Mitochondrial Cation-Hydrogen Ion Exchange

SODIUM SELECTIVE TRANSPORT BY MITOCHONDRIA AND SUBMITOCHONDRIAL PARTICLES*

(Received for publication, August 13, 1973)

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SUMMARY

The neutral exchange between Na⁺ or K⁺ ions and protons across the mitochondrial membrane was studied by means of passive swelling in isotonic acetate salts. The selectivity for Na⁺ over K⁺ was approximately 50:1. The rate of Na⁺-dependent swelling mediated by the natural exchange system was sensitive to pH and inhibited 70% or more by 20 mM Mg²⁺. Physiological Mg²⁺ levels, however, inhibited only about 15%. According to direct flux measurements with submitochondrial particles, the endogenous cation-H⁺ exchange possessed an apparent Na⁺:K⁺ selectivity ranging from 6 to 10:1 depending upon the experimental conditions. These ratios were found to be underestimates of the actual selectivity due to translocation of endogenous Na⁺ ions. Values corrected accordingly were comparable to those obtained for intact mitochondria.

Natural mitochondrial cation-H⁺ exchange was compared to exchange mediated by the "ionophore," nigericin. Kinetic studies with mitochondria and submitochondrial particles yielded a Na⁺:K⁺ selectivity ratio of 1.0 for nigericin. Cation discrimination was the major difference between endogenous and nigericin-dependent exchange. Differences between properties of ionophores in model systems and their kinetic behavior in biological membranes are discussed in the context of isolation-reconstitution studies.

The present studies deal with an electrically neutral cation for H⁺ exchange system of the inner mitochondrial membrane (1). Evidence for such an exchange was first provided by observations of Mitchell and Moyle (2) who examined the effects of Na⁺ and K⁺ upon mitochondrial proton ejection and passive swelling of mitochondria in isotonic weak acid salts of Na⁺ or K⁺ (3). These results were interpreted in terms of an endogenous monovalent cation for H⁺ exchange with preference for Na⁺ over K⁺ (see also Ref. 4). Further evidence for the existence of this exchange system was provided by the observation of energy-linked K⁺ uptake by beef heart submitochondrial particles (5, 6) which was stimulated by the model cation-H⁺ exchanger, nigericin (7, 8). Papa et al. (9) recently reported that Mg²⁺ATP submitochondrial particles also possess an apparent nigericin-like endogenous K⁺ permeability similar to that of A-particles (5). These authors demonstrated that Na⁺ and to a lesser degree K⁺ inhibited net H⁺ uptake by these particles (9) consistent with Na⁺ over K⁺ selectivity for mitochondrial exchange (1, 2-4).

Mitchell (10) suggested that neutral cation-H⁺ exchange could be important in osmotic regulation of mitochondria, i.e. to prevent swelling and uncoupling accompanying respiration-dependent uptake of anionic metabolites. We have been interested in this exchange system since it would influence the mitochondrial pH gradient and thus could effect the distribution of anionic substrates between mitochondria and cytoplasm and might, therefore, regulate cellular metabolism (11). Although a number of observations support the existence of neutral mitochondrial cation-H⁺ exchange system, this activity has not been treated quantitatively nor have suitable direct flux measurements been performed. These and related aspects of cation-H⁺ exchange are the subject of the present study.

Through Comparative studies of mitochondria and submitochondrial particles, it has been found that the natural cation-H⁺ exchange is highly selective for Na⁺ over K⁺. This system is quite active at physiological pH and Mg²⁺ levels although Mg²⁺ does inhibit the exchange. The natural exchange system has been compared to the ionophore, nigericin. These observations have not only shed light upon certain aspects of endogenous cation-H⁺ exchange but have revealed interesting differences between cation discrimination derived from kinetic studies with nigericin and assessments of selectivity based upon other techniques.

EXPERIMENTAL PROCEDURES

Preparations—Rat liver mitochondria were isolated by a modification of the method of Schneider (12). Submitochondrial particles (A-particles) were prepared from heavy layer rat liver mitochondria by the technique of Fessenden and Racker (13). All experiments were performed with freshly prepared mitochondria.

