Effects of Temperature on Conformation, Hydroxylation, and Secretion of Chick Tendon Procollagen*

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SUMMARY

When freshly isolated chick embryo tendon fibroblasts were incubated at 37° with [14Cproline and 0.5 mM α,α'-dipyridyl, a ferrous iron chelator which inhibits prolyl hydroxylase, the labeled unhydroxylated procollagen molecules accumulated intracellularly. When pepsin digestion at 15° was used as an enzymatic probe of conformation, these molecules were shown to be non-triple helical, indicating that hydroxylation of proline is necessary for triple helix formation at normal body temperatures. When cells which had accumulated unhydroxylated procollagen at 37° were then incubated at 25°, large amounts of unhydroxylated procollagen were secreted into the medium. Although these secreted molecules were sensitive to pepsin they were shown to be largely triple helical when prolyl hydroxylase was used as another probe of collagen conformation. The retained molecules were shown to be largely non-triple helical by this same test. This result suggests that triple helix formation may be necessary for normal secretion of procollagen by these cells.

When cells which had accumulated unhydroxylated procollagen at 37° were then incubated at different temperatures between 20° and 37° in the presence of excess ferrous iron to reverse the inhibition of prolyl hydroxylase, the accumulated molecules were secreted and the extent of hydroxyproline formation increased as the temperature during the reversal period increased. At 20° less than 1% of the [14C]proline in the secreted molecules was hydroxylated while at 37° this value increased to 33%. This result implies that under some conditions the body temperature of an organism may in part regulate the hydroxyproline content of the collagen molecules.

Recent studies from several laboratories have demonstrated that collagen is first synthesized as a precursor molecule called procollagen (1–4). The procollagen molecule is composed of three polypeptide chains which are larger than the α chains of interstitial collagen because of an extension present at the NH₂-terminus of each chain. These extensions are each approximately 25,000 daltons and differ from the α chain portion in their amino acid composition and conformation. The extensions are relatively poor in imino acids, relatively rich in acidic amino acids (5), and are susceptible to a number of proteolytic enzymes including pepsin (1–3). In contrast, the rest of the molecule is in a triple helical conformation, which is resistant to proteolysis by these enzymes and becomes susceptible to proteolysis only if the triple helical conformation is disrupted by heating or other means (6). Recent evidence indicates that the extensions present in the procollagen molecule are held together by disulfide bonds (7–9). The role of the extensions has not been clearly defined, but it has been postulated that they may serve to initiate triple helix formation by serving as registration peptides, they may facilitate secretion of collagen into the extracellular matrix, or they may control fibrillogenesis (10).

Although hydroxyproline occurs in all vertebrate interstitial collagens, it has been thought that it was not essential for the structural integrity of the collagen molecule, and its function has remained obscure (11). Recently we (12) and Berg and Prockop (13) have shown that hydroxyproline appears to stabilize the collagen triple helix. In further studies of a series of procollagen samples, in which the hydroxyproline content varied from <1% to 44% of the total imino acids, was prepared from chick tendon fibroblasts, and the thermal stability of these procollagen preparations was then investigated by using pepsin digestion at different temperatures as an enzymatic probe of conformation (14). With the use of this technique, the denaturation temperature of the procollagen was found to be directly proportional to the hydroxyproline content. A denaturation temperature of 23.5° was found for the unhydroxylated procollagen and 37.9° for fully hydroxylated procollagen. These values agree well with measurements of denaturation temperatures made by optical rotation on unhydroxylated and fully hydroxylated collagen (13). These results also agree with the observation that the synthetic peptide (Pro-Hyp-Gly)₁₀ denatures at a significantly higher temperature than (Pro-Pro-Gly)₁₀ and that proline hydroxylation increases the thermal stability of the peptide α₁CB2

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obtained by CNBr cleavage of collagen from rat tail tendon and rat skin (15, 16).

