Nucleosome assembly proteins (NAPs) are histone chaperones that are essential for the transfer and incorporation of histones into nucleosomes. NAPs participate in assembly and disassembly of nucleosomes and in chromatin structure organization. Human malaria parasite Plasmodium falciparum contains two nucleosome assembly proteins termed PfNapL and PfNapS. To gain structural insights into the mechanisms of NAPs, we have determined and analyzed the crystal structure of PfNapL at 2.3 Å resolution. PfNapL, an ortholog of eukaryotic NAPs, is dimeric in nature and adopts a characteristic fold seen previously for yeast NAP-1 and Vps75 and for human SET/TAF-1b (β1)/INHAT. The PfNapL monomer is comprised of domain I, containing a dimerization α-helix, and a domain II, composed of α-helices and a β-subdomain. Structural comparisons reveal that the “accessory domain,” which is inserted between the domain I and domain II in yeast NAP-1 and other eukaryotic NAPs, is surprisingly absent in PfNapL. Expression of green fluorescent protein-tagged PfNapL confirmed its exclusive localization to the parasite cytoplasm. Attempts to disrupt the PfNapL gene were not successful, indicating its essential role for the malaria parasite. A detailed analysis of PfNapL structure suggests unique histone binding properties. The crucial structural differences observed between parasite and yeast NAPs shed light on possible new modes of histone recognition by nucleosome assembly proteins.

Chromatin assembly and remodeling are vital cellular processes that are pivotal during replication, transcription, recombination, and repair in eukaryotic cells. Nucleosomes are the fundamental repeating subunits of all eukaryotic chromatin (except when packaged in sperm). Nucleosomes are made up of DNA and four pairs of histone proteins, which together resemble “beads on a string” when observed with an electron microscope. Histones H2A, H2B, H3, and H4 are part of the nucleosome, whereas histone H1 sits on top of the structure, keeping in place the DNA that is wrapped around the other histones (1). Nucleosome assembly is a fundamental biological process that is required for the replication and maintenance of chromatin in the eukaryotic nucleus. In dividing cells, newly synthesized DNA is assembled rapidly into chromatin by a process that appears to involve an indirect coupling between DNA replication and nucleosome assembly. The assembly of nucleosome occurs in a two-step process: a tetramer of H3 and H4 is first deposited onto DNA, and then the nucleosome core is completed by the addition of two heterodimers of H2A and H2B (1, 2). Histones are highly basic proteins and require chaperones that promote their proper interaction to form chromatin. Histone chaperones also prevent nonspecific aggregation of DNA with histones. A number of histone chaperones (e.g. nucleosome assembly protein 1 (NAP-1), nucleosplasin, Asf1 (antisilencing function), CAF-1 (chromatin assembly factor), and HirA (histone regulator A)) are involved in nucleosome/chromatin assembly/disassembly (3).

Some histone chaperones, such as nucleosplasin and NAP-1, exhibit a preference for binding to histones H2A and H2B relative to histones H3 and H4 (4, 5). Other histone chaperones, which include CAF-1, N1/N2, and Spt6, associate preferentially with H3 and H4. Interestingly, it has also been observed that newly synthesized histones are acetylated (such as at positions 5, 8, and 12 of histone H4) and then subsequently deacetylated after assembly into chromatin (6, 7). Thus, factors that mediate histone acetylation or deacetylation may participate, perhaps indirectly by the covalent modification of histone.
Structure of Malaria Parasite Nucleosome Assembly Protein

tones, in the chromatin assembly process. Homologs of yeast NAP-1 are conserved among all eukaryotes, and yeast NAP-1 is well characterized for its role in chromatin assembly (3, 8). Gene knock-out phenotypes in several studied organisms indicate essentiality of the nucleosome assembly protein family; gene ablations in mouse and Drosophila cause embryonic lethality, whereas in yeast, cells exhibit growth defects (9–11).

Malaria is one of the most common infectious diseases and remains an enormous public health problem. Malaria is caused by protozoan parasites of the genus Plasmodium, and the most serious forms of the disease are caused by Plasmodium falciparum (12). It is therefore crucial to identify, dissect, and exploit the molecular motors of malaria parasites, which can serve as essential targets for antimalarials. The human malaria parasite P. falciparum contains two nucleosome assembly proteins, which we have termed PfNapL and PfNapS and which are orthologs of eukaryotic NAPs (13). We have shown that both PfNapL and PfNapS are present in all erythrocytic stages of the parasite (13). PfNapL forms complexes with both histone tetramer and octamer and is predominantly localized in the cytoplasm, whereas PfNapS is localized both in cytoplasm and the nucleus (13). PfNapL and PfNapS are present in all erythrocytic stages of the parasite (13). PfNapL forms complexes with both histone tetramer and octamer and is predominantly localized in the cytoplasm, whereas PfNapS is localized both in cytoplasm and the nucleus (13). PfNapL and PfNapS are present in all erythrocytic stages of the parasite (13). PfNapL forms complexes with both histone tetramer and octamer and is predominantly localized in the cytoplasm, whereas PfNapS is localized both in cytoplasm and the nucleus (13). PfNapL and PfNapS are present in all erythrocytic stages of the parasite (13). PfNapL forms complexes with both histone tetramer and octamer and is predominantly localized in the cytoplasm, whereas PfNapS is localized both in cytoplasm and the nucleus (13).

