analyzed similarly, only FMN was detected in all instances by the specific enzymatic assays. Although the reason for the anomalous migration is not yet clear, it is possible that incomplete denaturation of the protein may hinder the movement of the flavin.

**Table I.** A small amount of FAD, approximately 5% of the total, was encountered occasionally in dehydrogenase preparations, but never when the enzyme had been fractionated with ammonium sulfate after solubilization. It should be noted that when FAD was admixed with the intact enzyme before the heat and acid denaturation, it was recovered quantitatively as judged by enzymatic assay. This experiment demonstrated that external FAD, at least, is not broken down under conditions which liberate the enzyme-bound flavin.

Paper Chromatography—When examined by paper chromatography with three different solvent systems, the dehydrogenase flavin migrated with the same RF value as FMN (Table I). No separation was observed when the dehydrogenase flavin was admixed with FMN before chromatography.

In order to investigate whether the migration of the flavin on paper chromatograms could be affected by the prior denaturation treatment, aliquots of the enzyme were exposed to various periods of heating, with and without subsequent acid treatment. A solution of the enzyme was placed in a boiling water bath and two aliquots were withdrawn at each of the following time intervals: 30 seconds, and 1, 2, 4, and 6 minutes. One of the two samples was treated with perchloric acid and the precipitated protein was removed by centrifugation. The second heat-denatured sample was centrifuged to remove denatured protein. Flavin from 90 mg of DPNH dehydrogenase was chromographed as described in "Experimental Procedure." In the elution profile the dehydrogenase flavin emerged as a single, symmetrical peak indistinguishable in its position from FMN (cf. Fig. 2). Admixing FMN with the dehydrogenase flavin before chromatography did not result in separation of the two substances.

**Fluorescence Studies**—Since the molar fluorescence of FAD at pH 7.0 is only approximately 10 to 15% as large as that of FMN (23), fluorescence measurements can be used to obtain additional evidence on the nature of the dehydrogenase flavin. Samples of FMN, FAD, and the dehydrogenase flavin were incubated with a purified nucleotide pyrophosphatase from *Aspergillus oryzae*. The fluorescence of each solution was measured before the addition of enzyme and again after incubation for 10 minutes at 30°. The fluorescence of the FAD sample increased approximately 45%.

![Fig. 1. Effect of denaturation process on RF of released flavin](http://www.jbc.org/)

![Fig. 2. Chromatography of dehydrogenase flavin on DEAE-cellulose. Flavin from 90 mg of DPNH dehydrogenase was chromatographed as described in “Experimental Procedure.” Aliquots, 2.5 ml, were collected in each tube. The positions of riboflavin, FMN, and FAD are shown in the elution profile.](http://www.jbc.org/)
TABLE II
Rate of degradation of FMN and dehydrogenase flavin by alkaline phosphatase

<table>
<thead>
<tr>
<th>Time of incubation</th>
<th>Hydrolysis</th>
</tr>
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<tbody>
<tr>
<td></td>
<td>Dehydrogenase flavin</td>
</tr>
<tr>
<td>min</td>
<td>%</td>
</tr>
<tr>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>5</td>
<td>25</td>
</tr>
<tr>
<td>15</td>
<td>55</td>
</tr>
<tr>
<td>30</td>
<td>75</td>
</tr>
<tr>
<td>60</td>
<td>100</td>
</tr>
</tbody>
</table>

10-fold after this treatment owing to the production of FMN, whereas that of FMN and the dehydrogenase flavin each remained essentially constant.

Degradation Studies—Equimolar quantities of FMN and the dehydrogenase flavin were incubated with alkaline phosphatase at 30°. Aliquots were withdrawn at 0, 5, 15, 30, and 60 minutes and the amount of FMN was estimated by the TPNH-cytochrome c aporeductase assay system. As shown in Table II, both samples were dephosphorylated at essentially the same rate. When the aliquots of these reaction mixtures were subjected to paper chromatography in three solvent systems, only FMN and riboflavin were found.

