Potassium Ion-dependent Hydrolysis of Adenosine Triphosphate Induced by Nigericin in Mitochondria*

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SUMMARY

K+ is required for the stimulation by the antibiotic nigericin of adenosine triphosphate hydrolysis in rat liver mitochondria. This action does not occur when K+ is replaced by other monovalent cations. It requires slightly hypotonic conditions and is not related to the swelling phenomena linked to the accumulation of K+ in mitochondria. ATPase is inhibited by small anion molecules to which the mitochondrial membrane is impermeable, less so if the anion is translocated inside the mitochondria. High concentrations of Rb+ or Na+ and low concentrations of Ca++ inhibit the ATPase stimulated by nigericin. The ATPase activity of submitochondrial sonic fragments, and of Racker's coupling factor F1, are not stimulated by nigericin.

β-Hydroxybutyrate and succinate partially inhibit the hydrolysis of ATP stimulated by nigericin in K+ media. Neither glutamate, malate, pyruvate, nor α-ketoglutarate inhibits this enzymatic activity. The inhibition of ATPase by oxidizable substrates is ascribed to the reincorporation of the liberated inorganic orthophosphate into ADP by way of the phosphorylation linked to the oxidation of succinate and β-hydroxybutyrate.

Low concentrations of nigericin partially inhibit the ATPase stimulated by uncoupling agents requiring the presence of alkali metal cations for their mechanism of action; the inhibition depends on the ionic environment of the mitochondria. Nigericin does not affect ATPase activities induced by agents with no alkali metal cation requirement. Dianemycin, an antibiotic with properties similar to nigericin regarding induced ion movement and spectra of respiratory inhibition, does not stimulate the hydrolysis of ATP in any of several media tried.

The K+-dependent ATPase stimulated by nigericin appears not to be related to a translocation of alkali metal cations induced by the antibiotic. The possibility that the induction of this activity is related to the participation of a potassium-dependent step in the sequence of reactions leading to the synthesis of ATP in mitochondria is discussed.

It has recently been shown that a group of compounds that specifically modify the vectorial movements of alkali metal ions in mitochondria also stimulate the hydrolysis of adenosine triphosphate with a strict dependency on the cation environment. Valinomycin (1) induces an increase in mitochondrial ATPase activity requiring K+, Rb+, or Cs+ (2); commercial triamcinolone (3, 4) activates ATPase in the presence of K+ or Rb+, less with Na+, and not with Li+ or Cs+; gramicidin (5) stimulates the hydrolysis of ATP in the presence of K+, Rb+, Cs+, Na+, or Li+; the nonactin homologues (6) are active in the presence of Na+, K+, Rb+, or Cs+, and have little activity with Li+. A common feature of these compounds is their ability to promote the active uptake of alkali metal cations in mitochondria (2) supported by the high energy intermediates—collectively labeled ~W (7)—derived either from ATP or from the electron transfer chain.

The previous findings suggest that the alkali metal cation-dependent stimulation of ATP hydrolysis produced by these compounds occurs because of the energy demands imposed by the transport of ions. However, evidence will be presented in this paper to show that the activation of the alkali metal cation-dependent ATPase from mitochondria is not necessarily related to a predicted change in the pattern of ion movements induced in the mitochondrial membrane.

Recently we reported (8) some properties of the antibiotin nigericin involving specific respiratory inhibition coupled to the loss of alkali metal cations induced by this compound in mitochondria. On the basis of these observations, we suggested that the site of action of this antibiotic must be at the level of the ion-translocating mechanism of the mitochondrial membrane.

The present paper deals with the description of a unique mitochondrial ATPase activity induced, specifically in K+-containing media, by relatively high concentrations of nigericin. This effect on ATPase activity seems to be distinct from the action of nigericin in blocking the mitochondrial ion pump. It may be the result of the interaction of the antibiotic with a mitochondrial component that requires K+ for its participation in the mechanism of oxidative phosphorylation.