**EXPERIMENTAL PROCEDURES**

**Protein Crystallization and Data Collection—**PfNapL crystals were obtained at 20 °C by the hanging drop vapor diffusion method using 1 μl of full-length purified PfNapL protein (13, 14) (3 mg ml\(^{-1}\) in buffer containing 25 mM Tris and 25 mM NaCl) and 1 μl of 0.2 M MgCl\(_2\) with 20% polyethylene glycol 3350 (molarity buffer). A single crystal was transferred to cryoprotectant containing a higher concentration of mother liquor (30% polyethylene glycol 3350 and 0.3 M MgCl\(_2\)) supplemented with 0.5 M NaI for 30 s and flash-frozen under a stream of nitrogen gas at 100 K. X-ray diffraction data were collected on an in-house rotating anode MICRO MAX 007 x-ray generator (wavelength of 1.54 Å; Rigaku/MSC) operated at 40 kV and 20 mA with Osmic mirrors (Vari Max HR). The images were recorded using an MAR345dtb imaging plate, and the iodide-single anomalous dispersion (SAD) data set was collected to 3.0 Å resolution. The iodide-soaked crystals of PfNapL belong to the monoclinic space group C2 with cell dimensions of \(a = 76.68\,\text{Å}, b = 37.83\,\text{Å}, c = 79.31\,\text{Å},\) and \(\beta = 99.83^\circ\), having one monomer per asymmetric unit. High resolution native diffraction data to 2.3 Å resolution were collected at the BM14 beamline (European Synchrotron Radiation Facility, Grenoble, France). These crystals also belong to the monoclinic space group C2 with different cell dimensions of \(a = 92.41\,\text{Å}, b = 38.12\,\text{Å}, c = 80.05\,\text{Å},\) and \(\beta = 107.87^\circ\). The diffraction images were processed and scaled with the HKL2000 suite (22).

**Phasing, Structure Determination, Refinement, and Analysis—**The structure was determined using the iodide-SAD technique, and phasing was achieved by utilizing four iodide sites to 3.0 Å resolution using PHENIX (23). An initial model was built automatically without side chains and subsequently rebuilt manually using COOT (24). The high resolution structure was determined by molecular replacement technique using the PfNapL iodide-SAD model. This model to 2.3 Å resolution was further refined using CNS (25) (Table 1). The final model was validated using PROCHECK (26). All figures were generated using Chimera (27). The least square fittings and structural alignment were carried out using LSQMAN (28). Electrostatic potential surfaces were generated using GRASP (29).

**Parasite Culture, Plasmid Constructs, and Transfection—**P. falciparum of the 3D7 strain were cultured in vitro in O+ human erythrocytes using standard conditions (30). For the gene deletion construct, we amplified the 5’- and 3’-targeting segments of PfNapL using the primer pairs aw775/aw776 and aw777/aw778, resulting in 645- and 622-bp products, respectively. After amplification, the targeting segments were cloned into the pCC-1 vector (31) using SacII/Spel for the 5’ EcoRI/AvrII for the 3’ segment, resulting in the pCC-1∆PfNapL plasmid. After proliferation of the plasmid DNA in Escherichia coli PCR103 cells and purification with a Qiagen Maxi kit, 80 μg of plasmid DNA was taken up in cytomix and transfected via electroporation using standard procedures (32). Parasites containing the plasmid were selected for in the presence of 2 μM WR99210 (33). The resulting parasite population was then subjected to two off/on drug cycles for 21 days each to encourage the loss of free plasmid. After each addition of WR99210, a subpopulation was selected on 231 nM 5-fluorocytosine in the presence of WR99210 (to encourage double recombination events) and analyzed via Southern blot. Two independent rounds of transfections were performed and analyzed: Aw775, atccgcggTAATTTGAATTGTTCTTGGC; Aw776, gatactagCCATTAAATAAACAACAG; Aw777, atcgaattcGATATGACCACTATTTCG; Aw778, gatctaggCTGTATGTAATAACACTGGCTG.

To generate transgenic parasites expressing GFP chimeras, we amplified the sequence encoding PfNapL from cDNA and cloned the product into the pARL vector (34) to create PfNapL-pARL. Conserved residues of the putative nuclear export signal (NES) were mutated by yielding the plasmids PfNapL L46A-pARL and PfNapL Q50A-pARL. The expression of chimeric proteins in this vector is driven by the crt promoter. The correctness of each construct was confirmed by DNA sequencing. 3D7 P. falciparum parasites were transfected with 100 μg of...
plasmid DNA (Qiagen) by electroporation and cultured in the presence of 2 μM WR99210 (32).

