Cyanochromes Are Blue/Green Light Photoreversible Photoreceptors Defined by a Stable Double Cysteine Linkage to a Phycoviologbilin-type Chromophore

Received for publication, June 29, 2009, and in revised form, August 3, 2009 Published, JBC Papers in Press, August 11, 2009, DOI 10.1074/jbc.M109.038513

Andrew T. Ulijasz¹, Gabriel Cornilescu⁵, David von Stetten⁵, Claudia Cornilescu⁵, Francisco Velazquez Escobar⁴, Junrui Zhang⁴, Robert J. Stankey⁴, Mario Rivera⁴, Peter Hildebrandt⁴, and Richard D. Vierstra⁴²

From the ¹Department of Genetics, University of Wisconsin, and the ⁵National Magnetic Resonance Facility at Madison, Madison, Wisconsin 53706, the ⁶Technische Universität Berlin, D-10623 Berlin, Germany, and the ¹Department of Chemistry, University of Kansas, Lawrence, Kansas 66045

Phytochromes are a collection of bilin-containing photoreceptors that regulate a diverse array of processes in microorganisms and plants through photoconversion between two stable states, a red light-absorbing Pr form, and a far red light-absorbing Pfr form. Recently, a novel set of phytochrome-like chromoproteins was discovered in cyanobacteria, designated here as cyanochromes, that instead photoconvert between stable blue and green light-absorbing forms Pb and Pg, respectively. Here, we show that the distinctive absorption properties of cyanochromes are facilitated through the binding of phycocyanobilin via two stable cysteine-based thioether linkages within the cGMP phosphodiesterase/adenyl cyclase/FhlA domain. Absorption, resonance Raman and infrared spectroscopy, and molecular modeling of the Tc-Pixj GAF (cGMP phosphodiesterase/adenyl cyclase/FhlA) domain associated with phycocyanobilin are consistent with attachments to the C₃ carbon of the ethylenedioxy side chain and the C₄ or C₅ carbons in the A–B methine bridge to generate a double thioether-linked phycociologbilin-type chromophore. These spectroscopic methods combined with NMR data show that the bilin is fully protonated in the Pb and Pg states and that numerous conformation changes occur during Pb → Pg photoconversion. Also identified were a number of photochromically inactive mutants with strong yellow or red fluorescence that may be useful for fluorescence-based cell biological assays. Phylogenetic analyses detected cyanochromes capable of different signaling outputs in a wide range of cyanobacterial species. One unusual case is the Synecocystis cyanochrome EtrI that also binds ethylene, suggesting that it works as a hybrid receptor to simultaneously integrate light and hormone signals.

Phytochromes (Phys)³ comprise a large and diverse superfamily of photoreceptors that regulate a wide range of physiological responses in plants, fungi, bacteria, and cyanobacteria (1–3). They are unique among photoreceptors by being able to photoconvert between two stable states, a red light-absorbing Pr form that is typically the dark-adapted and biologically inactive conformer and a far-red light-absorbing Pfr form that requires light for its production and is typically the biologically active conformer. By interconverting between Pr and Pfr, Phys act as light-regulated switches in controlling processes ranging from phototaxis and pigmentation in bacteria to seed germination, photomorphogenesis, and flowering time in higher plants.

Light absorption by Phys is directed by a bilin (or linear tetrapyrrole) chromophore produced by the oxidative cleavage of heme. Although bacterial and fungal Phys use the immediate cleavage product biliverdin (BV), cyanobacterial and higher plant Phys use phycocyanobilin (PCB) and phytochromobilin, respectively, produced by enzymatic reduction of BV (1, 2). The bilin is then covalently bound autocatalytically to the photosensory unit of the apoprotein, which typically contains a sequence of Per/Arndt/Sim (PAS), cGMP phosphodiesterase/adenyl cyclase/FhlA (GAF), and Phy-associated (PHY) domains. Intimate contact between the bilin and surrounding protein residues then generates the unique photochromic properties of Phys. Recent three-dimensional structures of the Pr form of several bacterial Phys (BphPs) and two cyanobacterial Phys (Cphs) have shown that the bilin is deeply buried within the GAF domain in a ZZZssa configuration and that the connection between the GAF and PAS domains is stabilized by a rare figure-of-eight knot involving the region upstream of the PAS domain being lassoed by a conserved loop within the GAF domain (4–9). Although the structure of Pfr remains unsolved, various physicochemical studies have proposed that photoconversion involves a rotation of one of the three methine bridges between the pyrrole rings (1, 10–14). This rotation then

Pb, blue-light-absorbing form of cyanochrome; Pg, green light-absorbing form of cyanochrome; Pfr, far-red-light-absorbing form of Phy; Pr, red light-absorbing form of Phy; BV, biliverdin IXα; Cyc, cyanochrome; GAF, cGMP phosphodiesterase/adenyl cyclase/FhlA; HK, histidine kinase; HSQC, heteronuclear single quantum coherence; IR, infrared; PAS, Per/Arndt/Sim; PCB, phycocyanobilin; PVB, phycoviologbilin; PEI, phycocerythrobilin; PUB, phycourobilin; ALA, α-aminolevulinic acid; RR, resonance Raman; Ni-NTA, nickel-nitritriacetic acid; SyB, Synechococcus sp. OS-B; Syn, Synecocystis sp. PCC6803; Te, Thermosynecococcus elongatus BP-1; Dr, Deinococcus radiodurans.

© 2009 by The American Society for Biochemistry and Molecular Biology, Inc. Printed in the U.S.A.
Double Bilin Linkage in Cyanochromes

induces much slower thermally driven movements of the protein to initiate signal transduction.

In microorganisms, Pfr can activate a variety of signaling systems using output motifs directly appended to the C-terminal end of the photosensory region. The most prevalent are histidine kinase domains that then begin specific two-component phosphorelay (3, 15, 16). Although the output of plant Phys remains unclear, the presence of a C-terminal HK-related domain suggests that they also work as light-regulated protein kinases (17).

In addition to the canonical Phys, it has become apparent through phylogenetic and biochemical studies that a heterogeneous collection of Phy-like photoreceptors exists (e.g. Refs. 3 and 18). These include Phys that prefer Pfr as the dark-adapted state (7, 19, 20), Phys that photoconvert from Pr to shorter wavelength-absorbing “near red” or Prn forms (6, 21), and Phy-like photoreceptors that bind bilins but instead photoconvert between forms with maximal absorption other than red and far-red light (22–25). Often these Phy-like sequences are missing key residues or domains common among canonical Phys, suggesting that they employ novel bilins as chromophores, bind the bilin in different architectures, and/or use distinct photochemistries.

One subclass of novel Phy-like photoreceptors present in a number of cyanobacteria, which we have designated cyanochromes (or Cycs) to better distinguish them from Cphs, is exemplified by Synecocystis sp. PCC6803 (Syn PixJ) (or TaxD1, locus sll0041) and its relatives. Syn-PixJ was discovered based on its involvement in blue light-mediated phototaxis in this mesophilic cyanobacterium (26, 27) with its close homolog Te-PixJ (locus tll0569) then found in the thermophilic cyanobacterium Thermosynechococcus elongatus BP-1 by sequence similarity (28). Like Cphs, the cyanochromes tested thus far covalently bind PCB but then generate photoreceptors that convert between blue and green light-absorbing forms designated Pb and Pg, respectively (22, 24, 29). Subsequent studies proposed that PCB is converted to phycoviolobilin (PVB) upon attachment to the apoprotein (30). PVB differs from PCB by having a methylene instead of a methine bridge between the A and B pyrrole rings, which blue-shifts the absorption of the chromophore by shortening the π-conjugation system. Phototransformation of Pb to Pg could then occur by a mechanism similar to Phys.