Possibility that Enzyme-bound Flavin is FAD—The above results provide strong evidence that when the DPNH dehydrogenase is denatured by heat, by acid, or by a combination of these procedures, the original amount of enzyme-bound flavin is accounted for quantitatively as FMN in the supernatant fraction. Furthermore, when the flavin is removed by other techniques, including dialysis or treatment with Florisil (see next section), only FMN is found. Even when the flavin is bound to the protein, it can be shown to be FMN by assay with the yeast TPNH-cytochrome c aporeductase. In the experiment described in Fig. 3, all of the components of the FMN assay system were present initially except for FMN. Addition of 0.2 µg of FMN, Curve A, or two levels of DPNH dehydrogenase containing 0.25 and 0.5 µg of flavin, respectively, Curves B and C, caused the reduction of cytochrome c. No activity was observed, however, with the DPNH dehydrogenase samples when TPNH or the TPNH-cytochrome c reductase was omitted. Separate experiments, in which flavin binding is measured fluorometrically (4), have established that the flavin does not dissociate from the protein under the conditions of this experiment. Thus, reactivation of the TPNH-cytochrome c aporeductase must be caused by the bound flavin, obviously FMN. This experiment is similar to the demonstration by Mahler (24) that DPN, bound to rabbit muscle 3-phosphoglyceraledehyde dehydrogenase, can be utilized as a substrate by the DPNH-cytochrome c reductase.

These observations, leading to the conclusion that FMN is the prosthetic group of the beef heart DPNH dehydrogenase, are further strengthened by finding, to be described below, that the apodehydrogenase is reactivated almost completely by FMN, but not by FAD. King has also concluded that his DPNH dehydrogenase preparation from beef heart contains FMN as its prosthetic group. Finally, it would not be unexpected to find FMN involved with a mammalian DPNH dehydrogenase since it is well established that the classical TPNH-cytochrome c reductase (TPNH dehydrogenase) from yeast has FMN for its prosthetic group (25).

Despite the weight of this evidence, the possibility must be considered that the enzyme-bound flavin is actually FAD, or an FAD-like dinucleotide, and that removal of the flavin from the protein by any method results in its breakdown to FMN. In support of this hypothesis are the observations of Mahler et al. (5) and Singer et al. (7, 9), who have reported finding approximately equimolar quantities of flavin and adenine in other

![Fig. 3. Enzymatic estimation of FMN bound to DPNH dehydrogenase. Standard TPNH-cytochrome c reductase assay as described in "Experimental Procedure." In Experiment A, 0.2 µg of FMN was added at the time indicated by the arrow; in Experiments B and C, DPNH dehydrogenase samples containing 0.5 and 0.25 µg of FMN, respectively, were added at the arrow.](http://www.jbc.org/)
Reconstitution of Dissociated DPNH Apodehydrogenase with FMN

As a consequence of the above findings, which demonstrated that the dehydrogenase contains only FMN, attempts were made to dissociate the bound flavin from the enzyme by relatively mild procedures and thereafter to reactivate the resulting apoenzyme with added FMN. It was found that dialysis of the enzyme for 24 hours against water resulted in a progressive loss of flavin which paralleled the loss of DPNH dehydrogenase activity with dichlorophenolindophenol or cytochrome c as acceptors (Fig. 4). The activity was not restored, however, by the addition of either FMN or FAD to the apoenzyme. When the enzyme was dialyzed for shorter time intervals (less than 12 hours), resulting in only a partial loss of flavin and activity, there was considerable stimulation of cytochrome c reductase activity upon the addition of FMN, but not FAD. Activity with dichlorophenolindophenol, on the other hand, could not be restored by addition of either flavin.

A rapid and more satisfactory method for the removal of flavin consisted of passing the enzyme through a small Florisil column. The flavin was adsorbed at the top of the column while the protein emerged in the effluent. Repetition of this procedure gave rise to a suitably resolved apoenzyme, recovered in about 80% yield as judged by protein analysis, and from which about 90% of the flavin had been removed.

When the apoenzyme, prepared by the above method, was tested in the presence of added FMN, approximately 75% of the original activity with cytochrome c as acceptor was restored (cf. Huennekens et al. (1), Fig. 1); the apoenzyme was not reactivated by FAD or riboflavin. This demonstration of the restoration of activity of the apoenzyme with FMN further establishes the identity of the prosthetic group of DPNH dehydrogenase.