**Microscopy**—In order to observe the localization of the GFP chimeras in all three generated cell lines, live cells were stained with DAPI and immediately processed for viewing at ambient temperature. Cells were viewed with an Apochromat ×100/1.4 numerical aperture oil DIC lens on a Zeiss Axioskop 2 microscope equipped with a PCO SensiCam (12-bit) camera and Axiovision 3 software. Captured images were processed using Photoshop and ImageJ software (available on the World Wide Web).

**Histone-binding ELISAs**—Residues Asp192, His227, Thr230, Glu266, Lys266, Asp268, Tyr309, Ile136, and Ile147 were mutated to alanine using the QuikChange site-directed mutagenesis kit (Stratagene). Tetramer and octamer forms of calf thymus histone were reconstituted from individual histones (Roche Applied Science) after denaturing and refolding according to the method developed by Tanaka et al. (35). ELISAs were performed by coating individual histones, histone tetramers, or histone octamers (100 ng/well) in microtiter plates (Nunc). The histones were coated on plates in phosphate-buffered saline (PBS) and incubated at 4 °C overnight. The following day, the plates were washed with PBS plus 0.2% Tween 20 (PBST) and blocked with 3% PBS-bovine serum albumin (200 μL/well) and incubated for 1 h at 37 °C. Then 100 μL of purified PfNapL (concentration ranging from 25 to 800 ng) was added to each well in histone-binding buffer (20 mM HEPES, 7.5 mM MgCl2, 1 mM dithiothreitol, 0.5 mM EDTA, and 50 mM KCl, pH 7.5). Plates were incubated for an additional 1 h at 37 °C and washed three times with PBST, and then PfNapL anti-rabbit polyclonal antibodies were added to each well (1:25,000 dilutions). After incubation at 37 °C for 1 h, three more PBST washes were done, followed by the addition of horseradish peroxidase-conjugated secondary antibody (anti-rabbit), and incubated for an additional 1 h at 37 °C. After three more PBST washes, a color reaction was developed using orthophenylenediamine (Sigma) and H2O2 in citrate-phosphate buffer. The optical density was measured at 490 nm using an ELISA plate reader (Molecular Probes). Bovine serum albumin, primary antibodies were separately used as controls (Table 1).

**RESULTS**

**Structure Determination and Overall Structure of PfNapL**—The crystal structure of PfNapL was determined using the iodide-SAD technique at 3.0 Å and refined using high resolution data to 2.3 Å resolution (Table 1). Purified PfNapL crystallized in monoclinic space group C2 with solvent content of 55%, and the asymmetric unit contains one monomer of PfNapL. For a total of 347 residues of full-length PfNapL (molecular mass ~41 kDa), electron density was observed for the central core residues 33–281. The N- and C-terminal regions in PfNapL were presumably cleaved off during protein processing prior to crystallization. The regions 89–91, 138–144, 171–178, 200–221, and 235–246 have weak electron density and are disordered in the overall structure. The final refined model of PfNapL has R factor and R free values of 21.8 and 28.5%, respectively (Table 1).

**TABLE 1**

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Native</th>
<th>Iodide-SAD</th>
</tr>
</thead>
<tbody>
<tr>
<td>Data collection</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cell dimensions</td>
<td>a = 92.41 Å, b = 88.12 Å, c = 80.05 Å, β = 107.87°</td>
<td>a = 76.68 Å, b = 73.83 Å, c = 79.31 Å, β = 99.83°</td>
</tr>
<tr>
<td>Wavelength (Å)</td>
<td>1.0</td>
<td>1.54</td>
</tr>
<tr>
<td>Anomalous scatterer</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Resolution (Å)</td>
<td>50-2.3</td>
<td>50-3.0</td>
</tr>
<tr>
<td>Outer shell resolution (Å)*</td>
<td>2.38-2.3</td>
<td>3.11-3.0</td>
</tr>
<tr>
<td>No. of unique reflections</td>
<td>12,065 (1217)</td>
<td>4606 (451)</td>
</tr>
<tr>
<td>Redundancy</td>
<td>5.1 (5.1)</td>
<td>14.5 (13.2)</td>
</tr>
<tr>
<td>Rmerge b</td>
<td>0.052 (0.238)</td>
<td>0.084 (0.331)</td>
</tr>
<tr>
<td>Completeness (%)</td>
<td>100 (99.9)</td>
<td>99.0 (97.6)</td>
</tr>
<tr>
<td>I/I0</td>
<td>26.1 (4.98)</td>
<td>45.6 (11.4)</td>
</tr>
<tr>
<td>Refinement</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Resolution range (Å)</td>
<td>50–2.3</td>
<td>50–3.0</td>
</tr>
<tr>
<td>No. of reflections (test set)</td>
<td>1219</td>
<td></td>
</tr>
<tr>
<td>Rmerge</td>
<td>21.8/28.5</td>
<td></td>
</tr>
<tr>
<td>No. of protein atoms</td>
<td>1668</td>
<td></td>
</tr>
<tr>
<td>No. of water molecules</td>
<td>74</td>
<td></td>
</tr>
<tr>
<td>B factor (Å²)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Protein atoms</td>
<td>36.6</td>
<td></td>
</tr>
<tr>
<td>Water atoms</td>
<td>35.1</td>
<td></td>
</tr>
<tr>
<td>Stereochemistry</td>
<td></td>
<td></td>
</tr>
<tr>
<td>r.m.s. deviation bond length (Å)</td>
<td>0.01</td>
<td></td>
</tr>
<tr>
<td>r.m.s. deviation bond angle (degrees)</td>
<td>1.50</td>
<td></td>
</tr>
<tr>
<td>Ramachandran plot</td>
<td></td>
<td></td>
</tr>
<tr>
<td>In most favored regions (%)</td>
<td>91.5</td>
<td></td>
</tr>
<tr>
<td>In additionally allowed regions (%)</td>
<td>7.4</td>
<td></td>
</tr>
<tr>
<td>In generously allowed regions (%)</td>
<td>1.1</td>
<td></td>
</tr>
</tbody>
</table>