How Te-PixJ and related cyanochromes bind PCB to generate more blue-shifted PVB-type chromophores remains unclear. Like Cphs, two cyanochromes examples link PCB via a thioether linkage between a cysteine in the Cyc-GAF domain and the C3' carbon of the ethyliendene side chain of ring A (24, 28). Additionally, loss of the C4=C5 double bond is necessary to generate PVB. One model by Ishizuka et al. (30) from studies with Te-PixJ proposed that the double bond moves from the C4-C5 position to the C2-C3 position by an autoisomerase activity intrinsic to the GAF domain. A more recent model by Rockwell et al. (24) using another Syn-PixJ relative in T. elongatus, Tlr0924, invoked the possibility of a second cysteine that also participates in PCB ligation. This cysteine was proposed to bind the bilin at the C10 position via a reversible thioether linkage. In the dark-adapted Pb state, the second linkage would then be formed to generate a rubin-like chromophore attached to the bridge between the B and C pyrrole rings. This bond would then break upon photoconversion to generate the more π-conjugated green light-absorbing photoprdut Pg.

In this report, we employed a number of physicochemical approaches to help resolve the unique chromophore architecture and photochemical properties of cyanochromes, using Te-PixJ as the example. By independently mutagenizing the cysteine that binds the A ring ethyliendene (Cys-522 (22)) and that proposed by Rockwell et al. (24) to reversibly bind the bilin at a second site (Cys-494), we demonstrate that both residues form light-stable covalent adducts with a PVB-type chromophore. In addition, we employed various spectroscopic methods to show that the bound PVB is fully protonated as both Pb and Pg, that only one pyrrole ring is active during photoconversion, and that the polypeptide may undergo extensive remodeling as Pb converts to Pg. We identified a set of conserved amino acids in Te-PixJ important for cyanochrome photochemistry, including several that when substituted generate yellow or red fluorescent chromoproteins potentially useful for cell biological applications. Phylogenetic analyses show that cyanochromes are widespread among cyanobacteria with their closest relatives being members of the red/far-red light-absorbing Phy subfamily defined by the absence of the N-terminal PAS domain (31).

EXPERIMENTAL PROCEDURES

Te-PixJ Cyc-GAF Domain Alignment and Secondary and Tertiary Structure Predictions—Phy/Cyc sequences were identified by performing BLAST searches of the GenBank™ data base (www.ncbi.nlm.nih.gov) using the Te-PixJ Cyc-GAF domain sequence as the query. Amino acid sequence alignments were generated with ClustalW (32) and displayed using MacBoxShade version 2.15 (Institute of Animal Health, Pirbright, UK). Secondary structure was predicted using the Scratch Protein Predictor (SSpro) (33, 34). The tertiary structure of Te-PixJ(Cyc-GAF) was predicted using the SWISS-MODEL online resource (35) by inputting an alignment of the SyB-Chp 1 GAF domain (Leu-31 to Gin-200) and Te-PixJ Cyc-GAF domain (Arg-440 to Phe-585) sequences along with the three-dimensional structure of SyB-Chp1(GAF) as the molecular template (Protein Data Bank (PDB) ID code 2K2N (8)). Representations of the predicted Te-PixJ (Cyc-GAF) structure were generated with PyMOL (DeLano Scientific LLC, San Carlos, CA).

Phylogenetic Analysis—Phy, Cyc and related sequences were obtained from the National Center for Biotechnology Information (NCBI) and the Joint Genomes Institute (JGI) data bases and are listed with their accession numbers in supplemental Table 1. Sequences were aligned using CLUSTALX (36) under default settings and trimmed to include only the GAF domain (supplemental Table 2). Phylogenetic analysis was performed with MrBayes version 3.1.2 (37) using the proportion of invariable sites γ rate model and the mixed amino acid model. The software was run for 1,350,000 generations, discarding the first 25% (337,500 generations) as “burn-in” after checking that the log likelihood values had plateaued. The strict consensus tree was displayed with TreeView (38). A similar tree architecture...
Double Bilin Linkage in Cyanochromes

Expression of Recombinant Cyc-GAF Domains—The coding regions for the Cyc-GAF domain from Te-PixJ (residues 430–591 from locus slr1212) and Syn-Etr1 (residues 466–624 from locus slr1212) were PCR-amplified from T. elongatus BP-1 genomic DNA and the Synechocystis phosphor YpCDE (40), respectively, and cloned into Ncol/HindIII-digested pBAD-6H expression plasmid using the melt and reanneal method as described (31). Te-PixJ(Cyc-GAF) mutants were generated by PCR as described (31), using two complementary primers containing the desired nucleotide change. Primers used in this study are shown in supplemental Table 3.

Recombinant GAF domain expression and purification were performed as described previously (31) using a modification of the dual plasmid expression system devised by Gambetta and Lagarias (41). For in vitro assembly, Escherichia coli crude extracts containing the apoprotein were incubated with 200 μM interference filters, respectively. For denaturation experiments, which were dialyzed into 25 mM Tris-HCl (pH 8.0) and 1 mM EDTA. Samples for NMR analysis were added to the minimal expression medium as described (31).

To isotopically label PCB with 13C at carbons 4, 5, 9, 10, 15, and 19 prior to assembly with the Te-PixJ(Cyc-GAF) apoprotein, 100 μM 5-[^13C]aminolevulinic acid (ALA) was added to the minimal expression medium as described (31). Incorporation of 15N into PCB was similarly accomplished by adding a final concentration of 100 μM[15N]ALA (Medical Isotopes Inc., Pelham, NH). Following a 2-h induction of the E. coli BL21-AI cultures with 1 mM isopropyl 1-thio-β-D-galactopyranoside and an overnight incubation at 18 °C with 0.2% arabinose, the expressed proteins were purified by nickel nitrilotriacetic acid agarose (Ni-NTA) chromatography (Qiagen, Valencia, CA), using 300 mM imidazole for elution. Most eluates were then dialyzed into 25 mM Tris-HCl (pH 8.0) using Amicon Ultra-15 centrifugal filters (10 kDa cutoff, Millipore, Billerica, MA), with the exception of samples used for resonance Raman (RR) or IR spectroscopy, which were dialyzed into 25 mM Tris-HCl (pH 8.0) and 1 mM EDTA. Samples for NMR analysis were further purified by fast protein liquid chromatography with a phenyl-HP column (GE Healthcare) (31), pooled, and then dialyzed into 10 mM Tris-DCI (pH 8.0). 5-[^13C]ALA-labeled Te-PixJ(GAF) was adjusted to a final concentration of 1.3 mM for IR and RR spectroscopy and a final concentration of 1.75 mM for NMR spectroscopy. The [15N]ALA-labeled Te-PixJ(GAF) samples were brought to a concentration of 1.5 mM before NMR analysis.