EXPERIMENTAL PROCEDURE

Mitochondria were prepared from livers of male rats weighing 150 to 220 g according to the procedure described by Johnson and Lardy (9). The mitochondria were stored in 0.25 M sucrose at 0° and used within 1 hour of preparation.

The ATPase activity was measured by the method of Lardy and Wellman (10); specific conditions are described in the tables.
**FIG. 1.** The effect of the concentration of nigericin on the activity of mitochondrial ATPase in the presence of different alkali metal cations. The reaction mixture contained 6 mM Tris-ATP (pH 7.4), 10 mM triethanolamine (Cl-) (pH 7.4), 50 mM alkali metal cation (Cl-), and mitochondria from 0.06 g of liver in 0.3 ml of 0.25 M sucrose in a final volume of 1.0 ml at pH 7.4. Nigericin was added in 0.01 ml of a 25% solution of N,N-dimethylformamide in 96% ethanol. In this and the following figures, $M_w$ on the ordinate denotes mitochondria washed with 0.25 M sucrose.

**FIG. 2.** The dependency on K+ of the ATPase activity stimulated by nigericin. Experimental conditions were the same as in Fig. 1, except for the varying concentrations of the salt used to support the increase in enzyme activity induced by the antibiotic. Inorganic phosphate was determined by the method of Sumner (11). The submitochondrial sonic fragments were prepared by the procedure described by Graven, Estrada-O., and Lardy (12). The coupling factor $F_1$ of Pullman et al. (13) was generously provided by Miss Chien Ho-Chiu.

Deionized water and highest purity chemicals were used throughout. The antibiotics used in this work were generous gifts from the following sources: monactin, Professor V. Prelog, Eidgenössischen Technischen Hochschule, Zurich; aurovertin, Drs. H. A. Nash and C. L. Baldwin, Pitman Moore Division, Dow Chemical Company; gramicidin B, Dr. Bernard Witkop, National Institutes of Health; valinomycin, Dr. J. C. McDonald, Saskatoon, Saskatchewan; 1,1,3-tricyano-2-amino-1-propene, Dr. Floyd Eberts, The Upjohn Company; BA 180265, Dr. K. V. Rao, Pfizer Laboratories; and nigericin, Dr. R. Harned, Commercial Solvents Company. The Tris salt of ATP was obtained from Sigma Chemical Company.

**RESULTS**

Nigericin in concentrations from 1 to 10 μg per ml induced the hydrolysis of ATP by mitochondria in media containing K+ (Fig. 1). Maximal stimulation of ATP hydrolysis was induced by nigericin at a concentration of 10 μg per ml; in other experiments, no further enhancement of this activation was observed at higher concentrations of the antibiotic. No significant activation of ATPase was detected when the concentration of nigericin in the medium was below 0.5 μg per ml.

Alkali Metal Cation Requirement of ATPase—The induction of ATPase activity by nigericin was related to the concentration of K+ in the medium (Fig. 2) with maximal activity between 25 and 50 mM. The apparent $K_m$ for K+ was 5 mM. KCl in concentrations above 50 mM inhibited the nigericin-stimulated ATPase, with complete inhibition at 300 mM. Although the potassium ion was responsible for part of the inhibition, as will be seen below, the anion (Cl-) accompanying the monovalent metal cation affected the extent of this inhibition.

In the presence of Rb+, nigericin stimulated slightly the hydrolysis of ATP (Fig. 2). However, whereas K+ has an apparent $K_m$ of 5 mM, Rb+ slightly stimulated the mitochondrial ATPase below 25 mM, and its activation was only 30% of that induced by nigericin in media containing K+. Na+ or NH4+ at concentrations in which K+ (25 to 50 mM) produced a full activation of the enzyme, permitted only a minimal activation of ATPase (0.4 μmole of Pi per 0.2 mg of mitochondrial nitrogen) (Figs. 1 and 2). Li+ and Cs+ did not support nigericin-induced ATPase activity at any concentration tested.