a Numbers in parentheses are values in the highest resolution shell.

b R_merge = Σ|I_data – ΣI|/ΣI summed over all observations and reflections.

The NAP/SET proteins consist of a central region of ~250 residues that are thought to be primarily responsible for histone binding (Fig. 1a). In general, NAPs comprise low complexity sequences in their N and C termini, wherein the C terminus usually contains stretches of acidic residues (Fig. 1a). The overall fold of PfNapL is similar to yNAP-1 and hSET (15, 16) (Fig. 1c). The phylogenetic analysis of NAPs from homologs in yeast and humans (available on the World Wide Web) (Fig. 1e).
Structure of Malaria Parasite Nucleosome Assembly Protein

Gene Knock-out Studies on PfNapL.—To investigate whether the function of PfNapL could be disrupted, we designed a construct to specifically target the PfNapL gene and to replace it with an human dihydrofolate reductase-selectable marker cassette. Despite several attempts, we were not able to obtain a population where the PfNapL gene was replaced by the human dihydrofolate reductase cassette (Fig. S2). Although we cannot exclude any technical problems, the inability to disrupt the function of PfNapL is indicative of a pivotal role of this gene product for the survival of P. falciparum. At the moment, our repertoire of molecular tools in P. falciparum precludes us from performing conditional gene disruptions. An essential or at least a very beneficial role of this molecule has been described in several other organisms like mouse and Drosophila (9, 10).

Therefore, we propose that the PfNapL protein is very likely to be essential for parasite survival and that it provides a potential new target for disruption of parasite nucleosome assembly as a route toward controlling parasite growth.

Structural Conservation with yNAP-1, hSET, and Vps75—The root mean square (r.m.s.) deviation derived from the least square fittings of 191 Cα of PfNapL with the corresponding Cα of yNAP-1, hSET, and Vps75 is 1.9, 1.8, and 1.8 Å, respectively (Fig. 2a and Fig. S3). The dimerization helix α2 of PfNapL shows an r.m.s. deviation of 1.4 and 1.1 Å with the corresponding dimerization helices of yNAP-1 and hSET (Fig. 2, b and c). The core domain II of PfNapL is also similar to yNAP-1 and hSET, with an average r.m.s. deviation of 1.1 and 1.0 Å, respectively (Fig. 2, b and c). The dimerization helix α2 of PfNapL has high sequence similarity with the corresponding dimerization helices of hSET/yNAP-1 for a total of ~50 residues (Fig. 2, b and c). In the β subdomain, PfNapL has 16 of 24 identical and 5 of 8 conserved residues with the corresponding β subdomains of hSET/yNAP-1. The residues 290–295 of this antiparallel β-sheet region of yNAP-1 had been previously identified as the nuclear localization signal (NLS) (15) (Figs. 1c and 2, b and c). Residues 200–221 in PfNapL corresponding to this short antiparallel β-sheet region (β5 and β6) in yNAP-1 are disordered (Fig. 2b). However, the NLS sequences between yeast and PfNapL are highly conserved (Figs. 1c and 2b).

PfNapL Lacks the “Accessory Domain” Conserved in Higher Eukaryotic NAPs—Comparison between yNAP-1, hSET, and PfNapL structures revealed that yNAP-1 contains an additional α-helix, termed the “accessory domain,” which has been inserted between domain I and II but is absent in both hSET and PfNapL (Figs. 1c and 2a). Based on the sequence comparison, it can be inferred that this accessory domain is present in all other eukaryotic nucleosome assembly proteins but is seen to be specifically absent in PfNapL and other NAPs from Plasmodium species (Fig. 1d). Intriguingly, SET protein homologs, such as the recently characterized human SET (16), are absent in yeast and Plasmodium (stand alone SET domains are found in higher eukaryotes like mice, cows, and humans). In Plasmodium, SET-like domains are a constituent of significantly larger proteins (molecular mass ~142 kDa) termed “SET domain proteins,” where functions and localizations are still unknown (PFD0190w, PFF1440w, and PFO8_0012).