† The abbreviations used are: A-particles, submitochondrial particles obtained by sonicating mitochondria in the presence of ammonia; FCCP, carbonyl cyanide trifluoromethoxyphenylhydrazone; EGTA, ethylene glycol bis(β-aminoethyl ether)-N,N'-tetraacetic acid.

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Mitochondrial Swelling Studies: Calculated Fluxes for Comparison with Submitochondrial Particle Measurements—Mitochondrial swelling was recorded continuously with a Beckman DU spectrophotometer as absorbance changes at 600 nm. In all experiments mitochondria were pretreated with respiratory inhibitors before addition to isotonic salt media. Swelling rates are expressed as either ΔA per min or Δ% initial A per min. The latter was used to calculate the net flux of monovalent cations associated with passive swelling. An inverse first power relationship between changes in absorbance and mitochondrial volume was assumed (15). A representative total mitochondrial volume of 3.0 μl per mg of protein (16-18) was used in these computations.

To compare Na+ and K+ fluxes (e.g. per mg of protein values) for mitochondria and submitochondrial particles, the following assumptions were necessary: (a) cation flux was uniform across the entire inner mitochondrial membrane surface; (b) the inner mitochondrial membrane protein constituted 52% of the total (19); (c) submitochondrial particles were derived exclusively from the inner membrane; (d) the membrane protein to surface area ratio was the same for submitochondrial particles and the mitochondrial inner membrane.

Except where indicated, maximum fluxes were reported (e.g. at pH 7.0 for mitochondria and pH 7.5 plus 7.5 mM MgCl2 for submitochondrial particles; see “Results”).

Iontropic Transport Measurements—The suspending medium Na+ or K+ activity was measured with a Beckman 39046 Cl or 39047 cation electrode, respectively, in combination with a remote junction calibration reference (Corning 70062) connected to the reference medium via an agar-KCl bridge. Electrode outputs were fed through Radiometer model 26 expanded scale pH meters to a multichannel potentiometric recorder. Proton transport was measured with a combination pH electrode (Beckman 39030). The combination pH- and cation-selective electrodes were mounted in the side of a 1.5 ml open vessel equipped with rapid overhead mixing by a modified titration assembly (Radiometer M-11).

Na+ and K+ Assays—A particles were extracted with 0.5 N HNO3. The extracts were analyzed for Na+ and K+ by means of a Perkin-Elmer atomic absorption spectrometer (model 303).

A particle Resin Treatment—As described under “Results,” it was necessary to remove endogenous monovalent cations from submitochondrial particles for certain measurements. This was accomplished by pretreating A-particles with Tris-Dowex 50W-X8 cation exchange resin (50 to 100 mesh, Bio-Rad Laboratories). The resin was prepared by washing on a column with 20 bed volumes of 1 N Tris-Cl at pH 7.2 followed by 20 volumes of distilled deionized water. A-particles (4 ml, 15 to 20 mg per ml) were treated with 2 g of Tris-Dowex resin at 0-4°C for 10 min immediately before use.

Reagents—NAD+, NADH, oligomycin, antimycin, rotenone, and yeast alcohol dehydrogenase (salt-free) were purchased from Sigma Chemical Co. Nigericin was supplied by Dr. D. T. Wong of Eli Lilly, Indianapolis, Ind. and Dr. R. L. Hamed, Commercial Solvents Corporation, Terre Haute, Ind. The FCCP used in these studies was contributed by Dr. P. G. Heytler, E. I. Dupont de Nemours and Co., Wilmington, Del.

RESULTS

Mitochondrial Swelling in Isotonic Salts—One assay of electrically neutral monovalent cation exchange by mitochondria is passive swelling of metabolically inhibited mitochondria in isotonic acetate salt media. Swelling presumably occurs by uptake of Na+ or K+ via neutral cation-H+ exchange (3). This establishes a pH gradient which drives acetate uptake (most likely as the free acid) and results in osmotically induced swelling (20).

The results obtained upon addition of respiratory-inhibited mitochondria to medium containing various isotonic salts are shown in Fig. 1. In the presence of Na+-acetate, the rate of mitochondrial swelling was severalfold more rapid than swelling in K+-acetate. These results were consistent with the presence of an endogenous monovalent cation-H+ exchange system with a high Na+:K+ selectivity and agreed qualitatively with previous observations obtained with both beef heart (4) and rat liver mitochondria (3).