Absorption and Fluorescence Spectroscopy—Absorption spectra were measured with a PerkinElmer Lambda 650 UV-visible spectrometer (PerkinElmer Life Sciences) with the samples dissolved in 25 mM Tris-HCl (pH 8.0). Pb → Pg and Pg → Pb photoconversions were achieved with white light filtered through appropriate 430- and 530-nm interference filters, respectively. Pr → Pfr and Pfr → Pr photoconversions were achieved with white light filtered through 630- and 690-nm interference filters, respectively. For denaturation experiments, the chromoproteins were either photoconverted with white light and the appropriate interference filter (430 nm for Pb and 630 nm for Pr) or kept in darkness followed by denaturation in 8 M urea (pH 2.0). Soluble and bound chromophores were separated following 10% trichloroacetic acid precipitation. The precipitated fraction was redissolved in acidic urea. Following UV-visible absorption measurements, denatured samples were then irradiated with the appropriate wavelength of light corresponding to their Pb/Pr and Pg/Pfr peak absorbance as follows: wild-type Te-PixJ(Cyc-GAF) (590 and 530 nm), Te-PixJ(Cyc-GAF) D494H (590 and 660 nm), Te-PixJ(Cyc-GAF) C522A (590 or 690 nm), Te-PixJ(Cyc-GAF) C494A, and SyB-Cph1(GAF) (590 and 660 nm).

Fluorescence excitation and emission spectra were recorded with a QuantaMaster model C-60/2000 spectrofluorometer (Photon Technologies International, Birmingham, NJ). Emission spectra were recorded with an excitation of 523–555 nm, depending on the sample. Wild-type Te-PixJ(Cyc-GAF) and the Y463H, D492H, and C522A mutants were excited with 360 nm light, and the C494H and C494A mutants were excited with 590 nm light. Excitation spectra were recorded with 551–651-nm emission wavelengths, depending on the sample.

IR Spectroscopy—IR spectra were recorded with 1064 nm excitation (Nd-YAG continuous wave laser, line width <1 cm⁻¹) using a RFS 100/S (Bruker Optics, Ettlingen, Germany) Fourier transform Raman spectrometer (4 cm⁻¹ spectral resolution). The near IR excitation line was sufficiently close to the first electronic transition to generate a strong preresonance enhancement of the chromophoric vibrational bands such that Raman bands of the protein matrix remained very weak in the spectra of the parent states (42–44). Typical sample volumes were 5 μl of a 1–2 mM protein solution in 25 mM Tris-HCl (pH 8.0) and 1 mM EDTA. All spectra were measured at −140 °C using a liquid-nitrogen cooled cryostat (Linkam, Waterfield, Surrey, UK). The laser power at the sample was set at ∼700 milliwatts, which did not damage the chromoproteins as checked by comparing spectra of the samples obtained before and after RR data acquisition. The total accumulation time was less than 2 h for each spectrum. For all RR spectra, the background was subtracted manually. Calculated RR and IR spectra were obtained by density functional theory using the B3LYP functional and the 6–31G* basis set. The force field was scaled by a set of global scaling factors as described elsewhere (13, 45).

To detect photoproducts that accumulate in blue light, the samples were irradiated at room temperature and then rapidly frozen to −140 °C. Green light-emitting diodes combined with a blue interference filter were used for photoconversion in the RR and IR experiments.

IR Spectroscopy—IR difference spectra were recorded at 20 °C in a Bruker Tensor IR interferometer equipped with an attenuated total reflection device. Typical sample volumes were 20 μl of a 1–2 mM protein solution in 25 mM Tris-HCl (pH 8.0) and 1 mM EDTA. Composite spectra were assembled from 200 scans (−90 s each); these spectra were recorded in batches of five separated alternately by saturating green and blue light irradiations. Difference spectra from 10–15 blue/green irradiation cycles were generated and added together without backround correction.
NMR Spectroscopy—Isotopically labeled forms of Te-PixJ-(Cyc-GAF) assembled with PCB (~2 mM) were dissolved in 93% H₂O, 7% D₂O, 10 mM deuterated Tris-DCl (pH 8.0), and 0.15 mM NaN₃, and placed in a 280-μl Shigemi microcell. NMR spectra were collected at 25 °C using an 800-mHz ¹H frequency Varian INOVA spectrometer (Varian Inc., Palo Alto, CA).
Double Bilin Linkage in Cyanochromes

equipped with a cryogenic probe. Pg samples were obtained by irradiating the microcell solution with saturating blue light (430 nm). Photoconversion of Pg → Pb was accomplished by irradiating the microcell with saturating green light (530 nm).

1H-15N heteronuclear single quantum coherence (HSQC) spectra with 15N-13C-labeled Te-Pix(J(Cyc-GAF) chromoprotein or unlabeled Te-Pix(J(Cyc-GAF) protein assembled with [15N]PCB were collected as 128*(t1, 15N) × 1022* (t2, 1H) data matrices. Acquisition times were 48 and 85 ms in the t1 and t2 dimensions, respectively. 1H-13C HSQC spectra, which centered on the methine region of unlabeled Te-Pix(J(Cyc-GAF) protein assembled with [13C]PCB, were collected as 32*(t1, 13C) × 769* (t2, 1H) data matrices. Acquisition times were 10 and 80 ms in the t1 and t2 dimensions, respectively. 13C direct detected spectra were acquired with 51,200 scans and a total experimental time of 23 h each. Data were processed and plotted using the NMRPipe software package (46). All NMR data were collected at the National Institutes of Health-sponsored National Magnetic Resonance Facility at Madison (NMRFAM) at the University of Wisconsin.

RESULTS

Architecture and Phylogeny of Cyanochromes—Pix(J) type biliproteins represent a novel class of Phy-type photoreceptors that photoconvert between blue (Pb) and green light-absorbing (Pg) forms (22, 28). To better define their distribution, we used the Cyc-GAF domain sequences of Syn-Pix(J) (TaxD1) and Te-Pix(J) to search the various genome data bases for potential homologs. This search retrieved numerous relatives in a variety of cyanobacteria, including T. elongatus, Synechocystis, Cyanotheca, Acaryochloris marina, Gloeobacter violaceus, Anabaena variabilis, and Anabaena (Nostoc) species (Fig. 1B and supplemental Table 1). Previously known Phy-type proteins that also clustered in this group included: Synechococcus elongatus CikA (involved in circadian entrainment (47)) and Synechocystis Etr1 (implicated in ethylene perception (40)), suggesting that these proteins are photochemically related to Pix(J). No Pix(J)-type sequences were found in any species outside of cyanobacteria, in agreement with the prediction that this photoreceptor type is restricted to the cyanobacterial kingdom (24, 29). Consequently, we have used the term cyanochromes (or Cycs) to name this unique group to better distinguish them from the more typical Cphs that are present together with Cycs in many cyanobacterial species.

When the obtained GAF domain sequences were analyzed using a Bayesian phylogenetic comparison, the resultant cyanochrome clade was clearly separate from others in the Phy superfamily, including BphPs, Cphs, diatom Phys (Dphs), fungal Phys (Fphs), and plant Phys (supplemental Fig. 1). Maximum likelihood-generated trees produced a similar result (data not shown). Their closest relatives appear to be the PAS-less Phys, the founding member of which is Synechococcus OS Type B’ (SyB)-Cph1 (8, 31). Members of this clade are missing the PAS domain that is upstream of the chromophore-binding GAF domain in other canonical Phys.

Alignment of the GAF sequence from various Phy types revealed a distinct organization for the cyanochrome family (Fig. 1B and supplemental Table 2). Most notable in cyanochromes is the absence of the ~23-amino acid region between the predicted β strand and the α3 helix in the Cyc-GAF sequences, which forms the lasso loop in the knot of canonical BphPs, Cphs, Fphs, and plant Phys (4). The DIP motif that participates in the hydrogen-bond network involving the pyrrole water in canonical Phys is also absent. Instead, we (like Rockwell et al. (24)) often found a DPCF or related ETCF motif at this position. Despite this divergence, several residues known to make important contacts with the bilin in canonical Phys are conserved in cyanochromes. These include the cysteine (Cys-522 in Te-Pix(J)) that likely binds PCB through the C31 carbon on the A ring ethylidene side chain, two histidine residues (His-523 and His-553) that bind the pyrrole water and the D ring, respectively, a tyrosine (Tyr-463) that is in close proximity to D ring and important for photoconversion, and an arginine (Arg-507) that forms a double salt bridge with the B-ring propionic acid side chain of PCB (Fig. 1B) (4, 5, 24, 28), suggesting that cyanochromes interact non-covalently with the bilin in a manner similar to other Phys.