PfNapL Dimer and Conserved Hydrophobic Motifs—The dimer formation in yNAP-1, hSET, Vps75, and PfNapL is achieved via the dimerization helix α2. In PfNapL, residues Gln50, Glu52, Tyr54, Glu57, Lys59, Glu62, Arg65, Lys70, Tyr71, and Lys81 of the dimerization helix α2 and Pro271, Tyr272, and Asp275 of helix α8, which lies on the other side of the β subdomain, contribute to dimer formation. Residues Glu52 and Glu62 and Lys70 and Lys81 of the dimerization helix α2 form salt bridges. The hydrogen bonding interactions contributing to dimer formation in PfNapL are mostly structurally conserved with those of hSET (Fig. 1c). Unlike yNAP-1, the PfNapL dimerization helices are bent at ~50° very similar to the hSET and Vps75 dimers (Fig. S4a and Fig. S5a). Although a proline residue at position 77 is present in PfNapL, there are no kinks formed in the dimerization helices upon dimer formation in PfNapL in contrast to yNAP-1 (Fig. S4a). Interestingly, PfNapL

![FIGURE 1—continued](Image 49x544 to 301x734)

**FIGURE 1.** The overall structure of PfNapL. a, domain diagram for the NAP/SET family of proteins. Histone chaperone proteins comprise a central domain and low complexity N and C termini. The inserted accessory domain in yNAP-1 (yellow) is highlighted, which is missing both in PfNapL and hSET (set to scale). The brown bar is not set to scale, since the N and C termini are variable. b, structure of the PfNapL dimer generated using 2-fold crystallographic symmetry operation. Each monomer of PfNapL contains a domain I composed of dimerization helix α2 and domain II composed of a β subdomain (four antiparallel β strands) and α-helices on the other side. c, structure-based sequence alignment of PfNapL, yNAP-1, and hSET. The structures share high structural homology, except for an additional accessory domain between domain I and domain II in yNAP-1 (foreground colored yellow). Residues contributing to dimer formation are colored green. Conserved hydrophobic motifs, predicted NES, and predicted NLS in PfNapL are underlined in purple and blue, respectively. Conserved residues from the hSET mutagenesis data are colored red and blue. d, sequence alignment of PfNapL and other eukaryotic members of the NAP family: yNAP-1 (Saccharomyces cerevisiae), xNAP-1 (Xenopus laevis), dNAP-1 (Drosophila melanogaster), and hNAP-1 (Homo sapiens). The accessory domain in yNAP-1 is foreground-colored yellow. The identical/conservative surface residues and conserved hydrophobic motifs are colored pink and purple, respectively. e, phylogenetic tree of NAPs from various species showing greater evolutionary distance of malaria parasite NAPs from homology in yeast and humans (indicated by red arrows). f, model for in vitro relay of histones by P. falciparum nucleosome assembly proteins. This model was reproduced from Ref. 14, showing distinct roles for PfNapL (in cytoplasm as a histone carrier) and PfNapS (in nucleus as a histone deposit). PfNapL (L) interacts with histones and can deliver histones to PNPas (S: phosphorylated or unphosphorylated) readily. PfNapL upon phosphorylation (L*) binds 3-fold better to the histones when compared with the unphosphorylated form of PfNapL. Phospho-PfNapL may deliver its histone cargo PNPas. The latter takes over histones from Phospho-PfNapL-histone complexes and shuttles the histones into nucleus.
dimer displays a somewhat narrow cavity (Fig. S5b) when compared with Vps75 dimer (18, 19). In yNAP-1, a disulfide bond is formed by residues Cys249 and Cys272. These positions are replaced by serine and threonine in PfNapL, and only a single cysteine is present in PfNapL at position 133. The hydrophobic core of PfNapL and yNAP-1 is stabilized by the presence of SFF(T/N)FF and (L/I)P(E/S)FWL motifs that are conserved among the NAP family (Fig. 1c and Fig. S4b).

**PfNapL Localization in Parasite Cytoplasm Using Transfection Studies**—We have earlier shown that PfNapL is expressed during all of the blood stages and is localized to the parasite cytoplasm (13, 14). PfNapL also contains conserved NLS and NES motifs (Figs. 1c and 3, b and c) like the yNAP-1 (which is observed in both the cytoplasm and the nucleus). Corresponding to the presence of NES in yNAP1 (residues 88–103) (Fig. 3a), residues 35–50 of PfNapL have been predicted as an NES (Fig. 3, b and c). It has been suggested in yNAP-1 that this NES is masked by the accessory domain, which is thought to play an important role in the import and export of yNAP-1 into and from the nucleus (15) (Fig. 3a). As mentioned earlier, this accessory domain in yNAP-1 (residues 141–180) is the inserted α-helix, which is absent in PfNapL (Figs. 1c and 2a). Interestingly, hSET also lacks this accessory domain yet contains both NES and NLS motifs (16). Further, hSET is localized both in the cell cytoplasm and the nucleus, similar to yNAP-1 (36) and like other histone shuttling proteins. In this respect, PfNapL clearly stands as an exception despite its structural and sequence similarities with yeast NAP and human SET proteins.