When the concentration of alkali metal cation was kept constant (75 mM), any replacement of K+ with Na+ resulted in a decrease in the ATPase induced by nigericin (Fig. 3). If NH4+, KCl 75 mM

**FIG. 3.** The inability of Na+ to replace K+ on the activity of ATPase stimulated by nigericin. Experimental conditions were the same as in Fig. 1.
Li<sup>+</sup> or Cs<sup>+</sup> was used in place of Na<sup>+</sup>, the results were similar to those presented in Fig. 3. Divalent metal ions did not support the induction of ATPase activity by nigericin.

**pH Dependency of Nigericin-stimulated ATPase**—In mitochondria incubated in a medium buffered with 8 × 10<sup>-2</sup> M histidine, 10<sup>-2</sup> M triethanolamine, and 20 mM acetate, nigericin induced an increase in ATPase activity with an apparent optimal peak at pH 6.3 and a progressive decrease of the enzyme activity as the medium approached pH 8.0 (•, Fig. 4). Substracting the spontaneous, bell-shaped ATPase activity (○) from those values results in a curve with an optimal ATPase activity induced by nigericin at pH 7.0 and above (△). Nigericin induced no ATPase activity when the pH of the medium was less than 6.1.

**Effect of Changes in Osmolarity on ATPase**—Changes in the osmolarity of the media were found to have a marked effect on the ATPase induced by nigericin. When 50 mM KCl was present in the incubation medium, the addition of increasing amounts of sucrose (from 80 to 300 mM) progressively inhibited the ATPase activity induced by nigericin until none was detected at 300 mM sucrose (Table I). It appears that hypotonic conditions are required for the activation of ATPase by this antibiotic.

**Effect of Anions on ATPase**—High concentrations of potassium chloride inhibited the nigericin-activated ATPase as effectively as equivalent molarities of sucrose (Table I; Figs. 2 and 5). Since the mitochondrial membrane is relatively impermeable to Cl<sup>-</sup> ions (14), the inhibition of ATPase was presumed to be a result of the change in osmolality and not an inhibition caused specifically by Cl<sup>-</sup>. To assess this possibility, the effect on nigericin-activated ATPase of increasing concentrations of different anions was studied (Table II). On the basis of the results, anions can be separated into two different groups: (a) those which inhibit the K<sup>+</sup>-dependent ATPase induced by nigericin (chloride, bromide, iodide, nitrate, and citrate) and (b) those which do not inhibit this ATPase activity (acetate, phosphate, and formate). Citrate at 50 μM inhibited ATPase 30% and at 150 μM inhibited completely. This inhibitory effect of citrate does not appear to be due to its chelating activity since EDTA at concentrations up to 60 mM did not inhibit the ATPase activity induced by nigericin. Bromide, nitrate, and iodide were less inhibitory than citrate, and chloride was even less effective.

As is shown in Table II, acetate, phosphate, and formate did not inhibit the ATPase activity appreciably (<30%, even at 150 mM). The anions which failed to inhibit the nigericin-induced ATPase are those to which the mitochondrial membrane is permeable (15). It would appear that the inhibitory effect of anions described above is not the result of simple change in the osmolality of the solution, but is related also to the permeability of the mitochondria to the anions. Support for this proposal comes from the results of the experiments in which l-malate (5 or 7.5 mM) was added to a medium containing increasing amounts of citrate (Table II and Fig. 5). L-Malate facilitates citrate penetration into mitochondria (16, 17). The presence of l-malate decreased the inhibition of nigericin-induced ATPase produced by citrate. Increasing the concentration of malate...
**TABLE II**

*Effect of various anions on K+-dependent ATPase of mitochondria stimulated by nigericin*