Like other Phy types (2, 3, 20, 31, 48), the cyanochromes have widely varied domain architectures N- and C-terminal to their signature Cyc-GAF domains (Fig. 1A) (29). Examples include Te-Pix(J), which has two predicted transmembrane domains at the N-terminal end, two HK/adenyl cyclase/methyl-binding protein-phosphatase (HAMP) domains bracketing the Cyc-GAF domain, and a methyl-accepting (MA) domain at the C-terminal end, and Syn-Etr1, which has a predicted ethylene-binding domain between two transmembrane domains followed by two PAS domains and a typical HK domain at the C-terminal end. Cyanochromes are notable among the Phy superfamily for not having an obvious PHY domain following the Cyc-GAF domain. In agreement with their predicted phylogenetic relationship to PAS-less Phys based on the GAF domain, the conserved knot lasso loop present in canonical Phys located the conserved knot lasso loop present in canonical Phys.

![FIGURE 1. Protein organization and GAF domain amino acid sequence alignment of representative Cys.](http://www.jbc.org/)

A domain architectures of representative Cycs as compared with the Cph Syn-Cph1 and the PAS-less Phy Syb-Cph1. EBD, ethylene-binding; HAMP, histidine kinase/adenyl cyclase/methyl-binding protein/phosphatase; MA, methyl-accepting; TM, predicted transmembrane. The motifs containing the cysteine(s) that bind(s) the bilin and the histidine in the HK domain that participates in phototransduction are indicated. B, alignment of the GAF domain from representative Cycs (brackets) with members of the BphP (Dr-BphP), Cph (Syn-Cph1), PAS-less Phy (Syb-Cph1), and plant Phy (At-PhyA) clades. The predicted α-helical and β strand secondary structures based on the Dr-BphP and Syb-Cph1 three-dimensional structures (5, 8) are shown above the alignment. The bracket locates the conserved knot lasso loop present in canonical Phys. The dashed line locates the Cyc-specific motif bearing the second bilin attachment site. The two cysteines (Cys-494 and Cys-522) that bind the bilin are indicated by the closed arrowheads. Other key residues, some of which impact the spectral properties of Te-Pix(J), are indicated by the open arrowheads. The amino acid numbering refers to the Te-Pix(J) sequence. Sequences in the alignment include: Te-Pix(J) from T. elongatus BP-1 (T101569), Syn-Pix(J) (TaxD1) from Synechocystis sp. PCC6803 (SL0041), ZP_01728723 from Cyanotheca sp. CCOY1018, AM1_0046 from A. marina (MBIC11017), G13432 from G. violaceus PCC7421, Syn-Etr1 from Synechocystis sp. PCC6803 (SL1212), Te-CikA from T. elongatus BP-1 (T101569), Syn-CikA from S. elongatus (slr1969), Ana-CikA from A. variabilis ATCC 29413 (Ava-1168), An(Nos)-Pix from Anabena (Nostoc) sp. PCC7120 (Ava11069), RcaE from F. diplosiphon (AA085875), Syb-Cph1 from Synechococcus sp. JA-2-3B a2(13) (CYB_2465), Dr-BphP from D. radiodurans R1 (DR0050), Syn-Cph1 from Synechocystis sp. PCC6803 (Slr0473), and At-PhyA from A. thaliana Columbia-0 ecotype (At1g09570). The accession numbers or locus tags are in parentheses.
domain sequence, some members of the cyanochrome family also do not have an obvious PAS domain N-terminal to the Cyc-GAF domain. Consequently, the photosensory domain of some cyanochrome members may be the smallest known within the Phy superfamily, with only their Cyc-GAF domain potentially required for full blue/green light-driven photochromicity.

**Te-PixJ Binds PCB via a Stable Double Cysteine Linkage**—To examine the photochemistry of cyanochromes, we used a recombinant assembly protocol in which the Cyc-GAF domain from *Te-PixJ* was expressed in *E. coli* with a C-terminal His₆ tag and either assembled *in vivo* with purified bilins or assembled *in vitro* by co-expression with bilin synthetic enzymes (16, 31, 41). Bilin attachment was then assessed by zinc-induced fluorescence of the purified chromoproteins following SDS-PAGE. As shown in Fig. 2A, purified *Te-PixJ*(Cyc-GAF) showed a strong preference for PCB and failed to bind BV or the bilin precursor protoporphyrin IXα *in vitro*. However, PCB binding was poor, suggesting that its incorporation is inefficient post-translationally. In contrast, robust PCB incorporation was evident *in vivo* co-synthesis of *Te-PixJ*(Cyc-GAF) with the heme oxygenase and PCB synthase activities that sequentially convert heme into PCB (Fig. 2B). Like previous studies (22, 24, 28), we found that the recombinant wild-type Cyc-GAF domain assembled with PCB displayed a blue absorption maximum at 430 nm for the dark-adapted Pb state, which could then be photoconverted by blue light to the Pg state with an absorption maximum at 534 nm (Fig. 2C).

In agreement with Ishizuka *et al.* (28), we found that Cys-522 is important for bilin attachment as its replacement with alanine substantially diminished PCB incorporation. However, PCB still bound (albeit poorly) with the assembled chromoprotein acquiring a new absorption maximum at ~600 nm in addition to the blue light maximum at 430 nm (Fig. 2, B and C). It also remained photochromic with blue light increasing its absorbance at 600 nm and shifting the blue light maximum to 370 nm. Taken together, the data implied that a second PCB-ligation site exists.

To locate other residues that could generate a second thioether linkage, we scanned our sequence alignment of the Cyc-GAF domains for additional conserved cysteines. One obvious candidate was Cys-494, which is present within the DPCF motif in canonical Phys, thus placing Cys-494 close to the chromophore based on the three-dimensional structures of the PAS-less Phy Syb-Cph1 (8). When tested recombinantly, we found that Cys-494 also contributes to bilin attachment; its replacement with alanine compromised chromoprotein assembly and prevented the residual chromoprotein from photoconversion (Fig. 2, B and C). Strikingly, the absorption maximum of the C494A mutant was remarkably similar to the PAS-less Phy Syb-Cph1(GAF) assembled with PCB (Fig. 2C and supplemental Fig. 2), suggesting that the chromophore was no longer PVB but PCB as predicted if a second binding site was eliminated. Analysis of a double cysteine mutant (C494A/C522A) then confirmed that both residues together are essential for proper PCB attachment. The C494A/C522A apoprotein failed to bind PCB covalently as judged by the lack of zinc-induced fluorescence and the lack of UV-visible absorption beyond 280 nm (Fig. 2, B and C).