Additional evidence that swelling involved an electrically neutral exchange was provided by the observation that the “model” cation-H+ exchanger, nigericin, stimulated the rate of mitochondrial swelling in either Na+ or K+-acetate medium. Appropriate controls (e.g. no swelling in the presence of the nonpermeant anion, chloride, or substantially more rapid swelling in K+ or Na+-acetate plus nigericin) indicated that spontaneous swelling depended solely upon cation-H+ exchange which was rate-limiting. Acetate penetration was not rate-limiting even in the presence of nigericin since spontaneous swelling in NH4+-acetate was more rapid than with the ionophore (not shown).

The influence of pH on exchange was examined since the participation of K+ and Na+ in cation-H+ exchange of rat liver mitochondria, according to oxygen pulse experiments, was reportedly pH dependent (2). In contrast, no significant effect of pH upon beef heart mitochondrial exchange activity was observed (21). As shown in Fig. 2 (left panel), swelling was more...
rapid in Na+ than in K+-acetate medium at all pH values tested although Na+-dependent swelling declined sharply at pH values greater than 7.0. Swelling rates for nigericin plus Na+ or K+ were virtually independent of pH except for a modest decline at pH 8.0.

The ratio of the initial swelling rates in Na+ and K+ media was employed to estimate Na+ : K+ selectivity values which are plotted as a function of pH in Fig. 2 (right panel). A high Na+ : K+ discrimination was evident for the natural exchange system, particularly at neutral or acid pH values. In contrast, the Na+ : K+ selectivity of the ionophore, nigericin, was 1:1 throughout the pH range examined. Variation in selectivity values for endogenous exchange from one mitochondrial preparation to another resulted from small, absolute differences in the rate of K+-dependent swelling. The average Na+ : K+ selectivity ratios for six preparations were 46, 43, 13, and 4.0 at pH values of 6.5, 7.0, 7.5, and 8.0, respectively. The normalized rates of Na+-dependent swelling from pH 6.5 to 8.0 were 22 ± 0.3, 22 ± 5.8, 16 ± 5.9, and 10 ± 4.3 Δ% initial A per min; see “Experimental Procedures.” Corresponding swelling rates in K+ medium at the above pH values were 0.5 ± 0.4, 0.6 ± 0.5, 1.2 ± 0.8, and 2.5 ± 1.5. Thus at acid pH the Na+ : K+ selectivity could exceed 100 if K+-dependent swelling was especially slow. Since K+-dependent fluxes were extremely small and variable the selectivity values reported here must be considered as approximations of the true Na+ : K+ discrimination of endogenous exchange.

Mg2+ influences the monovalent cation permeability of various membranes (22) and is the predominant intracellular divalent cation (23). Therefore, its effect upon endogenous exchange was examined (Fig. 3). Mg2+ strongly inhibited swelling in Na+ medium, i.e. the endogenous exchange. Maximal inhibition (70%) occurred at 20 to 25 mM Mg2+; half-maximal inhibition was obtained with 5.2 ± 0.4 mM Mg2+ (for six preparations). Mg2+ abolished K+-H+ exchange and also inhibited both Na+ and K+-dependent swelling mediated by nigericin (not shown). Since direct interaction between Mg2+ and nigericin is unlikely (24), inhibition of endogenous and nigericin-dependent exchange by Mg2+ may involve alteration of membrane surface charge or structure. The effect of Mg2+ was nonspecific since Ca2+ and La3+ also inhibited. On a concentration basis, Ca2+ and La3+ were 5 and 10 times, respectively, more effective inhibitors than magnesium.2

Inhibition of exchange by polyvalent cations could result from their influence upon the membrane surface potential by “shielding” and possible binding to fixed charges (25). Ionophore-sensitive cation conductances across lipid bilayers are very sensitive to changes in surface potential (25) and stimulation of electrically neutral mitochondrial anion exchanges by various cations has been attributed to alteration of surface charge (26). These latter results seemed particularly relevant since they indicated a possible reciprocal relationship between surface charge and mitochondrial exchange processes, i.e. cations facilitate neutral anion exchanges (26) whereas they inhibit neutral cation exchange. Because pH has characteristic influences upon surface charge and ionic fluxes (25, 26), the mode of action of Mg2+ was further probed by determining the pH profile for Mg2+-induced inhibition of cation-H+ exchange.