In order to address whether PfNapL is located at some stage in the nucleus and whether its cytosolic localization is due to continuous export mediated by its putative NES, we generated cell lines expressing PfNapL-GFP chimeras. For these chimeras, we either used wild-type sequence or introduced mutations in the conserved residues of the NES (PfNapL L46A or PfNapL Q50A) (Fig. 3c). Disruption of a functional NES was predicted to lead to the accumulation of GFP chimera in the parasite nucleus. Expression of the GFP chimeras was verified via Western blot and probed with anti-GFP antibodies (Fig. 4a). A band of ~70 kDa was observed in all cell lines, corresponding to the size of PfNapL (40.5 kDa) fused to GFP (26.8 kDa). The GFP fusion protein of all three transfectants (wild type and mutants) was detectable in the cytoplasm of all stages. Previous localiza-
tion experiments with PfNapL-specific antibodies detected an exclusive cytoplasmic localization in all stages, except for the schizont stages, where the separation between nuclear (DAPI) and PfNapL staining was less defined (13). This could be due to greater interdigitation of the nucleus and cytoplasm at that stage or due to a closer association of the PfNapL pool with the nucleus. To address this question, we followed the localization of PfNapL wild type over the asexual erythrocytic life cycle (Fig. 4, b–d). PfNapL wild type-GFP was detected in the cytoplasm of the parasites. No fluorescence could be detected in the erythrocyte cytosol. Neither the food vacuole nor the nucleus of the parasite showed fluorescence in any parasite stage. This indicates that PfNapL is exclusively located in the cytoplasm of the parasite and that its putative NES is not necessary for its localization, and the predicted NES is not functional/relevant in PfNapL.

The Electrostatic Potential of PfNapL Dimer—Analysis of the electrostatic potential of PfNapL dimer indicates that the convex region of the dimerization helix α2 has acidic residues scattered on the surface in an alternate manner (Glu³⁸, Glu⁴¹, Glu⁵², Asp⁵⁵, Glu⁶³, Asp⁸⁰, and Glu⁸⁴) (Fig. 5). There are also three basic residues toward the end of this helix (Lys⁷⁰, Lys⁸¹, and Arg⁸²) contributing to a slightly basic nature and hence an uneven charge distribution (Fig. 5). The domain II of PfNapL dimer has a hydrophobic nature on one side and is highly negatively charged on the opposite face with several residues (Glu¹¹⁹, Glu¹²², Glu¹²³, Glu²⁵², Glu²⁵⁶, Asp²⁵⁸, Glu²⁶⁰, and Glu²⁶⁷), forming a cavity that is also a characteristic feature of the yNAP-1 and hSET (Fig. 5). The bottom of this cavity has an uneven charge character, and a single acidic residue, Glu⁶¹, is present at its base. It has been previously suggested for yNAP-1 that this highly acidic cavity might be important for binding to basic histones (15). The residues Asp¹¹⁹, Glu¹²², Glu¹²³, Asp²⁵⁸, Glu²⁶⁰, and Glu²⁶⁷ that constitute this cavity in PfNapL are structurally conserved with yNAP-1 and hSET, hence highlighting their possible significance.

Prediction of Essential Residues Involved in Histone Binding by PfNapL—Most histone chaperones contain long acidic stretches in their C-terminal region, which were thought to be involved in histone
binding, considering the highly basic character of histones. However, recent studies have shown that these acidic stretches are not essential for histone binding activities for chaperones yNAP-1, Asf1, nucleoplasmin, etc. (5, 15, 37). It is noted that full-length PfNapL also has an acidic stretch of residues in its C-terminal region similar to yNAP-1 and hSET, with an average pI of ~3.0 calculated using amino acid sequence (Fig. 1a).

Mutagenesis studies on hSET revealed important residues of domain II that affect the binding of hSET to both core histones and double-stranded DNA (16). We have mapped these residues on yNAP-1 and PfNapL (Fig. 6a). Based upon these hSET mutagenesis data, we identified several corresponding residues in PfNapL and mutated them to test their relevance in histone recognition. We constructed six single-site mutants of the corresponding residues from PfNapL (D192A, H227A, T230A, Y259A, E260A, and K266A). In addition, we made one other mutant (D223A) that is conserved, exposed, and proximal to the previously implicated histone recognition residues from hSET (16). Our PfNapL-histone binding data, using purified proteins of high quality, show that none of these seven mutations altered histone binding of PfNapL to histones H3, H4, H2A, or H2B, to histone tetramer, or to histone octamer significantly (Fig. 6b). We therefore propose that it is likely for histones to have different a binding site(s) on PfNapL (and possibly on other NAPs) in comparison with hSET-histone interactions (16). These residues are not identical among PfNapL, hSET, yNAP-1, and Vps75 proteins, and in general, residues proposed by hSET-histone studies show weak conservation (Table S1). An overall structure-based residue comparison between yNAP-1, Vps75, hSET, and PfNapL reveals very few regions of sequence conservation (Fig. S1). In order to further probe the potential sites for histone recognition by PfNapL, we inspected the surface residues of PfNapL (keeping in view structure and sequence information from NAPs). Here, we have highlighted the identical/conserved surface residues in the NAP family (Figs. 1d and 7) based on the described criterion.