Rockwell *et al.* (24) also identified a comparably positioned cysteine in the Cyc-GAF domain of a paralog of *Te-PixJ*, *T. elongatus* Tlr0924, as potentially forming a second binding site with the chromophore but proposed that this linkage was broken upon photoconversion from Pb to Pg. If true, we reasoned that the bilin bound to the C522A variant would lose its only covalent thioether linkage (provided by Cys-494) upon blue light exposure and thus readily dissociate from the apoprotein after denaturation under acidic conditions. To test this hypothesis, wild-type and C522A mutant chromoproteins were heated in SDS-PAGE buffer either before or after photoconversion with blue light and then examined for the amount of bilin...
bound by zinc-induced fluorescence. As can be seen in Fig. 2D, the state of the photoreceptor had no obvious impact on bilin attachment, indicating that the Cys-494 linkage was equally stable in both spectral forms. Collectively our data point to both Cys-522 and Cys-494 binding the chromophore independently and indicate that the stability of the Cys-494 linkage is unaffected by the spectral form of the photoreceptor.

**Molecular Modeling Points to Cys-494 Binding PCB via the C4=C5 Bridge**—Under the assumption that Cys-494 also participates in PCB binding via a thioether linkage, we attempted to determine its linkage site to the chromophore. As a first step, we generated a homology model of Te-PixJ(Cyc-GAF) using as a template the three-dimensional solution structure of its closest available relative (*i.e.* the PAS-less Phy Syb-Chp1 (8)) in combination with our alignment of cyanochromes (Fig. 3A). The predicted chromophore pocket was similar to other Phys, including the appropriate placement of residues known to be key to Phy photochemistry (5, 8, 31, 42), confirming that the model represents a good reflection of the Te-PixJ(Cyc-GAF) structure, especially for conserved features (Fig. 3, A and B).

When the predicted position of Cys-494 was examined specifically, we found that it could extend toward PCB but from the opposite side as Cys-522, and importantly, point toward the C4=C5 thioether bond (Fig. 3, A and B). Together with the prediction that PCB converts to a PVB-like adduct upon attachment (30), a likely scenario is that Cys-494 spontaneously attacks the C4=C5 bond to form a thioether linkage with either the C4 or the C5 carbon, which then generates a PVB-type chromophore (Fig. 3C). The C5 carbon is the more likely target given its closer predicted distance (3.05 Å versus 4.28 Å) to the Cys-494 sulfhydryl group.

For further support of a double thioether linkage of Te-PixJ to a PVB-type chromophore, we compared the absorption spectra of wild-type Te-PixJ(Cyc-GAF) with the C494A and C522A chromoproteins after denaturation in acidic urea, which can better distinguish between PCB and PVB due to reduced modification of bilin photosorption by non-covalent contacts with the apoprotein. As reported previously (30), wild-type Te-PixJ(Cyc-GAF) has two absorption maxima at 564 and 597 nm as Pb in acidic urea with the denatured chromoprotein being poorly photochromic after blue light irradiation (Fig. 4).

Denatured Pg has a maximal absorption at 541 nm, which is then red-shifted by blue light, suggesting that the bilin in Pg retains some ability to photoisomerize back to the Pb geometry without non-covalent protein contacts. In agreement with a PVB-type chromophore, these spectra were substantially blue-shifted by 50–70 nm as compared with the Pr and Pfr forms of Syb-Chp1(GAF), which bears a PCB chromophore linked via a single thioether linkage to the C3 carbon (8) (Fig. 4).

We predicted that elimination of the Cys-494 thioether linkage in Te-PixJ(Cyc-GAF) should now generate a PCB-type chromophore attached by a single thioether linkage between Cys-522 and the C3 carbon. In agreement, the absorption spectrum of the non-photochromic C494A mutant in acidic urea was identical to that of Syb-Chp1(GAF) denatured as Pr (Fig. 4). Both spectra were slightly blue-shifted relative to free PCB (maximum absorbance of ~687 nm), implying that the bilin in Te-PixJ(Cyc-GAF) remained bound to the Cyc-GAF domain (Fig. 4). Similar analysis of the C522A mutant denatured as Pb, which we predicted would bind PCB via a thioether linkage at the C4 or C5 position to generate a PVB-type chromophore, did not initially generate the expected PVB spectra in acidic urea. Instead, its absorption spectrum closely matched that of free PCB (Fig. 4 and supplemental Fig. 3A), suggesting that the chromophore was released from Pb in acidic urea. However, analysis of the Pg form of the C522A mutant after photoconversion implied that the chromophore remained bound. Its absorption in acidic urea was substantially blue-shifted from free PCB and remained photochromic, a property not observed with free PCB (Fig. 4).

To further explore the stability of the bilin linkage between Cys-494 and the chromophore, we first dissolved the assembled C522A mutant in acidic urea as Pb or Pg, precipitated the chromoprotein with trichloroacetic acid, and then measured the absorption of the soluble and precipitated fractions (supplemental Fig. 3). As observed above, the precipitated forms of Pb and Pg retained a near equal amount of chromophore following SDS-PAGE with about 50% released into the soluble fraction, indicating that the linkage of the bilin to Cys-494 was equally stable in both spectral forms. The bound forms had absorption spectra similar to PVB (absorption maximum at 611 nm). The released form from Pg had a similar absorption, which likely

![](http://www.jbc.org/)
NMR Spectroscopy of the Te-PixJ Chromophore in Pr and Pb States—To help define the chemistry and architecture of the bilin during photoconversion, we examined by NMR Te-PixJ(Cyc-GAF) assembled with isotopically labeled PCB. Using our previously described labeling scheme with $^{15}$NALA and 5-$^{13}$CALA (8, 31), we selectively introduced $^{15}$N into all four pyrrole ring nitrogens or $^{13}$C into the C4, C5, C9, C10, C11, C15, and C19 carbons of the chromophore (Fig. 5A). Data from the $^1$H-$^{15}$N two-dimensional correlation revealed five distinct $^{15}$N chemical shifts of 127.50, 131.25, 132.25, 132.40, and 145.00 ppm, representing the four pyrrole amides (Fig. 5B). These data unambiguously demonstrate that all four nitrogens are protonated in Pb, with the smaller split peaks at $^{13}$H11011 ppm most likely reflecting a single N–H moiety experiencing minor conformational exchange between two distinct states.

Upon photoconversion to Pg, all four N–H peaks remained evident with the 132 ppm peaks appearing to merge into a single more intense peak with a chemical shift at 132.25, demonstrating unambiguously that the Pg state is fully protonated as well (Fig. 5B). Of the four pyrrole nitrogens, only the $^1$H–$^{15}$N cross-peak at 9/145 ppm experienced a dramatic change in chemical shift, most likely moving to the 10.3/135 coordinates (Fig. 5B). This simultaneous $^1$H chemical shift increase (or deshielding) and $^{15}$N decrease (or shielding) during the Pb to Pg photoconversion could result from either a change in the hydrogen bonding of this nitrogen or a strong geometric strain. Taken together, it appears that: (i) the protonated pyrrole nitrogens in the Cyc-GAF domain are fully protected from the solvent in both conformers (Pb and Pg), (ii) the Pg chromophore is more stable/homogenous than Pb, and (iii) only one of the N–H moieties experiences a significant change in its chemical environment during photoconversion.