<table>
<thead>
<tr>
<th>Anion</th>
<th>P&lt;sub&gt;i&lt;/sub&gt; liberated at anion concentrations of 0.2 mEq</th>
<th>0 mM</th>
<th>10 mM</th>
<th>30 mM</th>
<th>50 mM</th>
<th>75 mM</th>
<th>100 mM</th>
<th>150 mM</th>
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<tr>
<td>Bromide</td>
<td>3.0</td>
<td>3.0</td>
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<td>0.75</td>
<td>0.48</td>
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<td>Nitrate</td>
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<td>3.0</td>
<td>0.65</td>
<td>0.48</td>
<td>0.25</td>
<td>0.04</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Iodide</td>
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<td>3.09</td>
<td>2.82</td>
<td>2.11</td>
<td>1.20</td>
<td>0.36</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Chloride (Tris salt)</td>
<td>2.5</td>
<td>2.42</td>
<td>1.90</td>
<td>1.65</td>
<td>1.25</td>
<td>0.85</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Chloride (NH₄ salt)</td>
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<td>3.1</td>
<td>2.75</td>
<td>2.6</td>
<td>2.5</td>
<td>1.75</td>
<td></td>
<td></td>
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<td>Citrate</td>
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<td>2.28</td>
<td>1.12</td>
<td>0.43</td>
<td>0.06</td>
<td>0.0</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Citrate + l-malate</td>
<td>2.5</td>
<td>2.28</td>
<td>1.15</td>
<td>1.35</td>
<td>1.05</td>
<td>0.85</td>
<td></td>
<td></td>
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<tr>
<td>Acetate</td>
<td>3.0</td>
<td>2.95</td>
<td>2.75</td>
<td>2.60</td>
<td>2.40</td>
<td>2.20</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Formate</td>
<td>3.0</td>
<td>2.30</td>
<td>2.36</td>
<td>2.36</td>
<td>2.23</td>
<td>2.28</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Phosphate*</td>
<td>3.0</td>
<td>3.0</td>
<td>3.0</td>
<td>2.9</td>
<td>2.7</td>
<td>2.4</td>
<td></td>
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</table>

* In the presence of high phosphate, liberated P<sub>i</sub> was determined by using an aliquot of a zero time sample as the blank in the spectrophotometer.

Inhibitory Effect of Rb⁺, Na⁺, and Ca++ on K⁺-dependent ATPase—The inhibition of nigericin-induced ATPase was greater with NaCl and RbCl than with triethanolamine chloride at comparable concentrations. It is very probable that this is the result of a specific inhibitory effect of Rb⁺ and Na⁺ on the K⁺ requirement for the ATPase because neither lithium nor cesium chloride increased the inhibition above that observed with triethanolamine chloride. There was less inhibition with NH₄Cl than with triethanolamine chloride. The replacement of Cl⁻ by acetate decreased the inhibition produced by Na⁺ (Fig. 5) as well as by higher concentrations of potassium ions.

Fig. 6. The effect of substrate oxidation on the net formation of P<sub>i</sub> in the presence of nigericin. Experimental conditions were the same as in Fig. 1. Mitochondria were incubated for 3 min at 30°C with substrates before ATP and nigericin were added. Substrates neutralized to pH 7.4 with triethanolamine were used in this experiment. Rotenone (10⁻⁵ M), added in 0.01 ml of a 75% ethanol-H₂O solution, was used to inhibit β-hydroxybutyrate oxidation.

Lardy, Johnson, and McMurray (18) reported that nigericin (2 µg per ml) induced a striking inhibition of the ATPase induced by 10⁻⁴ M CaCl₂. As will be seen below, CaCl₂ also inhibited close to 70% of ATPase activity induced by nigericin when Ca++ was tested at concentrations of 2.5 × 10⁻³ M.

*Inhibitory Effect of Substrates—*Succinate and β-hydroxybutyrate at 5 mM inhibited the ATPase induced by nigericin 34 and 44%, respectively (Fig. 6). Further increases of succinate or β-hydroxybutyrate concentrations increased the inhibition slightly. However, the addition of both succinate and β-hydroxybutyrate enhanced the inhibition to 68%. Rotenone (10⁻⁵ M) reversed the inhibition of ATPase induced by 10 mM β-hydroxybutyrate (Fig. 6). None of the other substrates—glutamate, malate, pyruvate, or α-ketoglutarate—inhibited the ATPase activity at concentrations between 1 and 40 mM.