Previous experiments with embryonic chick tibiae (17) and fibroblasts (18-21) indicated that when the hydroxylation of proline and lysine was inhibited, unhydroxylated collagen molecules were secreted at a markedly reduced rate and accumulated intracellularly. In the case of one type of fibroblast, it has been shown that the unhydroxylated molecules which were secreted were susceptible to pepsin at 15°C and, therefore, they were not entirely in a triple helical conformation (22). Since triple helix formation normally occurs intracellularly before collagen is secreted (23), it is possible that a reduction in hydroxyproline content could prevent such helix formation and cause a significant decrease in the rate of secretion.

In the present study we demonstrate that (a) intracellular unhydroxylated collagen is not triple helical at 37°C, which indicates that hydroxylation of proline is necessary for triple helix formation at normal body temperatures; (b) unhydroxylated procollagen molecules can be secreted by cells incubated at temperatures at which the molecules are largely triple helical, suggesting that triple helix formation may be required for normal secretion; and (c) when cells which have accumulated unhydroxylated procollagen molecules at 37°C are allowed to hydroxylate these molecules at different temperatures, then the degree of hydroxylation which can be achieved increases as the temperature increases. This result implies that under some conditions the body temperature of an organism may in part regulate the hydroxyproline content of the collagen molecules.

MATERIALS AND METHODS

Materials—[U-14C]Proline, 200 μCi per μmole, was purchased from New England Nuclear (Boston, Mass.); crystalline pepsin, colchicine, cycloheximide, sodium dodecyl sulfate, and mercaptoethanol were purchased from Sigma Chemical Co. (St. Louis, Mo.). Reagents used for acrylamide gel electrophoresis and α,α′-dipyridyl were purchased from Eastman Kodak Co. (Rochester, N. Y.).

Isolation of Fibroblasts and Incubation Conditions—Fibroblasts were isolated from 17-day-old chick embryo tendons as previously described (24). Subsequently 109 cells per ml were incubated at 37°C in Krebs medium containing 2% fetal calf serum, 25 μg per ml of ascorbic acid, [14C]proline, and either 0.5 mM α,α′-dipyridyl or 1 μM colcemide. The cells were incubated at the temperatures indicated and for the intervals shown for each experiment. In experiments in which it was desired to reverse the inhibition of prolyl hydroxylase by α,α′-dipyridyl, the cell suspensions were centrifuged and the cells resuspended in fresh media containing 1 mM FeSO4 and the incubations continued for the desired intervals. Further details of particular experiments are given in the tables and figures.

Treatment of Collagen Preparations with Pepsin—To determine whether the intracellular collagen synthesized by the cells under various experimental conditions was triple helical, limited pepsin digestion was used as an enzymatic probe of conformation (6, 12).

To avoid changes in the conformation of the molecules, the samples were rapidly titrated to pH 2.0 with 1 N HCl, and pepsin previously dissolved in 10-2 M HCl was added to a final concentration of 100 μg per ml. All manipulation of samples after termination of the incubations was carried out at room temperature as opposed to the usual temperature of 4°C in order to prevent possible reformation of triple helical molecules. The total elapsed time from the end of the incubation until the addition of pepsin was always less than 2 min. The pepsin digestion was performed at 15°C for 6 hours. In some experiments the duration of pepsin digestion was extended to 16 hours without any apparent change in results. At the end of the digestion, the solutions were neutralized, sodium dodecyl sulfate and mercaptoethanol were added to final concentrations of 1%, and the samples immediately placed in a boiling water bath for 9 min. The samples were then dialyzed against 0.01 M sodium phosphate, pH 7.0, 0.1% sodium dodecyl sulfate, and 0.1% mercaptoethanol and aliquots run on sodium dodecyl sulfate polyacrylamide gels or on 6% agarose columns.

Disc Gel Electrophoresis and Agarose Chromatography on Sodium Dodecyl Sulfate—Disc gel electrophoresis in sodium dodecyl sulfate was performed as described previously (12) with the use of 5% acrylamide gels and half the standard amount of cross linker. The gels were immediately chilled and cut into 1.5-mm fractions in order to minimize losses, and the fractions were solubilized with 30% H2O2 and counted in a scintillation counter. Duplicate gels were run containing radioactive samples and purified α and β rat tail collagen. These gels were stained with Coomassie blue and the radioactive peaks located relative to these markers. The positions of the radioactive peaks were identical in the stained and unstained gels when corrected for gel swelling during destaining. Radioactively labeled collagen samples were chromatographed on sodium dodecyl sulfate-agarose columns (Bio-Gel P-150, 200 to 400 mesh, Bio-Rad Laboratories, Rockville Centre, N. Y.) as described previously (3).