As shown in Fig. 4A, Mg2+ inhibition was greatest at acid pH although swelling rates in the absence of Mg2+ were substantially slower at alkaline pH values. The absolute inhibition by Mg2+ as a function of pH is plotted in Fig. 4B. This more clearly indicates that Mg2+ inhibition is most evident at low pH. This was the opposite effect predicted from a strict surface charge influence, i.e. protons should compete with Mg2+ (26) for inhibition of exchange. The influences of Mg2+ and pH may, therefore, be more complex and involve changes in membrane structure or direct effects upon the exchange system. Alternatively, inhibition by exogenous Mg2+ may be underestimated at high pH due to inhibition by endogenous Mg2+. High pH could, therefore, inhibit exchange by facilitating binding of endogenous Mg2+ to the inner membrane. This proposal has the advantage of providing a single explanation for inhibition by high pH and polyvalent cations (Mg2+, Ca2+, and La3+), i.e. diminution of exchange by positive charges within the membrane or at its surface. This relatively simple mechanism for a control of cation-H+ exchange by the membrane surface potential is offered as a working hypothesis and requires additional study.

1 M. Douglas and R. Cockrell, manuscript in preparation.
In order to assess better the endogenous neutral exchange system, submitochondrial particles were examined since they provided the most convenient system for direct measurement of the relevant cation fluxes.

Monovalent Cation Accumulation by A-particles—A-particles (13) or EDTA-particles (see Ref. 27) were utilized because they can be “coupled” by oligomycin (9, 27–31). In sucrose plus glycylglycine medium at pH 7.5 the respiratory control indices with FCCP were 6.0 ± 0.5 for seven preparations. In the presence of high levels of Tris-NO₃ (which were optimal for K⁺ and Na⁺ ion transport measurements) the respiratory control indices were significantly lower (3.7 ± 0.3 for three preparations also at pH 7.5). The difference in respiratory control in the presence and absence of Tris-NO₃ cannot be readily explained at the present time.

Maximal cation fluxes in submitochondrial particles require the presence of a suitably permeant anion (5, 9, 31). Uncoupling of submitochondrial particles by NH₄⁺ salts can be correlated with the ability of anions to permeate as charged species, i.e., electrogenically (31). Although SCN⁻ was the most permeable anion (Table I), NO₃⁻ was routinely employed as the permeant anion of choice since SCN⁻ has complex secondary effects (32, 33).

Representative tracings of energy-linked Na⁺ and K⁺ uptakes by A-particles are provided in Fig. 5. The initial rate and extent of Na⁺ uptake was 5- to 6-fold greater than the corresponding values for K⁺ accumulation. Cation uptake was energy-linked since it was reversed by uncoupling agent (FCCP) or abolished if uncoupler was added prior to initiation of respiration (not shown). Evidence that cation uptake was mediated by an endogenous neutral exchange system was provided by the observation that nigericin stimulated both Na⁺ and K⁺ accumulation, each of which was reversed by FCCP (Table II). The initial rate and net uptake of Na⁺ promoted by nigericin was slightly less than K⁺ uptake.

Further documentation that net influx of Na⁺ (K⁺ not shown) as well as proton uptake by these submitochondrial particles was energy-linked is provided by the results summarized in Table III. Cation uptake required coupling by oligomycin and was abolished by uncoupling agent or respiratory inhibitors. Therefore, the metabolically dependent uptake of protons by submitochondrial particles (i.e., in the opposite direction of intact mitochondria, cf. Refs. 30, 34, and 35) provides the pH gradient necessary to drive monovalent cation accumulation facilitated by the endogenous cation-H⁺ exchange system or the carboxylic ionophore, nigericin.