The Michaelis constant of $2.3 \times 10^{-4}$ for the reactivation of the apoenzyme by FMN is relatively large for flavoproteins, and provides an explanation for the facile dissociation of the coenzyme from the protein during dialysis. It should be noted that approximately 15 to 20 times as much FMN as was present originally in the holoenzyme must be added to fully reconstitute the activity of the apoenzyme.

The activity of the enzyme with dichlorophenolindophenol as acceptor drops from 40 to 3 μmoles of DPNH oxidized per minute per mg of protein upon removal of the flavin, and in contrast to the results with cytochrome c, this activity is not restored upon the addition of FMN or FAD. On the other hand, the activity with ferricyanide as acceptor is not appreciably diminished when the flavin is removed from the enzyme and, hence, no increase in activity is observed upon the addition of FMN or FAD. These results are summarized in Table III. The finding that DPNH oxidation coupled with ferricyanide is apparently not dependent upon the presence of bound flavin provides further evidence that the DPNH dehydrogenase of Ringler, Minakami, and Singer (7) could be an apodehydrogenase with respect to flavin since these investigators have routinely used a ferricyanide assay.

* Activity is expressed as micromoles of DPNH oxidized per minute per mg of protein.

![Fig. 4. Effect of dialysis on flavin content and enzyme activity](http://www.jbc.org/)

### Table III

<table>
<thead>
<tr>
<th>Electron acceptor</th>
<th>Activity*</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Holoenzyme</td>
</tr>
<tr>
<td>Cytochrome c</td>
<td>3.9</td>
</tr>
<tr>
<td>Dichlorophenolindophenol</td>
<td>40</td>
</tr>
<tr>
<td>Ferricyanide</td>
<td>38</td>
</tr>
</tbody>
</table>

DPNH dehydrogenase preparations. An attempt was made, therefore, to detect AMP or other adenine-containing fragments in the present DPNH dehydrogenase, especially after release of FMN had occurred. With adenyllic deaminase as the assay system (17), no AMP could be detected in the native or denatured enzyme, or in the flavin extracts. Alternatively, a chemical method (18) was used which responds to adenine, adenosine, or AMP. Samples of the intact enzyme, the dehydrogenase flavin, FMN, and FAD were hydrolyzed with 6 N HCl in a sealed tube at 100° and 4-amino-5-imidazole carboxamide was estimated with the Folin reagent. A positive reaction was obtained only with FAD. These results appear to exclude the possibility that FMN originates from cleavage of bound FAD during the denaturation process.

Reconstitution of Dissociated DPNH Apodehydrogenase with FMN

The high molecular weight enzyme (~20 = 12.6) with the following properties: an FMN to iron ratio of 5.6 with the following properties: an FMN to iron ratio of 2 or 4:1, reversible resolution and reactivation with FMN, and ability to use cytochrome c as acceptor.
Flavin Content of Various Particulate Precursors of DPNH Dehydrogenase

The identification of FMN as the DPNH dehydrogenase flavin has made it of interest to determine the FMN and FAD content of various particulate preparations which are precursors of, or related to, the soluble dehydrogenase. Of the following preparations which were assayed, all but the last two were particulate: beef heart homogenate, beef heart mitochondria, Nagarse mitochondria, electron transport particle, DPNH oxidase, DPNH-cytochrome c reductase (Amytal- and antimycin-sensitive), DPNH-coenzyme Q reductase, DPNH-cytochrome c reductase from pig heart (3), and succinic dehydrogenase from electron transfer particle (26). Each sample was digested with trypsin, followed by denaturation with heat and acid to release the total flavin, which was determined by the change in absorbancy at 450 mg upon addition of hydrosulfite. The same treatment, but

**TABLE IV**

Recovery of flavin and enzyme activity during solubilization of DPNH dehydrogenase