Preparation of Prolyl Hydroxylase and Unhydroxylated Procollagen—Prolyl hydroxylase was purified from 13 day old chick embryos as described by Hamo et al. (25). The purification was carried through the DEAE-cellulose chromatography step, and the final enzyme preparation had a specific activity of 45 units per mg of protein when measured with 125 pg per ml of (Pro-Gly-Pro), molecular weight 2400, as substrate (purchased from Miles Laboratories, Inc., Kankakee, Ill.). One unit is defined as that amount of enzyme required to synthesize 1 μg of hydroxyproline in 1 hour at 37°C under conditions in which the concentrations of all other cofactors are saturating. Unhydroxylated [14C]procollagen-labeled procollagen was prepared by incubating isolated fibroblasts with 0.5 mM α,α′-dipyridyl for 3 hours and then extracting the [14C]procollagen from the cells with 0.5 M acetic acid at 2°C (21). The extract was centrifuged at 12,000 × g for 15 min, and the supernatant was then dialyzed exhaustively against 0.6 M acetic acid. This dialyzed material served as substrate for the hydroxylase. Conditions of the hydroxylation reaction are described in Table 1.

RESULTS

Conformational State of Unhydroxylated Procollagen Retained Intracellularly at 37°C When the cells were incubated with [14C]proline and either 0.5 mM α,α′-dipyridyl or 1 μM colcemide for 2 hours, the procollagen synthesized by the cells was secreted at a markedly reduced rate relative to control cells, and the [14C]procollagen accumulated intracellularly. In the case of the α,α′-dipyridyl-treated cells, the [14C]procollagen contained essentially no [14C]hydroxyproline while the [14C]procollagen from the colcemide-treated cells was fully hydroxylated (28). In order to avoid changes in the conformation of the molecules, the cell suspensions were quickly centrifuged at the end of the incubation and aliquots of the resuspended cells immediately incubated with or without pepsin. The digest mixtures were neutralized and subjected to electrophoresis on sodium dodecyl sulfate polyacrylamide gels (Fig. 1). The fully hydroxylated collagen synthesized by the colcemide-treated cells was recovered as procollagen when pepsin was omitted from the digestion. After pepsin digestion, 85% of the radioactivity originally in the procollagen peak was recovered as α chains, thus indicating that it was predominantly triple helical. In contrast, less than 7% of the radioactivity originally in the unhydroxylated procollagen was recovered as α chains after pepsin digestion, and smaller molecular weight products were observed near the tracking dye. This result demonstrates that the unhydroxylated procollagen within the cells at 37°C was not triple helical.

It has been proposed that the additional amino acid residues at the NH₂ termini, in which the chains are linked together by disulfide bonds, may serve as a nucleation site for triple helix formation. It was possible, therefore, that α,α′-dipyridyl could have interfered with the formation of these disulfide link-
Fibroblasts (10⁷ per ml) were incubated with 3 μCi per ml of [¹⁴C]proline and 0.5 mM α,α’-dipyridyl in Krebs medium for 1 hour at 37°C. The cell suspension was then centrifuged at 1500 × g for 2 min and the cells resuspended at 10⁷ per ml in a modified Krebs medium containing 0.1 M N-2-hydroxyethylpiperazine-N’-2-ethanesulfonic acid, pH 7.4, in place of sodium phosphate buffer, 7 mM KCl in place of KH₂PO₄, 0.1 mM α,α’-dipyridyl, and 25 μg per ml of catalase (Sigma Chemical Co.), 0.5 mM FeSO₄, 0.5 mM a-ketoglutarate, 0.1 mM dithiothreitol, 2 mM ascorbate, and 50 mg of bovine serum albumin. The suspensions then centrifuged. The cells were resuspended in an equal volume of fresh N-2-hydroxyethylpiperazine-N’-2-ethanesulfonic acid medium, sonicated briefly at 30°C, and 1-ml aliquots of the supernatants and cells incubated at 25°C or 37°C for 1 hour in a 2-ml reaction mixture, each ml of which contained 4 units of prolyl hydroxylase, 1 mg of bovine serum albumin, 0.5 mg of catalase (Sigma Chemical Co.), 0.5 mM FeSO₄, 0.5 mM α-ketoglutarate, 0.1 mM diithiothreitol, 2 mM ascorbate, and 50 mM Tris-HCl buffer, pH 7.8. Unhydroxylated procollagen which had been denatured by heating to 45°C for 45 min in the N-2-hydroxyethylpiperazine-N’-2-ethanesulfonic acid-Krebs medium immediately prior to incubation with the enzyme was used as a control. The incubations were terminated by addition of 2 ml of 12 M HCl, and the mixtures were then hydrolyzed at 108°C for 24 hours. The [¹⁴C]hydroxyproline content of the samples was then assayed by a specific chemical procedure (27). The values in the table represent the average of duplicate incubations.