**TABLE III**

*Effect of nigericin and uncoupling agents on different ATPase preparations*

The reaction mixtures contained 10 mM triethanolamine-HCl, 50 mM K⁺ acetate, 6 mM Tris-ATP, 200 mM sucrose, and the indicated additions in a total volume of 1 ml; 10-min incubation at 30°C. No sucrose was added for measuring the ATPase induced by nigericin in fresh mitochondria. MgSO₄ (10 mM) was added to the media for measuring ATPase from Racker’s coupling factor F<sub>i</sub> with its coupling factor F<sub>i</sub>. The F<sub>i</sub> preparation had a specific activity of 42 when assayed according to Pullman et al. (13) with an ATP-regenerating system that removes inhibitory ADP.
alkali metal cation requirements for the hydrolysis of ATP induced by those compounds.

Nigericin at a concentration of 2 µg per ml inhibited the ATPase stimulated by valinomycin, nonactin, gramicidin D, and stendomyein A in K⁺ media 14 to 67%; the action of the antibiotic BA 180265A was inhibited 31% (Table IV). As previously mentioned, the uncoupling of oxidative phosphorylation by valinomycin, gramicidin (5), the nonactin homologues (6), and stendomyein A (7), and the induction of ATPase by these agents as well as by BA 180265A (7), is dependent on the presence of alkali metal cations. Lardy et al. (18) also reported that nigericin at 2 µg per ml inhibited 24% of the ATPase induced by

<table>
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<th>Agent</th>
<th>Control</th>
<th>Nigericin, 2 µg per ml</th>
<th>Inhibition</th>
</tr>
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<tbody>
<tr>
<td>None</td>
<td>0.23</td>
<td>0.62</td>
<td></td>
</tr>
<tr>
<td>2-Amino-3,5-diodobenzoate, 10⁻⁴ M</td>
<td>2.23</td>
<td>1.98</td>
<td>11</td>
</tr>
<tr>
<td>1,1,3-Trieyano-2-amino-1-propene, 5 X 10⁻⁴ M</td>
<td>1.60</td>
<td>1.56</td>
<td></td>
</tr>
<tr>
<td>BA 180265A, 1 µg</td>
<td>2.69</td>
<td>1.85</td>
<td>31</td>
</tr>
<tr>
<td>Monactin, 10⁻⁴ M</td>
<td>2.81</td>
<td>1.52</td>
<td>46</td>
</tr>
<tr>
<td>Valinomycin, 10⁻⁴ M</td>
<td>3.31</td>
<td>1.66</td>
<td>53</td>
</tr>
<tr>
<td>Gramicidin B, 10⁻⁴ M</td>
<td>3.32</td>
<td>1.51</td>
<td>57</td>
</tr>
<tr>
<td>Stendomyein A, 22.8 µg</td>
<td>1.99</td>
<td>0.66</td>
<td>67</td>
</tr>
</tbody>
</table>

**Table V**

Effect of different uncoupling agents on ATPase stimulated by nigericin

The reaction mixture contained 6 mM ATP, 50 mM KCl, 25 mM sucrose, 10 mM triethanolamine buffer (pH 7.4), 10 µg of nigericin, and mitochondria from 0.06 g of liver in 0.3 ml of 0.25 M sucrose in a total volume of 1 ml and was incubated for 10 min at 30°. Values represent the average of duplicate experiments which agreed closely.