A high Na⁺:K⁺ selectivity for endogenous exchange (Fig. 5) was also evident from the extent of energy-dependent proton uptake by A-particles in Na⁺ versus K⁺ medium (Fig. 6A). In K⁺ medium, net proton uptake was approximately one and one-half times the extent of H⁺ accumulation in Na⁺ medium. In agreement with the lack of discrimination by nigericin between Na⁺ and K⁺ (Table II), the ionophore virtually abolished proton uptake in the presence of either cation (Fig. 6A). It is noteworthy that Mg²⁺ increased the extent of H⁺ uptake in both Na⁺ and K⁺ medium to approximately the same value (Fig. 6B). This is consistent with Mg²⁺ inhibition of mitochondrial cation-H⁺ exchange (Fig. 4); however, additional studies revealed that the influence of Mg²⁺ is more complex.

Kinetic studies demonstrated saturation behavior for endogenous neutral exchange. Table II presents the initial rate and net uptake of Na⁺ and K⁺ by A-particles and the kinetic parameters derived from these measurements. These data included 7 experiments per point, with an additional 2 experiments for Na⁺ net uptake conducted with untreated A-particles. The kinetic constant for Na⁺ accumulation was 1.5 × 10⁻⁵ M, while that for K⁺ accumulation was 1.3 × 10⁻⁶ M. The initial uptake rate of Na⁺ was 3.0 ng atom O/mg min, while that of K⁺ was 1.7 ng atom O/mg min. The initial rate of Na⁺ uptake was 2.6, while that of K⁺ was 1.3 ng atom O/mg min.

The endogenous cation-H⁺ exchange system was inhibited by FCCP, a mitochondrial uncoupler that dissipates the proton gradient. FCCP was added to the medium as indicated in the legend of Fig. 5. The initial uptake rate of Na⁺ was 2.0 ng atom O/mg min, while that of K⁺ was 1.0 ng atom O/mg min. The initial rate of Na⁺ uptake was 1.5, while that of K⁺ was 0.8 ng atom O/mg min.

The endogenous cation-H⁺ exchange system was also inhibited by ADH, an alcohol dehydrogenase that catalyzes the oxidation of ethanol. ADH was added to the medium as indicated in the legend of Fig. 5. The initial uptake rate of Na⁺ was 1.0 ng atom O/mg min, while that of K⁺ was 0.5 ng atom O/mg min. The initial rate of Na⁺ uptake was 0.5, while that of K⁺ was 0.25 ng atom O/mg min.

The endogenous cation-H⁺ exchange system was also inhibited by ethanol, an alcohol that is oxidized by ADH. Ethanol was added to the medium as indicated in the legend of Fig. 5. The initial uptake rate of Na⁺ was 0.5 ng atom O/mg min, while that of K⁺ was 0.25 ng atom O/mg min. The initial rate of Na⁺ uptake was 0.25, while that of K⁺ was 0.125 ng atom O/mg min.

The endogenous cation-H⁺ exchange system was also inhibited by ethanol, an alcohol that is oxidized by ADH. Ethanol was added to the medium as indicated in the legend of Fig. 5. The initial uptake rate of Na⁺ was 0.25 ng atom O/mg min, while that of K⁺ was 0.25 ng atom O/mg min. The initial rate of Na⁺ uptake was 0.25, while that of K⁺ was 0.25 ng atom O/mg min.

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TABLE III

Energy-linked cation uptake by submitochondrial particles

A-particles (2.6 mg treated with 2.5 μg of oligomycin per mg) were equilibrated for 2 min in 1.0 ml of 250 mM sucrose, 50 mM Tris-NO₃, 20 mM glycylglycine (pH 7.2) and 0.5 mM NaCl for monitoring sodium movements or 250 mM sucrose, 50 mM KNO₃, and 3 mM glycylglycine (pH 7.2) in the case of proton measurements. Cation flux was initiated by alcohol dehydrogenase addition as indicated in Fig. 5.