Labeling the chromophore with 5-$^{13}$CALA allowed us to examine the environment of the methine bridges between the pyrrole rings and the D-ring carbonyl during Pb → Pg photoconversion (14) (Fig. 5A). Combined one-dimensional (by $^{13}$C direct detection) and two-dimensional ($^1$H–$^{13}$C HSQC) NMR detected distinct $^{13}$C peaks at 108.25 and 108.75 ppm and a cluster at 110 ppm within the methine region (Fig. 5C). The cluster at 110 ppm clearly represented a single C–H group experiencing either a shifting proton or slight changes in geometry, as evidenced by the single $^{13}$C peak seen in the one-dimensional spectrum and the heterogeneity in the $^1$H dimension of our $^1$H–$^{13}$C HSQC plot. The peaks at 108.25 and 108.75 ppm

represents free PVB. However, the released form from Pb had a red-shifted absorption spectrum, which could reflect either photoisomerized free PVB or release of a non-covalently bound PCB-type bilin (absorption maximum at 694 nm).
could reflect either a single C–H moiety experiencing conformational exchange or two distinct methine groups. In the latter case, the less intense peaks might have a reduced signal in the two-dimensional plot from solvent exchange. Although we cannot yet assign these peaks to specific methine carbons (C4, C5, C10, and C15), the $^{13}$C NMR spectrum detected at least two for Pb. Strikingly, upon photoconversion to Pg, all these peaks disappeared, and an intense singlet emerged at 102.5 ppm (Fig. 5C), suggesting that substantial changes occur in or around the three bridges. Because we assume that the B–C and C–D methine bridges in PVB remain intact to generate the required extended $\pi$-conjugation system, conformational exchange likely explains the loss of these peaks.

When the C=O region of the $^{13}$C direct detect spectrum was analyzed, which would measure the environment of the C19 carbonyl of the D ring, we found a single peak at 175.92 ppm for Pb. Its sharpness implies that the D-ring is held rigidly in this state, most likely via hydrogen bonds (Fig. 5D). Upon photoconversion to Pg, this peak moved ~2 ppm and split into a less intense doublet (174.47 and 174.14 ppm), indicating conformational exchange/motion between two closely related yet distinct geometries (Fig. 5D). Conformational exchange for the D-ring carbonyl combined with environmental changes surrounding a single pyrrole N–H point to movement in and around the D ring during Pb $\rightarrow$ Pg photoconversion. This motion could reflect the proposed Z to E rotation around the C15–C16 bond of the bilin, changes in the immediate chemical environment of the D-ring, or formation or loss of hydrogen bond(s) during Pg formation (e.g. with the His-553 side chain (Fig. 3B)) (4, 5, 8).

NMR Spectroscopy Reveals Global Light-induced Movements of Te-PixJ(GAF)—To study the global movements of the Te-PixJ(Cyc-GAF) protein moiety during photoconversion, we examined by two-dimensional NMR the holoprotein isotopically labeled with $^{15}$N. $^{15}$N was uniformly incorporated by expressing the recombinant polypeptide in minimal medium containing $^{15}$NH$_4$Cl as the main nitrogen source along with excess non-labeled ALA to minimize bilin labeling. Our initial analysis with wild-type protein assembled with PCB generated poorly resolved $^1$H-$^{15}$N HSQC spectra consistent with the possible presence of multiple states caused by oligomerization (data not shown). One possible culprit was Cys-555, which was predicted from the three-dimensional model to be solvent-exposed and thus could form an intermolecular disulfide bridge (Fig. 3A). To avoid this potential complication, we generated a C555A mutant of Te-PixJ(Cyc-GAF). This mutant retained its ability to assemble with PCB and had normal Pb and Pg absorp-

FIGURE 5. NMR spectroscopy of Te-PixJ(Cyc-GAF) assembled with $[^{15}$N]PCB or $[^{13}$C]PCB. A, proposed structure of PVB bound via Cys-494 and Cys-522. The filled circles locate the carbons labeled with $^{13}$C. B, two-dimensional $^1$H-$^{15}$N HSQC NMR spectra of $[^{15}$N]PCB-labeled Te-PixJ(GAF) before (Pb) and after (Pg) irradiation with saturating blue light. The arrows highlight the $^{15}$N chemical shifts that change position upon Pb $\rightarrow$ Pg photoconversion. C, one-dimensional $^{13}$C and two-dimensional $^1$H-$^{13}$C NMR spectra of $[^{13}$C]PCB-labeled Te-PixJ(Cyc-GAF) in the methine region of the spectra before (Pb) and after (Pg) irradiation with saturating blue light. D, one-dimensional $^{13}$C NMR spectra of $[^{13}$C]PCB-labeled Te-PixJ-(Cyc-GAF) before (Pb) and after (Pg) irradiation with saturating blue light in the C=O region of the spectra (170–180 ppm).
Double Bilin Linkage in Cyanochromes

**FIGURE 6.** Two-dimensional $^1$H-$^{15}$N HSQC NMR spectra of $[^{15}$N]Te-PixJ-(Cyc-GAF) in the Pb and Pg states. NMR data were collected with the C555A variant of Te-PixJ(Cyc-GAF) with the polypeptide uniformly labeled with $^{15}$N and assembled with unlabeled PCB. Blue, NMR spectrum of Pb. Green, NMR spectrum of Pg. Magenta, anti-alias chemical shifts. The inset shows a magnification of a region in the $^1$H-$^{15}$N HSQC spectra that includes a number of peaks that substantially change position upon Pb to Pg photoconversion.

Importantly, it also provided highly resolved $^1$H-$^{15}$N HSQC spectra with uniformly sharp peaks, suggesting that this mutant remains monomeric even at high protein concentrations (Fig. 6 and data not shown). The number of peaks in the $^1$H-$^{15}$N HSQC spectrum for Pb closely matched the predicted number of backbone and side chain N–H resonances in this 177-amino acid chromoprotein.

When photoconverted to Pg by blue light, the C555A mutant of $[^{15}$N]Te-PixJ(Cyc-GAF) generated a markedly different $^1$H-$^{15}$N HSQC spectrum (Fig. 6). Close inspection revealed appreciable chemical shifts (reflecting chemical and magnetic environmental changes) for ~50% of the residues, indicating that the GAF domain of Te-PixJ may experience numerous conformational changes in its backbone, side chain, and secondary structure upon photoconversion. Some smaller secondary peaks were also observed that could reflect minor substructures or multiple stable conformations for several amide groups. Many of the chemical shifts present in Pb were completely absent in Pg (Fig. 6). Taken together with their non-overlapping absorption spectra in the blue/green spectral region for Pb and Pg (Fig. 2B), it appears that Te-PixJ(Cyc-GAF) contains little residual Pb upon photoconversion to Pg, unlike canonical Phys, which can contain upwards of 25% Pr contamination when saturating red light is used to generate Pfr (49). Moreover, the $^1$H-$^{15}$N HSQC peaks lost upon photoconversion to Pg did not return even after a 3-month incubation in darkness at 25°C (data not shown), indicating that the Pg form of Te-PixJ is remarkably stable against thermal reversion.

**IR and Resonance Raman Spectroscopic Support for a PVB Chromophore**—To provide further support for a PVB-type bilin in Te-PixJ(Cyc-GAF), we employed both IR and RR spectroscopy and compared these spectra with those calculated for *in vacuo* model bilins. We restricted our *in vacuo* models to several derivatives of PCB that represented the range of likely candidates for the bound chromophore (24, 30). These included: PCB with and without a second thioether linkage at C10, PCB with or without a second thioether linkage at C4 or C5, phycoerythrobilin (PUB) with a second thioether linkage at C4 or C5, and phycoerythrobilin (PEB) with a second thioether linkage at C15. In all cases, the C3 ethylidene group was replaced with an ethyl to mimic the thioether linkage to Cys-522. Except for PCB-B and -C, which lack the proton at the ring B and C nitrogens, respectively, the chromophores were modeled as fully protonated. In addition, chromoproteins assembled with $[^{13}$C]-labeled PCB at the C4, C5, C9, C10, C11, C15, and C19 carbons (Fig. 5A) were used for the RR and IR analyses to support the vibrational assignments. For IR spectroscopy, we generated a Pg-Pb difference spectrum to minimize signals from the protein moiety. From these analyses, we tried to identify candidate bilins for which the calculated spectra reproduced the main features of the experimental spectra, rather than seeking perfect agreement for the position of individual bands.