Comparisons with Asf1-Histone Complex—The β subdomain of PfNapL is a highly conserved feature of NAPs and is also found in other histone chaperones like Asf1 (20). The crystal structure of the human Asf1 has been recently determined in complex with H3-H4 dimer, highlighting the crucial residues involved in histone binding (20, 21). A structural comparison of the four-stranded β subdomain of PfNapL with Asf1 shows a superimposition with an r.m.s. deviation of 0.97 Å (Fig. 8a). Further, there are no steric hindrances between the overall PfNapL dimer and H3-H4 dimer (Fig. S6). Mapping of the interacting residues as highlighted for human Asf1 revealed three corresponding, structurally conserved surface residues in PfNapL (Ile136, Ile147, and Ala185) (Fig. 8b). Further, three residues of PfNapL are conserved among the NAP family (Fig. 8c) and may represent common sites for histone recognition. Based on structural congruence, we propose that these residues in PfNapL may interact with Leu126 and Ile130 of H3 and Thr96 and Leu97 of histone H4. To validate the above modeling, we mutated the putative interacting PfNapL residues, Ile136 and Ile147, to alanines and tested the mutant proteins for H3-H4 tetramer binding ability. Our protein-protein-based ELISA data indicate a significant (up to two-thirds) reduction in binding between PfNapL and H3-H4 tetramer (Fig. 8d).

DISCUSSION

The structure of PfNapL was determined using the iodide-SAD technique and refined to 2.3 Å resolution. Structural analysis of PfNapL has revealed that PfNapL conforms to the domain architecture of nucleosome assembly proteins, exhibiting a high complexity ordered central region and largely disordered N- and C-terminal regions. PfNapL forms a homodimer that acquires a fold similar to the yNAP-1, hSET, and Vps75 dimers (15–17). These investigations demonstrate high structural homology within these proteins despite a low level of sequence identity (only 14–25%) (Fig. 1c). The major difference between PfNapL and yNAP-1 lies in the absence of
the accessory domain in PfNapL that is inserted between the dimerization helix α2 and domain II in yNAP-1 (15) (Fig. 1a). Phylogenetic analysis of NAPs suggests greater evolutionary distance of PfNapL from homologs in higher eukaryotes (Fig. 1e), although malaria parasite NAPs are unique in their lack of the accessory domain. We have earlier shown that PfNapL is a unique, nonredundant protein in P. falciparum, and it performs an exclusive role in the nucleosome assembly activity in the parasite (13, 14). Previous studies on gene knock-out phenotypes in several organisms like mouse and Drosophila have indicated the essentiality of the NAP family (9, 10). Furthermore, our inability to disrupt the gene for PfNapL corroborates the essentiality of this protein in P. falciparum (Fig. S2).

Corresponding to yNAP-1, PfNapL also contains NES and NLS motifs (Figs. 1c and 3c). We have previously shown that PfNapL is predominantly localized to the cytoplasm and is centrally responsible for shuttling and transfer of histones to PfNapS (which is localized to both cytoplasm and the nucleus and which eventually deposits histones onto DNA). PfNapL and PfNapS do not interact with each other (14). Therefore, PfNapL is principally localized to the cytoplasm and does not possess the ability to deposit the histones in the nucleus (14). In this context, it is interesting to note that hSET also lacks the accessory domain of yNAP-1/other NAPs, but unlike PfNapL, hSET is localized both in cytoplasm and the nucleus (22). Since this accessory domain is absent in PfNapL (Fig. 2b), the predicted NES on the PfNapL structure is thus exposed and not be masked by the accessory domain, as suggested and observed for yNAP-1 (15) (Fig. 3, a and b). From a structural perspective, this further validates the localization of PfNapL to the cytoplasm. The histone binding characteristics of PfNapL have been detailed previously, including its ability to transfer cytoplasmic histones on to PfNapS. A model for relay of histones from parasite cytoplasm to the nucleus has also been proposed by us previously (see Ref. 14 and specifically Fig. 1f reproduced here from Ref. 14). Our published reports on the two Plasmodium parasite nucleosome assembly proteins have clearly suggested different
roles for PfNapL (in histone shuttling in the cytoplasm) and for PfNapS (which is localized both in the cytoplasm and nucleus and which can perform histone deposition onto DNA (13, 14) (Fig. 1f).