<table>
<thead>
<tr>
<th>Addition</th>
<th>P₄ liberated</th>
<th>Inhibition</th>
</tr>
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<tbody>
<tr>
<td></td>
<td>μmoles/0.2 mg N</td>
<td>%</td>
</tr>
<tr>
<td>None</td>
<td>3.06</td>
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</tr>
<tr>
<td>Valinomycin, 10⁻⁴ M</td>
<td>1.68</td>
<td>45</td>
</tr>
<tr>
<td>Dinactin, 10⁻⁴ M</td>
<td>1.71</td>
<td>44</td>
</tr>
<tr>
<td>Gramicidin B, 10⁻⁴ M</td>
<td>1.77</td>
<td>42</td>
</tr>
<tr>
<td>CaCl₂, 2.5 X 10⁻³ M</td>
<td>0.97</td>
<td>68</td>
</tr>
<tr>
<td>MgCl₂, 2.5 X 10⁻³ M</td>
<td>2.83</td>
<td>7.5</td>
</tr>
<tr>
<td>1,1,3-Trieyano-2-amino-1-propene, 5 X 10⁻³ M</td>
<td>3.63</td>
<td>0</td>
</tr>
<tr>
<td>2-Amino-3,5-diodobenzoate, 10⁻⁴ M</td>
<td>2.94</td>
<td>3.9</td>
</tr>
<tr>
<td>Oligomycin, 2 µg</td>
<td>0.18</td>
<td>94</td>
</tr>
<tr>
<td>Aurovertin, 2 µg</td>
<td>3.02</td>
<td>0</td>
</tr>
</tbody>
</table>

10⁻⁴ M dinitrophenol, 75% of that induced by 100 µg of deoxycholic acid per ml, and 90% of the enzyme induced by 10⁻⁴ M Ca++. In this regard it is known that a portion of the dinitrophenol-stimulated ATP hydrolysis in liver mitochondria is also dependent on Na⁺ or K⁺ (10, 19) and the ATPase activity induced by deoxycholic acid is greater with K⁺ than with Na⁺ (4). Also, it is known that Ca⁺⁺ interacts in an apparent competition with the mechanism of K⁺ movements in mitochondria (20).

Nigericin (2 µg per ml) did not inhibit the ATPase induced by 5 X 10⁻⁴ M 1,1,3-tricyano-2-amino-1-propene or 2-amino-3,5-diodobenzoate (Table IV). The failure of nigericin to inhibit the ATP hydrolysis stimulated by triiodothyroacetate has been reported previously (18). No ion requirement is known for the hydrolysis of ATP induced by these last three compounds.

It was also found that the compounds with ability to induce hydrolysis of ATP that was partially inhibited by low concentrations of nigericin, inhibited the K⁺-dependent ATPase stimulated by higher concentrations of nigericin. Valinomycin, dinactin, and gramicidin as well as Ca⁺⁺ inhibited the hydrolysis of ATP induced by nigericin 40 to 68%, while trieyanoaminopropene and 2-amino-3,5-diodobenzoate were not inhibitory (Table V). Oligomycin completely inhibited the hydrolysis of ATP induced by nigericin, whereas aurovertin did not inhibit (Table V).

**Discussion**

Nigericin is produced by a streptomyecete isolated originally by Harned et al. from a Nigerian soil (22). It is a powerful inhibitor of the growth of a variety of microorganisms with the striking peculiarity that its lethal effect is reversed by the addition of K⁺ to the assay medium (23).

In the initial experiments with this antibiotic for studies of mitochondria (18), it was found to inhibit the oxidation of most of the DPN-linked substrates, but not of β-hydroxybutyrate and succinate. At low concentrations, nigericin did not uncouple oxidative phosphorylation with any oxidizable substrate but it inhibited the exchange of ³²P between inorganic phosphate and ATP. The same effects were produced in mitochondria by the antibiotic diamycin.

In recent studies (8) it was found that both nigericin and diamycin induced a rapid loss of alkali metal cations previously accumulated in mitochondria under the influence of valinomycin, monactin, or gramicidin B. The inhibition of the oxidation of glutamate and most other DPN-dependent substrates apparently

1 S. N. Graven, S. Estrada-O., and H. A. Lardy, unpublished experiments.
results from the necessity of a high salt concentration for these substrates, since it was reversed by 75 mM alkali metal cations. The oxidation of succinate, β-hydroxybutyrate, and, under some conditions, isocitrate appears to be less dependent on the ionic environment (8).

The present paper shows that nigericin induced a specific K+-dependent ATPase activity in mitochondria. The central question in these studies was whether this K+-dependent ATPase is (a) an essential part of the series of events which lead to the blocking of the uptake, or to the release of intramitochondrial alkali metal cations induced by nigericin, or (b) a collateral effect not directly related to the primary interaction of the antibiotic with the ion-translocating system of the mitochondrial membrane. The latter possibility appears to be the most likely based on the following considerations.