The experimental IR difference spectrum of Te-PixJ(Cyc-GAF) had a number of positive and negative peaks between 1500 and 1700 cm$^{-1}$ that included chromophore bands as well as amide-I bands from the protein (Fig. 7A and supplemental Fig. 4B). Several of these peaks matched calculated IR spectra for a variety of PCB derivatives with the best fit being the PVB modified at either the C4 or the C5 carbon. A surprisingly poor fit was evident for PCB modified at the C10 carbon, contradicting the proposal that this site links the bilin (24). The calculated spectrum for PCB-C10 predicted prominent peaks only above 1700 cm$^{-1}$, which contain the stretching modes of the propionate side chain C=O groups of the bilin (Fig. 7A and supplemental Fig. 5B). These peaks were absent in the experimental data, suggesting either that Pb → Pg photoconversion does not drastically perturb these carboxyl groups or that they form salt bridges with the Cyc-GAF domain, thus moving the respective C=O stretching modes out of this spectral region.

RR spectroscopy of Te-PixJ(Cyc-GAF) was performed in H$_2$O and D$_2$O buffers to determine the protonation state of the bilin by the presence or absence of the characteristic N–H in-plane bending mode at ~1570 cm$^{-1}$, which downshifts by ~500 cm$^{-1}$ upon H/D exchange (Fig. 7C). The RR spectra of Te-PixJ(Cyc-GAF) as Pb and Pg displayed vibrational band patterns similar to canonical Phys, with the most prominent bands above 1600 cm$^{-1}$ readily attributable to modes dominated by C=C stretching coordinates (Fig. 7C and supplemental Fig. 4A). Among the calculated RR spectra, the best fit to the exper-
calculated RR spectrum for PUB is similar to the calculated and experimental RR spectra for hexamethylypyromethene (50).

As evidence against a PEB-like chromophore, the experimental RR spectrum of the 13C-labeled sample displayed isotopic shifts that are clearly better matched by the calculated 30 cm⁻¹ downshift of the prominent 1622/1633 cm⁻¹ band of a PUB chromophore rather than the substantially larger 50 cm⁻¹ downshift predicted for PEB (supplemental Fig. 5A). This difference is due to the different labeling patterns of the A–B and C–D methine bridges where the most intense C=C stretching bands in PEB and PVB are located, respectively. Residual intensities at 1660 cm⁻¹ are readily attributed to amide-I bands.

Finally, the RR calculations for various PCB, PEB, and PVB species predict three or four modes between 1600 and 1660 cm⁻¹, which mainly include the C=C stretching coordinates of ring D and the A–B and C–D methine bridges. At first glance, none of these calculated spectra satisfactorily fit the two-banded pattern at 1620 and 1658 cm⁻¹ in the experimental spectra (Fig. 7C). However, if we take into account: (i) that a considerable contribution to the 1660 cm⁻¹ peaks originates from the Raman band envelope of the amide I and (ii) that structural distortions of the bilin with respect to the in vacuo geometry may cause a redistribution of the internal coordinates among the modes in this region, it is possible that the C–D methine bridge stretching, which is known to provide the largest contribution to the RR signal, may be divided into two modes when bound to the protein instead of one modeled in vacuo.

Absorption spectra predicted that the chromophore of the C494A mutant of Te-PixJ(Cyc-GAF) is PCB rather than PVB in wild type. RR spectra of the mutant confirmed this expectation (supplemental Fig. 2B). Only a single band was observed for the C494A mutant, which was similar to that generated for Syb-Cph1 and Syn-Cph1 shown previously to bind PCB via a single thioether linkage involving the C3¹ carbon (8, 31). Accordingly, the IR

![FIGURE 7. IR and RR spectroscopy of Te-PixJ(Cyc-GAF). A, experimental Pg-Pb difference IR spectra as compared with calculated IR spectra for various chromophore variants. PUB-C4, PUB-C5, and PUB-C10 denote PUB and PCB variants bound via the C4, C5, and C10 carbons, respectively. B, experimental Pg-Pb and calculated (ZZZssa-ZZZssa) IR difference spectra for chromophores with protonated (solid line) and deuterated (dashed line) pyrrole nitrogens (top two spectra) and for the natural abundance (solid line) and the 13C-labeled (dashed line) chromophores (bottom two spectra). For the location of the 13C-labeled carbons, see Fig. 5A. Solid lines = H₂O or 12C samples. The calculated spectra refer to the PUB chromophore. C, experimental RR spectra of the Pb and Pg states as compared with calculated RR spectra of PUB in the ZZZssa and ZZEssa geometries. Solid and dashed lines refer to chromophores protonated and deuterated at the pyrrole nitrogens, respectively.](http://www.jbc.org/byguestonjanuary192018)
Double Bilin Linkage in Cyanochromes

and RR spectra favor PVB as the chromophore in wild-type Te-Pix.

ZZZssa and ZZEssa Geometries Are Predicted for Pb and Pg, Respectively—Although the experimental IR and RR spectra support a PVB-like chromophore in both the Pb and the Pg states (Fig. 7 and supplemental Fig. 5), calculated spectra fail to distinguish between a thioether attachment to the C5 or C4 atom or just the addition of a hydrogen at either position. Moreover, it remains to be explained how PVB can undergo such a substantial red shift of its absorption (~106 nm) during the Pb → Pg conversion. It is conceivable, for example, that Z/E photoisomerization of the B–C methine bridge could occur simultaneously with photoisomerization of the C–D methine bridge. However, such a scenario can be ruled out by our NMR analyses, which showed that only one N–H group is active during photoconversion (Fig. 5B). Thus, the absorption changes of Te-Pix(Cyc-GAF) upon photoconversion are likely to be ascribed to differentially twisted methine bridge conformations tuning the extended π-conjugation system.

In the experimental RR spectra, the two prominent bands in the C=C stretching region shifted down by ~5 cm⁻¹ upon conversion from Pb to Pg (Fig. 7C). These downshifts were very similar to those predicted upon transition from ZZZssa to ZZEssa and agreed well with experimental analyses of biliproteins containing PVB (51). However, the intensities of the bands in this region were not well reproduced, even taking into account a contribution from the broad amide-I band to the highest frequency mode at 1660 cm⁻¹ (Fig. 7C). The analysis of the N–H bending modes and the H/D isotopic shifts in these spectra suggested that hydrogen-bond interactions of the N–H groups of the rings B and C cause this discrepancy. First, the characteristic in-phase N–H bending, which was seen as a prominent peak between 1550 and 1590 cm⁻¹ for the calculated ZZZssa and ZZEssa geometries of protonated bilins, was weak in the experimental spectra of Pb and Pg (Fig. 7C). Second and paralleling our NMR data (Fig. 5B), the C=C stretching at 1620 (Pb) and 1615 cm⁻¹ (Pg) was nearly insensitive toward H/D exchange, again unlike the calculations for protonated bilins. Conversely, the IR difference spectrum (Pg minus Pb) in the N–H out-of-phase in-plane bending (1510–1530 cm⁻¹) was satisfactorily reproduced by the calculated ZZEssa–ZZZssa difference spectrum (Fig. 7B).