<table>
<thead>
<tr>
<th>Preparation</th>
<th>DPNH oxidase</th>
<th>DPNH dehydrogenase</th>
<th>Recovery</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Protein</td>
<td>Activity</td>
<td>FMN</td>
</tr>
<tr>
<td>I</td>
<td>mg</td>
<td>units</td>
<td>mg</td>
</tr>
<tr>
<td>II</td>
<td>5,600</td>
<td>1620</td>
<td>567</td>
</tr>
<tr>
<td>III</td>
<td>7,000</td>
<td>2240</td>
<td>840</td>
</tr>
<tr>
<td>Average</td>
<td>12,000</td>
<td>3840</td>
<td>1350</td>
</tr>
</tbody>
</table>

*Flavin content is expressed as millimicromoles per mg of protein.
† - Value not determined.

**TABLE V**

Flavin content* of various particulate precursors of DPNH dehydrogenase and other related preparations

<table>
<thead>
<tr>
<th>Preparation</th>
<th>Trypsin, heat, and acid</th>
<th>Heat and acid</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Total flavin</td>
<td>FAD</td>
</tr>
<tr>
<td>Homogenate</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Mitochondria</td>
<td>0.20</td>
<td>0.10</td>
</tr>
<tr>
<td>Nagarse mitochondria</td>
<td>0.44</td>
<td>0.3</td>
</tr>
<tr>
<td>DPNH oxidase</td>
<td>0.57</td>
<td>0.08</td>
</tr>
<tr>
<td>Electron transport particle</td>
<td>0.46</td>
<td>0.17</td>
</tr>
<tr>
<td>DPNH-cytochrome c reductase</td>
<td>0.84</td>
<td>0.03</td>
</tr>
<tr>
<td>DPNH-Q reductase</td>
<td>1.4</td>
<td>0.03</td>
</tr>
<tr>
<td>DPNH dehydrogenase</td>
<td>8.2</td>
<td>0.3</td>
</tr>
<tr>
<td>from DPNH oxidase</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>DPNH-cytochrome c reductase from pig heart</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Succinic dehydrogenase</td>
<td>-</td>
<td>-</td>
</tr>
</tbody>
</table>

*Flavin content is expressed as millimicromoles per mg of protein.
† - Value not determined.

It contains progressively larger amounts of FMN relative to FAD, and this is paralleled by the increase in DPNH dehydrogenase activity. Finally, all of the bound flavin is FMN in the highly purified, soluble DPNH dehydrogenase which presumably has no other enzymatic activities.

It is of interest to calculate the recovery of flavin and enzymatic activity (DPNH oxidation linked to cytochrome c or dichlorophenolindophenol) during the solubilization of the dehydrogenase from the DPNH oxidase (Table V). In three experiments, an average of 86% of the activity was recovered with a retention of 63% of the flavin. Since the amount of FMN in the dehydrogenase did not exceed in any instance the amount originally present in the DPNH oxidase, further evidence is thereby provided that FAD is not broken down during solubilization of the enzyme. The lower recovery of flavin, relative to the recovery of activity, may be due to the fact that all of the FMN in the particulate DPNH oxidase is not associated with the DPNH dehydrogenase.

**SUMMARY**

1. The flavin component of the soluble reduced nicotinamide adenine dinucleotide dehydrogenase from beef heart was shown to be flavin mononucleotide (FMN), as judged by enzymatic, chromatographic, fluorometric, and degradative methods.
2. The enzyme was resolved into an aporeductase and FMN.
by passage through a Florisil column; FMN, but not flavin adenine dinucleotide (FAD), reactivated the aporeductase \( (K_m = 2.3 \times 10^{-4} \text{ M}) \) with cytochrome c as the terminal electron acceptor. The aporeductase retained almost all of its activity with ferricyanide as the acceptor and FMN or FAD did not stimulate this system.

3. The possibility that enzyme-bound FAD was degraded to FMN upon release from the enzyme was invalidated by the inability to find adenosine 5'-phosphate after dissociation of the enzyme.

4. FMN was estimated quantitatively in various particulate precursors of the reduced nicotinamide adenine dinucleotide dehydrogenase, and was shown to be concentrated, relative to FAD, during the course of purification of the enzyme.

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REFERENCES

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