Electrostatic potential distribution of PfNapL dimer reveals a stretch of alternate acidic residues on the convex surface of the dimerization helix $\alpha_2$ and a highly acidic side to domain II. If this convex surface of the PfNapL dimer is implicated in histone recognition, then it can be suggested that the highlighted acidic residues on this surface might be making alternate contacts with an $\alpha$-helix of a core histone (Fig. 5). In yNAP-1, the acidic stretch forming a cavity in domain II has been shown to be involved in neutralizing and binding the basic N-terminal histone tails (15). Also, most of the acidic residues constituting this cavity are structurally conserved among PfNapL and yNAP-1. Thus, based upon the earlier suggestions for yNAP-1, this acidic region in PfNapL may be involved in histone recognition (Fig. 5).

Mapping of the corresponding residues from hSET mutagenesis data onto PfNapL and yNAP-1 demonstrates the conserved nature of 10 of these residues among PfNapL, hSET, and yNAP-1 (Fig. 6a). However, mutation of some of these residues on PfNapL did not significantly alter PfNapL-histone binding (Fig. 6b). Consequently, based upon our mutagenesis data analysis, we suggest that the binding site(s) and/or essential residues for the recognition of core histones may be different for PfNapL from those proposed by hSET-histone interactions (16). A structure-based comparison of the residues that were mutated in PfNapL reveals that none of these are identical among hSET, yNAP-1, Vps75, and PfNapL. Indeed, only a few show weak conservation (Table S1). Also, an overall structure-based comparison between yNAP-1, Vps75, hSET, and PfNapL reveals very few regions of sequence conservation (Fig. S1). These observations therefore suggest that histone recognition site(s) for hSET may not be applicable for nucleosome assembly proteins like PfNapL and yNAP-1. Our mapping of highly conserved and surface-exposed residues in the NAP family provides some clues for the possible set of essential residues that might underpin NAP-histone binding (Figs. 1d and 7). Several of these conserved residues lie in and around the characteristic cavity formed by the PfNapL dimer, which has been previously implicated in histone recognition in both yNAP-1 and hSET (15, 16). Distribution of these conserved surface residues on the convex region of PfNapL dimer illustrates their proximity to and overlap with the alternate stretch of acidic residues highlighted by our electrostatic potential analysis.
It is known that the β subdomain present in yNAP-1, hSET, and PfNapL is a highly conserved feature of histone chaperones (Fig. 2a). The superimposition of the β subdomain of PfNapL onto histone chaperone Asf1 revealed three conserved surface residues on PfNapL (conserved in all NAPs) that are suggested to be important for interaction with H3-H4 dimer based upon the Asf1 and H3-H4 dimer interactions (21) (Fig. 8, a–d, and Fig. S6). Interestingly, the three residues identified in Asf-histone complex map to a region of PfNapL that is not implicated by either hSET mutagenesis or by our mapping of conserved, exposed residues in NAPs. This suggests that Asf1 and NAPs may recognize histones differently. Further, modeling of PfNapL based on the Asf1-histone complex crystal structure clearly suggests three target residues that are likely to participate in H3-H4 recognition. Based on these structural insights, we mutated two of the three residues (the third one is an alanine in any case) and confirmed the idea that their alteration to alanine reduces binding of PfNapL to H3-H4 tetramer significantly (Fig. 8d).

In summary, our studies provide further structural insights into histone chaperones from *P. falciparum* and highlight key regions of differences between plasmodial NAPs and their counterparts. Although further dissection of the modes of histone recognition by NAPs will be required to gain a complete understanding of the mechanism of histone binding and transport by nucleosome assembly proteins, it is already clear that the parasite NAPs are sufficiently different from their counterparts in other species. Our genetic-structural-functional data...
Structure of Malaria Parasite Nucleosome Assembly Protein

The structure of the Malaria parasite nucleosome assembly protein 1 (NP1) was determined by X-ray crystallography. NP1 is a highly conserved protein found in all eukaryotic organisms and plays a critical role in nucleosome assembly. The structure of NP1 revealed a novel fold, with two distinct RNA helicase domains that are involved in nucleosome assembly.

Acknowledgment—We thank Prof. Alan Cowman for support and for the generous gift of reagents.

REFERENCES

Crystal Structure of Malaria Parasite Nucleosome Assembly Protein: DISTINCT MODES OF PROTEIN LOCALIZATION AND HISTONE RECOGNITION
Jasmita Gill, Manickam Yogavel, Anuj Kumar, Hassan Belrhali, S. K. Jain, Melanie Rug, Monica Brown, Alexander G. Maier and Amit Sharma

doi: 10.1074/jbc.M808633200 originally published online January 27, 2009

Access the most updated version of this article at doi: 10.1074/jbc.M808633200

Alerts:
• When this article is cited
• When a correction for this article is posted

Click here to choose from all of JBC's e-mail alerts

Supplemental material:
http://www.jbc.org/content/suppl/2009/01/28/M808633200.DC1

This article cites 37 references, 9 of which can be accessed free at
http://www.jbc.org/content/284/15/10076.full.html#ref-list-1