<table>
<thead>
<tr>
<th>Additions</th>
<th>Net cation uptake (meq/mg)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Complete system</td>
<td>Na⁺</td>
</tr>
<tr>
<td>Oligomycin</td>
<td>19.7</td>
</tr>
<tr>
<td>+ FCCP (4 μM)</td>
<td>0</td>
</tr>
<tr>
<td>+ Rotenone (0.4 μg/mg)</td>
<td>0</td>
</tr>
<tr>
<td>+ Antimycin (0.2 μg/mg)</td>
<td>0</td>
</tr>
<tr>
<td>+ KCN (1 mM)</td>
<td>0</td>
</tr>
</tbody>
</table>

A-Particle Proton Uptake

Fig. 6. Influence of Na⁺ and K⁺ upon submitochondrial particle H⁺ ion transport. A-particles (2.2 mg) treated with oligomycin (2.0 μg per mg) were preincubated in 1.0 ml of K⁺ or Na⁺ medium containing NAD⁺ and ethanol as described in the legend of Fig. 5. The media contained final concentrations of either 112 mM KC1 or NaCl plus 75 mM sucrose and 2.0 mM glycylglycine (at pH 7.5 and ambient temperature). Proton uptakes on addition of alcohol dehydrogenase are shown in A for: 1, Na⁺ medium; 2, K⁺ medium; 3, Na⁺ medium plus nigericin (0.2 μg per mg); 4, K⁺ medium plus nigericin (0.2 μg per mg). In B, tracings 5 and 6 were obtained with particles in Na⁺ and K⁺ medium, respectively, plus 5 mM MgCl₂. Proton accumulation was completely reversed by 4 μM FCCP (indicated by arrows). Addition of FCCP prior to alcohol dehydrogenase abolished H⁺ uptake in all cases (e.g. as shown in trace 7 for KCl medium).

ous cation-H⁺ exchange (Fig. 7). The apparent Kₘ value of the endogenous exchange system was about 1 mM for Na⁺ or K⁺, well below the intracellular concentration of these ions. Initial studies revealed that submitochondrial particles cation uptake rates were unchanged or slightly decreased at increasing K⁺ or Na⁺ levels. It was, therefore, necessary to remove endogenous cations by resin treatment (see “Experimental Procedures”) in order to obtain the results shown in Fig. 7. Resin treatment of A-particles reduced endogenous Na⁺ from 12.4 to 5.0 neq per mg of protein and K⁺ was decreased from 2.8 to <0.1 neq per mg. Respiration rates and respiratory control ratios were not significantly affected by resin treatment. The kinetics of endogenous exchange, however, appear to be altered (e.g., the apparent Kₘ was increased); furthermore, cation uptakes of different preparations varied more than those of un-

Fig. 7. Kinetics of K⁺ and Na⁺ accumulation by submitochondrial particles. A-particles (3.2 mg) which had been pretreated with cation exchange resin (see “Experimental Procedures”) were suspended in medium at a pH of 7.5. Further experimental details are the same as those in the legend of Fig. 5 except for differences in the initial Na⁺ or K⁺ concentrations.

Fig. 8. Influence of pH upon Na⁺ and K⁺ fluxes in submitochondrial particles. The conditions were as described in the legend of Fig. 5 except 5.2 mg of A-particles were incubated in media containing either 0.5 mM NaCl or KC1 at the indicated pH values. In these experiments, Na⁺ or K⁺ ion fluxes were each measured with the Beckman 39047 electrode to avoid artefacts due to pH changes registered by the Na⁺ selective electrode at acid pH where buffer capacity was limited.

trated particles. Resin-treated submitochondrial particles were, therefore, not employed routinely but only in these kinetic studies.

Since the Na⁺:K⁺ selectivity for exchange in A-particles was significantly lower than that of mitochondria, the experimental conditions were modified in an attempt to maximize the Na⁺:K⁺ selectivity of submitochondrial particles. As observed in the case of mitochondrial swelling, the rate of Na⁺ flux via the natural exchanger was found to be pH dependent (Fig. 8). The maximum Na⁺ uptake rate and, in the experiment shown, the maximum Na⁺:K⁺ selectivity, occurred at pH 7.5. The highest Na⁺:K⁺ discrimination ratios for three preparations examined under these conditions averaged 6.0 ± 0.8 at pH 7.25 to 7.50.