**Optimal Concentration of Nigericin for ATPase and Ion Movements**—Less than 0.3 μg of nigericin per ml (1 to 2 mg of mitochondrial nitrogen per ml) is sufficient to block the uptake of K+ or Na+ with the instantaneous inhibition of the oxidation of glutamate, malate, α-ketoglutarate, citrate, or pyruvate (8). However, no detectable ATPase activity is induced by nigericin below 0.5 μg per ml (0.2 to 0.3 mg of mitochondrial nitrogen per ml); 1 to 10 μg of nigericin per ml are required to induce ATPase (Fig. 1). Increasing the concentration of nigericin from 1 to 10 μg per ml did not change the pattern and efficiency of inhibition of respiration or of cation release from that induced by 0.3 μg per ml.

**Optimal Osmolarity for Induction of ATPase Activity**—The osmolarity required to demonstrate the induction of the K+-dependent ATPase is different from that required to demonstrate the effect of nigericin on ion movements and substrate oxidation in mitochondria. As seen in Table I, hypotonic conditions are required for the stimulation of the K+-dependent ATPase by nigericin. When nigericin was tested in an isotonic medium which was optimal for its effect on substrate oxidation and ion movements, it induced little ATPase activity.

**Correlation between Ion Movements and ATPase Activity**—As described in previous papers (8, 12) the addition of nigericin to mitochondria prior to the addition of valinomycin and monactin prevents the uptake of K+ induced by the latter compounds. Addition of nigericin after introduction of monactin or valinomycin in a medium containing K+, phosphate, Mg++, and an oxidizable substrate causes the ejection of K+. The same pattern of responses is obtained with nigericin when gramicidin B is used to induce the uptake of Na+ or Li+. However, no ATPase activity is induced by nigericin in the presence of Na+ or Li+ (Figs. 1 and 3). The K+ specificity of the ATPase induced by nigericin does not appear to be related to the inhibition of alkali metal cation accumulation by liver mitochondria.

**Effect of Inorganic Phosphate on ATPase and Alkali Metal Cation-Dependent Respiration**—We shall present data elsewhere showing that high concentrations of inorganic orthophosphate overcome the inhibitory effect of nigericin both on the active uptake of phosphate into mitochondria and on the oxidation of the substrates that have an alkali metal cation requirement for oxidation. However, high concentrations of inorganic phosphate do not inhibit the K+-dependent ATPase activity induced by nigericin (see Table II) as would be expected if the phosphate uptake and respiration coupled to the movement of monovalent cations, and the activity of ATPase, were directly related.

**Differences between Nigericin and Dianemycin**—Both nigericin and dianemycin induce the loss of intramitochondrial cations in a system having a rapid turnover of alkali metal cations (8, 12). However, dianemycin did not induce ATPase under a variety of conditions.

These several facts establish that the stimulation of K+-dependent ATPase and the changes in the movements of alkali metal cations linked to the respiratory activity of the mitochondria are independent but possibly related effects of nigericin. The partial inhibition by nigericin of the hydrolysis of ATP stimulated by valinomycin, the gramicidins, the nonactin homologues, etc., could be mediated by its interference with the incorporation of monovalent ions induced by these antibiotics in mitochondria. The ability of nigericin to induce the release of K+ ions was greater than that of dianemycin when valinomycin or the nactins induced the accumulation of K+, while the ability of dianemycin to induce the release of Li+ was greater than that of nigericin when gramicidin B was used to induce Li+ accumulation in the mitochondria (12). In media containing Li+ or Na+ as the sole alkali metal cation, nigericin showed little capacity to inhibit the ATPase induced by gramicidin B, while dianemycin inhibited significantly (Table VI). Thus, nigericin and dianemycin follow a pattern of specificity for Li+ or K+ release similar to that found for their corresponding effects on ATPase stimulated by gramicidin, valinomycin, or the nactins in this paper.