<table>
<thead>
<tr>
<th>Hydroxylation temperature</th>
<th>25°C</th>
<th>37°C</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Total Cpm X 10⁶</td>
<td>Degree of hydroxylation</td>
</tr>
<tr>
<td>Denatured unhydroxylated procollagen</td>
<td>36.1</td>
<td>32.3</td>
</tr>
<tr>
<td>Cell incubation (25°C)</td>
<td>Secreted</td>
<td>91.4</td>
</tr>
<tr>
<td></td>
<td>Intracellular</td>
<td>97.2</td>
</tr>
<tr>
<td>Cell incubation (37°C)</td>
<td>Secreted</td>
<td>45.5</td>
</tr>
<tr>
<td></td>
<td>Intracellular</td>
<td>97.6</td>
</tr>
<tr>
<td></td>
<td>Intracellular plus secreted, without enzyme</td>
<td>177.9</td>
</tr>
</tbody>
</table>

a Values are 100 X [¹⁴C]hydroxyproline per total [¹⁴C].

Prepared as described under "Materials and Methods." The degree of hydroxylation was <1%.

Fig. 1. Polyacrylamide sodium dodecyl sulfate gel electrophoresis of intracellular procollagen after incubation with or without pepsin. Aliquots of cells which had been incubated with 1 μM colcemide, A, or 0.5 mM α,α’-dipyridyl, B, and [¹⁴C]proline for 2 hours were then incubated with or without 100 μg per ml of pepsin at pH 2.0 for 6 hours at 15°C. At the end of the digestion the solutions were neutralized, prepared for electrophoresis, run on 5% polyacrylamide gels, and the gels fractionated and counted as described under "Materials and Methods." ○, incubation with pepsin; □, without pepsin; ●, rat tail α chains; ●, rat tail β chains; ○, tracking dye.

at or below 25°C at neutral pH (30, 31). We therefore incubated cells, containing accumulated unhydroxylated procollagen, at 25°C or 37°C in order to compare the rate at which they secreted the unhydroxylated molecules at the two temperatures (Fig. 3). When the cells were incubated at 25°C there was a progressive secretion of the accumulated unhydroxylated molecules so that after 3 hours 41% of the nondialyzable radioactivity was found in the medium (Fig. 3). When the cells were incubated at 37°C, the secretion was irregular, and only 26% was found in the medium. Corresponding to the increase in radioactivity in the medium at 25°C, there was a decrease in the radioactivity found intracellularly. At 37°C the secreted radioactivity was slightly less than the corresponding intracellular decrease.