The influence of Mg²⁺ was studied since it effected mitochondrial exchange activity (Fig. 3) and proton uptake by submitochondrial particles (Fig. 6B). Titration with Mg²⁺ to a concentration
of 7.5 mM stimulated Na+ flux (Fig. 9) and increased Na+ : K+ selectivity (Fig. 9, inset). Higher concentrations of Mg2+ (as in mitochondria) reduced both the rate of Na+ transport and the Na+ : K+ selectivity of the endogenous exchange. Maximum Na+ : K+ selectivity values of 10.6 ± 3.1 were obtained with 3.0 to 7.5 mM Mg2+ by similar titrations of several submitochondrial particle preparations. It should be noted that Mg2+ did not significantly stimulate K+ uptake in spite of its marked enhancement of Na+ transport. This could indicate that flux of these cations is mediated by separate systems.

Stimulation of submitochondrial particle Na+ flux by low Mg2+ concentrations is clearly different from the strictly inhibitory influence of this cation upon the exchange system in mitochondria. The more rapid rates of Na+ uptake may reflect a "coupling" influence of Mg2+, presumably related to the enhancement of other energy-linked reactions of submitochondrial particles by low levels of Mg2+ (30). This would be consistent with the observation that Mg2+ increases the rate and extent of proton uptake by A-particles (Table IV) at the same concentration which enhances Na+ accumulation via the endogenous exchange. On the other hand, this same Mg2+ level inhibited Na+ exchange mediated by nigericin (Table IV) indicating that ionophore-mediated transport may be more sensitive to this divalent cation. Higher Mg2+ concentrations inhibited Na+ flux mediated by endogenous exchange (Fig. 9) as well as nigericin (not shown).

Net H+ uptake remained elevated even at high Mg2+ levels (not shown) indicating that inhibition of Na+ flux was not a consequence of diminished H+ uptake but more likely an effect upon exchange activity per se (as in mitochondria; cf. Fig. 3).

As alluded to earlier, the influence of Mg2+ upon submitochondrial particles is complex. Mg2+ could stimulate H+ uptake by either a coupling effect or by inhibiting cation-H+ exchange. Since low levels of Mg2+ also stimulate Na+ accumulation (Fig. 9), the observed Na+ and H+ fluxes most likely result from a dual influence of Mg2+, i.e. coupling and inhibition of exchange.

The general properties of natural cation-H+ exchange are summarized in Table V for comparison with the model exchanger, nigericin. As in mitochondria (Fig. 2), the endogenous exchange system of submitochondrial particles invariably exhibited a significant preference for Na+ over K+. The Na+ : K+ selectivity ratio for nigericin was 1.0 in these studies under all experimental conditions employed (cf. Fig. 2 and Table II). Under optimum conditions, the rate of Na+ uptake mediated by the natural exchange system approached that promoted by the ionophore (Table II). Saturation kinetics were demonstrated for endogenous (Fig. 7) or nigericin-dependent exchange. The latter had an apparent Kₘ for Na+ or K+ of approximately 2 mM. These comparisons indicate that the natural cation-H+ exchange system although similar in certain respects to the model exchanger, nigericin, differs markedly in cation selectivity.

DISCUSSION

Passive mitochondrial swelling and cation uptake by submitochondrial particles have provided reliable means for assessing neutral cation-H+ exchange as indicated by comparison of flux values obtained with these two techniques. The Na+ flux calculated from passive swelling (88 neq per mg-min; see "Experimental Procedures") agrees well with the net Na+ flux in A-particles measured directly (75 neq per mg-min). Passive swelling measurements thus appear quite reliable but also offer certain advantages over other methods such as the "oxygen pulse" tech-
nique of Mitchell and Moyle (2, 37) which requires extensive anaerobic equilibration of mitochondria (e.g. 20 min; see Ref. 37) during which their membrane permeability may change.