The stimulation of the K+-dependent hydrolysis of ATP induced by nigericin may be the result of a specific interaction of the antibiotic with a mitochondrial component that requires K+ for its participation in the mechanism of oxidative phosphorylation (24-26). Evidence bearing on this point will be published elsewhere.

The indication from Table I that nigericin induces ATPase only under hypotonic conditions requires revision in view of the fact that some electrolytes are not inhibitory. The effect of various anions on the K+-dependent ATPase induced by nigericin indicates the relation of this activity to the selective permeability properties of the mitochondrial membrane. Chloride, bromide, iodide and nitrate, when present at relatively high concentrations, inhibited the ATPase stimulated by nigericin (Ta-
ble I). Among these inhibitory anions are those demonstrated by Chappell and Crofts (14) and Chappell and Haakoff (18) to be excluded from mitochondria. Conversely, phosphate, acetate, and formate, which are known to penetrate mitochondria, are far less inhibitory. Finally, citrate, which does not permeate (27, 28) and is inhibitory (Table I and Fig. 5), is rendered non-inhibitory by the addition of small quantities of l-malate, which permits citrate to penetrate mitochondria (16, 17, 29). That high concentrations of NH₄Cl only slightly inhibited the ATPase induced by nigericin (Table II) may be related to the observation that mitochondria take up NH₄⁺ with NH₃ being the probable penetrating species (5).

Thus the data of Chappell et al. (14, 15) obtained in swelling experiments and our observations on the activity of nigericin-induced ATPase in mitochondria lead to similar classifications of the various anions (Table VII). An association of ATPase activation with mitochondrial swelling has already been noted by Dianzani (30) in mitochondria suspended in hypotonic conditions. In certain cases ATPase activation may be accounted for by an altered permeability of the mitochondria which makes ATPase more accessible to its substrate (31). However, this does not seem to be simply the case for the K⁺-dependent ATPase stimulated by nigericin, because this antibiotic blocks the mitochondrial swelling (8, 12) induced in iso- or hypotonic solutions of KCl with substrate and acetate even in the presence of valinomycin or the nonactin homologues, which swell mitochondria when K⁺ is present (32, 33). It is of interest that mitochondria exposed to hypotonic conditions appear to maintain a specific permeability barrier to small anions. However, this same behavior of the membrane has been observed by others (34).

The apparent inhibition of nigericin induced ATPase by succinate and β-hydroxybutyrate is characterized by a low substrate reincorporation of the liberated Pi into ATP by

<table>
<thead>
<tr>
<th>Nonpenetrating anion (inhibit K⁺ ATPase)</th>
<th>Penetrating anion (slight or no inhibition of K⁺ ATPase)</th>
<th>Permissive molecule</th>
</tr>
</thead>
<tbody>
<tr>
<td>Chloride</td>
<td>Acetate</td>
<td>Δ-Malate for citrate</td>
</tr>
<tr>
<td>Bromide</td>
<td>Formate</td>
<td></td>
</tr>
<tr>
<td>Iodide</td>
<td>Phosphate</td>
<td></td>
</tr>
<tr>
<td>Nitrate</td>
<td>Phosphate</td>
<td></td>
</tr>
<tr>
<td>Citrate</td>
<td>Phosphate</td>
<td></td>
</tr>
</tbody>
</table>

In conclusion, nigericin is an antibiotic which seems to have biphasic properties. At concentrations below 1 μg per ml it specifically interacts with the ion-translocating system of the mitochondrial membrane (8, 12) without induction of ATP hydrolysis. Above this concentration it induces an ATPase activity in mitochondria, adding its alkali metal cation-dependent uncoupling action (37) to its specific inhibitory effect on the mechanism of accumulation of K⁺ (8, 12) and inorganic phosphate (37, 38) in mitochondria.

REFERENCES

Potassium Ion-dependent Hydrolysis of Adenosine Triphosphate Induced by Nigericin in Mitochondria
Sergio Estrada-O, Stanley N. Graven and Henry A. Lardy


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