These results suggested that differences might exist between the structure of the protein secreted at 25°C and 37°C, and aliquots of the secreted proteins were, therefore, subjected to electrophoresis on polyacrylamide gels. The results (Fig. 4) demonstrated that over 60% of the radioactivity in the proteins secreted at 25°C was recovered in the molecular weight range of 125,000. However, when the secreted proteins were digested with pepsin at 15°C before electrophoresis, only lower molecular weight peptides were recovered. This result indicates that although the chains had a molecular weight equal to that of procollagen, they were either completely denatured or contained regions of local...
luted procollagen independently of the effect of temperature on the rate of protein synthesis, the following experiments were performed. Unhydroxylated procollagen was accumulated intracellularly at 37° by incubating cells with 0.5 mm α, ω-dipyridyl for 1 hour. The cells were then washed and resuspended in fresh medium containing 25 µg per ml of cycloheximide to stop protein synthesis and 1 mM FeSO₄ to reverse the inhibition of prolyl hydroxylase. Aliquots of the cell suspension were then incubated at 20°, 25°, 30°, or 37° for 3 hours. At hourly intervals 1-ml aliquots were taken, the cells separated by centrifugation, and the resuspended cells and supernatants dialyzed separately against running tap water. The dialyzed samples were then hydrolyzed and assayed for total radioactivity and [¹⁴C]hydroxyproline content. Zero time in this experiment is after the initial 1-hour incubation at 37°.

<table>
<thead>
<tr>
<th>Cell incubation temperature</th>
<th>Intracellular</th>
<th>Secreted</th>
<th>Degree of hydroxylation</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Total dpm X 10^4</td>
<td>%</td>
<td>Total dpm X 10^4</td>
</tr>
<tr>
<td>Zero time</td>
<td>17.6</td>
<td>0.5</td>
<td>9.4</td>
</tr>
<tr>
<td>20°</td>
<td>21.9</td>
<td>0.7</td>
<td>3.9</td>
</tr>
<tr>
<td>2 hours</td>
<td>20.0</td>
<td>1.8</td>
<td>4.3</td>
</tr>
<tr>
<td>3 hours</td>
<td>19.6</td>
<td>2.3</td>
<td>4.7</td>
</tr>
<tr>
<td>25°</td>
<td>20.0</td>
<td>2.6</td>
<td>3.8</td>
</tr>
<tr>
<td>2 hours</td>
<td>19.5</td>
<td>4.2</td>
<td>4.4</td>
</tr>
<tr>
<td>3 hours</td>
<td>15.2</td>
<td>5.1</td>
<td>5.1</td>
</tr>
<tr>
<td>30°</td>
<td>20.3</td>
<td>8.3</td>
<td>4.2</td>
</tr>
<tr>
<td>2 hours</td>
<td>15.7</td>
<td>9.6</td>
<td>5.8</td>
</tr>
<tr>
<td>3 hours</td>
<td>17.2</td>
<td>8.0</td>
<td>6.9</td>
</tr>
<tr>
<td>37°</td>
<td>19.8</td>
<td>14.2</td>
<td>5.7</td>
</tr>
<tr>
<td>1 hour</td>
<td>17.7</td>
<td>15.5</td>
<td>7.3</td>
</tr>
<tr>
<td>2 hours</td>
<td>15.6</td>
<td>15.6</td>
<td>8.7</td>
</tr>
</tbody>
</table>

* Values are 100 X [¹⁴C]hydroxyproline per total [¹⁴C].

Fig. 4. Polyacrylamide gel electrophoresis of unhydroxylated procollagen from cells incubated at 25°. Cells were incubated as described in Fig. 3, and aliquots of the secreted and intracellular proteins from the 2-hour time point were incubated with or without 100 µg per ml of pepsin for 6 hours at 15°. The incubation mixtures were prepared for electrophoresis and then subjected to electrophoresis as described in Fig. 1. A, secreted proteins; B, intracellular proteins; , incubation with pepsin; , incubation with pepsin, without pepsin. 1, rat tail β chains; 2, rat tail α chains; 3, tracking dye.

DISCUSSION

It was previously reported (21) that unhydroxylated procollagen which was retained intracellularly was triple helical since it was resistant to pepsin digestion at 15° and formed normal segment-long spacing aggregates at 4°. However, more recent experiments (12-16), which demonstrated that hydroxyproline is crucial to the thermal stability of collagen and procollagen, suggested that unhydroxylated procollagen might not be triple helical within the cell at 37°. In the present experiments the conformation of the intracellular unhydroxylated procollagen was examined by pepsin digestion under conditions designed to mini-
mize the opportunity for conformational changes in the protein. Under these conditions 93% of the unhydroxylated molecules were digested while only 15% of the hydroxylated intracellular molecules were digested. By this test then the unhydroxylated molecules were largely not triple helical while the hydroxylated molecules were triple helical. Since most of both the unhydroxylated and hydroxylated chains appeared to be held together by disulfide linkages (Fig. 2), the difference in the conformation of the two types of molecules cannot be due to prevention of disulfide bond formation by α,α'-dipyridyl. In the earlier report (21) the unhydroxylated collagen was extracted from the cells and apparently reformed native molecules during the extraction and prolonged dialysis period at 4°C.