Incubation of mitochondria anaerobically for 20 min in the absence of EGTA causes uncoupling as evidenced by loss of respiratory control (38). We have observed that after 20 min, aerobic incubation without exogenous substrate, Na\(^{+}\), but not K\(^{+}\) or choline, uncouples mitochondria (11). This may be a consequence of Mg\(^{2+}\) loss (39) which can increase mitochondrial Na\(^{+}\) conductance (40). Na\(^{+}\)-dependent uncoupling would reflect Na\(^{+}\) conductance plus neutral Na\(^{+}\)-H\(^{+}\) exchange resulting in cyclic Na\(^{+}\) and proton flow. This mechanism might explain early reports of different influence of Na\(^{+}\) and K\(^{+}\) upon mitochondrial respiration and phosphorylation (41, 42). Therefore, caution must be exercised in interpreting measurements of monovalent cation-H\(^{+}\) exchange after extensive incubation at a relatively low energy state such as prevails in oxygen pulse experiments (3). Permeability properties may change as already mentioned and could be part account for certain apparent discrepancies between previous observations related to cation-H\(^{+}\) exchange, e.g. pH dependence (3, 21).

According to oxygen pulse measurements, Na\(^{+}\)-H\(^{+}\) exchange declines as the pH is increased from 6.8 to 8.0; furthermore, K\(^{+}\)-H\(^{+}\) exchange activity is extremely low except at pH values of 6.0 or below (3). Brierley et al. (21) observed that apparent endogenous Na\(^{+}\)-H\(^{+}\) exchange was relatively insensitive to pH although these results were obtained with beef heart rather than rat liver mitochondria employed in the oxygen pulse studies. The results of the present study (cf. Fig. 2) for the pH dependence of endogenous exchange agree qualitatively with those reported by Mitchell and Moyle (3) at least over the pH interval from 6.5 to 8.0. Our observations do not agree with the report of substantial K\(^{+}\)-H\(^{+}\) exchange at low pH (2, 3). The almost negligible K\(^{+}\)-H\(^{+}\) exchange described here supports Caswell’s suggestion (43) that no significant K\(^{+}\)-H\(^{+}\) exchange occurs in isolated mitochondria.

In the present investigation mitochondrial- and submitochondrial particles-catalyzed K\(^{+}\)-H\(^{+}\) exchange differed significantly. The apparent K\(^{+}\) flux computed for submitochondrial particles, 0.2 neq per min·mg was approximately 2.5 times the calculated K\(^{+}\) flux derived from passive mitochondrial swelling experiments (2.4 neq per mg·min). The K\(^{+}\) flux in mitochondria was calculated from values obtained at pH 7.0 where these were the maximum rates attributable to exchange. Although not shown, slow but significant mitochondrial swelling occurred in KCl medium at pH values of 7.5 and greater; therefore, at high pH swelling could involve K\(^{+}\)-H\(^{+}\) exchange and salt penetration unrelated to exchange.

4 The K\(^{+}\) flux in mitochondria was calculated from values obtained at pH 7.0 where these were the maximum rates attributable to exchange. Although not shown, slow but significant mitochondrial swelling occurred in KCl medium at pH values of 7.5 and greater; therefore, at high pH swelling could involve K\(^{+}\)-H\(^{+}\) exchange and salt penetration unrelated to exchange.

### Table VI

<table>
<thead>
<tr>
<th>Assay system</th>
<th>K(^{+})/Na(^{+}) selectivity</th>
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<tr>
<td>Two phase cation distribution</td>
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</tr>
<tr>
<td>Three-phase cation migration</td>
<td>3 to 5:1</td>
</tr>
<tr>
<td>Phospholipid vesicle volume changes</td>
<td>4:1</td>
</tr>
<tr>
<td>Cation release from erythrocyte “ghosts”</td>
<td>2:1</td>
</tr>
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<td>Mitochondrial swelling</td>
<td>1:1</td>
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<tr>
<td>Submitochondrial particle cation uptake</td>
<td>1:1</td>
</tr>
</tbody>
</table>

* Ref. 40.
* B. Pressman, personal communication.
* Calculated from Ref. 8.
* Present studies.

ACKNOWLEDGMENT—The authors wish to thank Mr. R. B. Work who performed the swelling experiments.

## REFERENCES

34. Pressman, B., and Hеб, M. (1971) Biophys. Soc. Abs. 11, 301a
Mitochondrial Cation-Hydrogen Ion Exchange: SODIUM SELECTIVE
TRANSPORT BY MITOCHONDRIA AND SUBMITOCHONDRIAL
PARTICLES

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