TABLE III

<table>
<thead>
<tr>
<th>Temperature of hydroxylation</th>
<th>Intracellular</th>
<th>Secreted</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>peak total cpm × 10^3</td>
<td>Degree of hydroxylation</td>
</tr>
<tr>
<td>20°C</td>
<td>55.7 ± 4.2</td>
<td>44.6 ± 0.8</td>
</tr>
<tr>
<td>25°C</td>
<td>47.5 ± 10.6</td>
<td>60.5 ± 3.0</td>
</tr>
</tbody>
</table>

* Values are 100 × [14C]hydroxyproline per total 14C.

The five peak fractions containing the procollagen in each of the four chromatograms illustrated in Fig. 5 were pooled and an equal volume of 12 N HCl added. The samples were hydrolyzed for 24 hours at 108°C and analyzed for their [14C]hydroxyproline content. Pooled peak fractions from similar chromatograms of the intracellular labeled proteins were also analyzed.

Experiments using fibroblasts in culture in which hydroxyproline formation was inhibited by incubation in ascorbate-deficient media (20, 22, 32) or through Fe chelation (18) have indicated that markedly underhydroxylated molecules can be secreted although probably at a reduced rate relative to the secretion of fully hydroxylated molecules. Experiments using freshly isolated tendon fibroblasts showed that total inhibition of hydroxylase prevented secretion of whole procollagen molecules at 37°C. The reason for the differences between the cultured fibroblasts and the freshly isolated fibroblasts is not clear. Perhaps the secretory ability of the freshly isolated cells may have been impaired by the proteolytic digestion required to free them from the extracellular matrix, or alternatively the ability of the fibroblasts in culture to discriminate triple helical from non-helical molecules may be impaired. The present results indicate that the inability of the isolated tendon cells to secrete unhydroxylated procollagen can be overcome by incubating the cells at 25°C, since large amounts of unhydroxylated procollagen were secreted at 25°C and essentially none at 37°C. This is all the more remarkable since we would expect all cellular functions including secretory ability to be considerably slower at 25°C than at 37°C.

The unhydroxylated molecules secreted at 25°C were not perfectly triple helical, however, since they were sensitive to pepsin. Pepsin digestion, as presently used, is a stringent test of triple helicity since a single disruption of the triple helix probably leads to digestion. Recent experiments studying the ability of unhydroxylated procollagen to serve as a substrate for prolyl hydroxylase as a function of temperature have strongly indicated that only procollagen molecules which are not in a triple helical conformation can be hydroxylated (30, 31). We therefore used prolyl hydroxylase as another probe of collagen conformation. The results with the secreted unhydroxylated proteins were fairly clear. The unhydroxylated procollagen secreted at 25°C was hydroxylated to a much greater extent when incubated with prolyl hydroxylase at 37°C than at 25°C. This result suggested that large regions of the molecules were triple helical at 25°C. Control experiments using denatured procollagen proved that completely denatured molecules could be fully hydroxylated at 25°C. The proteins found in the medium at 37°C were relatively poor substrates at both hydroxylation temperatures of 25°C and 37°C, probably because these proteins were either non-collagenous or degraded unhydroxylated collagen. Since the molecules retained

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**Fig. 5.** Agarose gel chromatography of secreted procollagen which was hydroxylated intracellularly at different temperatures. Cells were incubated as described in Table II except that 3 μCi per ml of [14C]proline was used. At the end of the 3-hour reversal period at the different temperatures, the secreted proteins were prepared for chromatography as described under "Materials and Methods." One hundred thousand counts per min of each sample were chromatographed on a column (1.5 × 80 cm) of 6% agarose (Bio-Gel A-5m, 200 to 400 mesh, Bio-Rad Laboratories, Rockville Centre, N. Y.). The fraction size was 2.0 ml. A, 20°C; B, 25°C; C, 30°C; D, 37°C; 1, marker β rat tail collagen; 2, marker α rat tail collagen.

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**TABLE II**

<table>
<thead>
<tr>
<th>Temperature of hydroxylation</th>
<th>Intracellular</th>
<th>Secreted</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>cpm × 10^3</td>
<td>%</td>
</tr>
<tr>
<td>20°C</td>
<td>55.7 ± 4.2</td>
<td>44.6 ± 0.8</td>
</tr>
<tr>
<td>25°C</td>
<td>47.5 ± 10.6</td>
<td>60.5 ± 3.0</td>
</tr>
</tbody>
</table>

* Values are 100 × [14C]hydroxyproline per total 14C.
intradcellularly at 25° were equally good substrates for hydroxyla-
tion at 25° and 37°, they appeared to be less triple helical than
the secreted ones. Although the findings with the molecules
retained at 37° qualitatively agree with the interpretation that
they were largely non-triple helical, one quantitative in-
consistency exists. That is the fact that these molecules were some-
what better substrates at 37° than at 25°. We have no certain
explanation for this observation, but it is possible that limited
helical regions in fact existed intracellularly or were formed either
during the work up of the cells at room temperature prior to the
hydroxylation at 25° or during the hydroxylation at 25° itself.
Nevertheless, by the criterion of their ability to be hydroxylated,
the secreted procollagen molecules were largely triple helical, and
the retained intracellular molecules were largely non-helical, and
it is possible that the proper conformation may be essential for
rapid secretion of procollagen by these cells.

When unhydroxylated procollagen was incubated with purified
prolyl hydroxylase in a cell-free system at different temperatures,
the amount of hydroxyproline which could be formed increased
sharply between 26° and 30°, provided the substrate was not
denatured prior to incubation with the enzyme (30, 31). These
experiments suggested that under certain conditions temperature
increased during the reversal period. One possible
explanation of these results is that partial triple helix formation
was reversed. Hydroxyproline determinations on the iso-
lated procollagen (Table III) showed that the degree of
hydroxylation of the molecules increased markedly when the
enzyme was reversed. Hydroxyproline determinations on the iso-
lated procollagen then hydroxylated intra-
cellularly by reversal of inhibition of prolyl hydroxylase at differ-
temperatures showed that the hydroxylation reaction was es-
tentially complete 2 hours after the inhibition of prolyl hydroxyl-
ase was reversed. Hydroxyproline determinations on the iso-
lated procollagen (Table III) showed that the degree of
hydroxylation of the molecules increased markedly when the
temperature increased during the reversal period. One possible
explanation of these results is that partial triple helix formation
occurs more readily at lower temperatures, and the triple helical
regions of the molecule cannot be hydroxylated further, result-
ing in lower degrees of hydroxylation. These experiments sug-
gest that under some conditions the body temperature of an
organism may in part regulate the hydroxyproline content of
collagen molecules.

Acknowledgments—We thank Miss Sandy Schnee and Mrs.
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REFERENCES


U. S. A. 68, 1138


248, 2285

6. Rubin, A. L., Pfaah, D., Speckman, F. T., Davison, P. J.,
and Schmit, F. O. (1963) Science 193, 37


Sci. U. S. A. 70, 361


Chemistry (Anfinsen, C. B., Jr., Eds., J. T., and Rich-

Biochem. Biophys. Res. Commun. 52, 115

Res. Commun. 52, 115

Arch. Biochem. Biophys. 158, 478

15. Sakakibara, S., Inouye, K., Shudo, L., Kishida, Y.,
Kobayashi, Y., and Prockop, D. J. (1973) Biochim. Biophys.
Acta 303, 198


17. Juv, K., Prockop, D. J., Cooper, G., and Lash, J. (1966)
Science 152, 92

Biophys. 147, 612

Biochem. Soc. Lett. 15, 59

49, 1343

(1973) J. Biol. Chem. 248, 720


Fed. Proc. 32, 649

240, 358

Biophys. Acta 198, 460

242, 4007


Acta 264, 375


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