Cognizant of the limitations of in vacuo spectra, the ¹³C/¹²C isotopic shifts in the RR and IR spectra were largely consistent with the assignment of the Pb and the Pg chromophores as PVB in a ZZZssa and ZZEssa configuration, respectively (supplemental Fig. 5A). In the spectral region between 600 and 850 cm⁻¹, the bands for Pb were relatively weak such that only three features could be identified that were consistent with the predictions for the ZZZssa geometry (Fig. 7C). In Pg, however, this region was dominated by two bands of medium RR intensity at 656 and 820 cm⁻¹, which agreed with the calculated modes at 652 and 806 cm⁻¹, respectively (Fig. 7C). For these modes, with the latter one (806 cm⁻¹) being dominated by the C–H out-of-plane coordinates of the C–D methine bridge, the calculated D/H and ¹³C/¹²C isotopic shifts agreed well with the experimental RR data (Fig. 7C and supplemental Fig. 5A).

The IR difference spectra do not reveal changes in the configuration of the C–D methine bridge but can provide complementary information about protein structural changes associated with the Pb → Pg transition. The positive and negative signals at 1680 and 1670 cm⁻¹ shown in Fig. 7, A and B, could be attributed to a change of the amide I bands in view of its insensitivity toward ¹³C/¹²C substitution. The height of the difference signal relative to the absolute intensity suggests that only a few peptide bonds in the polypeptide backbone undergo a light-induced conformational change. Thus, Pb to Pg phototransformation may be associated mainly with changes in amino acid side chain conformations.

Te-Pix/C Mutagenesis Yields Insights into Cyanochrome Function—To assess the contributions of residues conserved within cyanochromes, we examined the photochemical impact of several in Te-Pix(Cyc-GAF) by site-directed mutagenesis. For example, Asp-492 lies within the conserved DPCF motif unique to cyanochromes. Based on its predicted position near the chromophore, it may have an analogus function to the aspartic acid in the DIP domain of canonical Phys, which plays a key role in the deprotonation/protonation cycle of the bilin by helping form the hydrogen-bond network between the bilin and the pyrrole water (4, 42, 43) (Fig. 1B). D492A, D492H, and D492N variants all bound PCB as judged by zinc-induced fluorescence of the chromoprotein following SDS-PAGE (Fig. 8A). However, instead of forming Pb, they generated a substantially red-shifted pigment with an absorption maximum at 562 nm (Fig. 8B). The mutant chromoproteins were also photochemically locked during green light irradiation and were intensely fluorescent with excitation and emission maxima at 552 and 578 nm, respectively (Fig. 8C), indicating that Asp–492 is critical for proper cyanochrome photochemistry. Spectral analysis of denatured D492H chromoprotein suggests that these mutants contain a mixture of bound PVB- and PCB-type chromophores (Fig. 4).

The thioether linkage to the C3 carbon is also important for photochemistry. In addition to impaired bilin binding and aberrant photoconversion, the C522A mutant was strongly fluorescent with 525 and 577 nm excitation and emission maxima, respectively (Fig. 8C). Our Cys–494 Te-Pix(Cyc-GAF) substitutions, which removed the second bilin attachment site, were also strongly fluorescent but were substantially red-shifted as compared with the Cys-522 mutants (excitation and emission maxima at ~620 and 650 nm, respectively (Fig. 8C and supplemental Fig. 2). The absorbance, fluorescence, and RR spectra of these Cys-494 mutants were almost identical to comparable mutants in the PAS-less Phy Syb-Cph1 (31) and had similar fluorescence properties to mutants affecting the Cyc-GAF domain of the Te-Pix relative, Tlr0924 (24).

Two conserved arginines that interact with the propionic acid side chains of the bilin were previously shown to be important for photochemistry and Pfr stability of Deinococcus radio- durans BphP and Syb-Cph1 (Arg-133 and Arg-101 in Syb-Cph1) (8, 42). In cyanochromes, Arg-133 has been generally replaced by an alanine (Ala-518 in Te-Pix), whereas Arg-101 has been replaced by a glutamine (Gln-509), indicating that the interaction of the protein with the bilin propionates is different in cyanochromes. In support, the Q509A mutant of Te-Pix-
(Cyc-GAF) bound PCB and was photochemically indistinguishable from wild type (supplemental Fig. 6). However, we noticed that Arg-507 might provide a substitute connection to the B-ring propionate; it is conserved among cyanochromes but not among canonical Phys and is present two residues upstream from Gln-509. The R507A variant assembled with PCB but less efficiently than wild type (supplemental Fig. 6). The resulting chromoprotein had a substantially altered and weak absorption spectrum with a maximum at 531 nm (supplemental Fig. 6B). Additionally, the R459A mutant was fluorescent with excitation and emission spectra similar to C522A (Fig. 8A and data not shown). From these data, we can infer that Arg-459 is partially responsible for generating the Pb state but is not necessary for Pb → Pg photoconversion and stabilization of the Pg photopproduct.

The Synechocystis Ethylene Receptor Is a Cyanochrome—One novel member of the cyanochrome clade is Synechocystis sp. PCC6803 Etr1 ( locus slr1212), which was previously shown to bind the gaseous hormone ethylene via its transmembrane receptor domain (40). Its Cyc-GAF domain contains all the conserved motifs found in the cyanochrome family including Cys-561 that could bind PCB via the C3' position and a second cysteine in the ETCF motif (Cys-533) that could provide a second docking site to the C4/5 position (Fig. 1B). When expressed recombinantly, we found that the Cyc-GAF domain of Syn-Etr1 readily bound PCB as determined by zinc-induced fluorescence of the chromoprotein following SDS-PAGE (Fig. 2B). The resulting biliprotein photointerconverted between Pb and Pg states with absorption maxima at 400 and 534 nm, respectively (Fig. 2C). This bilin ligase activity implies that Syn-Etr1 can act both as an ethylene receptor and as a light receptor.

DISCUSSION

Biochemical and phylogenetic analyses indicate that the cyanochromes represent a novel clade of Phy-type photoreceptors that predominately respond to blue and green light. By applying a combination of molecular modeling, biochemistry, and various spectroscopic methods to a representative, Te-PixJ, we provide evidence that cyanochromes are structurally similar to canonical Phys but that they bind the bilin via a unique architecture using two stable thioether linkages.
Double Bilin Linkage in Cyanochromes

The first binding site is presumed to be identical to that of canonical Phys, which binds the bilin via a thioether bond between a positionally conserved cysteine (Cys-522 in Te-PixJ) and the C3 of the A-ring ethylidene side chain. Like Rockwell et al. (24), we found a second binding site involving a second cysteine in a conserved DPCF motif. Although they proposed that the second cysteine binds to the C10 carbon of PCB to generate a rubin-like adduct that disengages upon photoconversion of Pb to Pg (24), our data are more consistent with PCB binding via the C4/C5 carbon to generate a PVB-like adduct that remains stable as both Pb and Pg. Ishizuka et al. (30) also proposed a PVB-type chromophore for these cyanochromes but speculated that reduction of the C4=C5 double bond occurred upon PCB binding by an intrinsic autoisomerase activity that moved this double bond to the C2/C3 position. Based on our data, the substantially blue-shifted absorption spectra of cyanochromes is explained by a shorter π-conjugation system, resulting from reduction of the A-B methine bridge during attachment of PCB to the second site at Cys-494. Once bound, the PCB chromophore for Te-PixJ(Cyc-GAF) is protonated (cationic) in both the Pb and Pg states similar to previous observations for the Pr and Pfr forms of canonical Phys (31, 54).

We note that several Phy sequences from cyanobacteria cluster phylogenetically within the cyanochrome family but are without the second cysteine, including members of the Frenyella diplisophon RcaE subgroup (55) and possibly a PixJ relative from Anabena (Nostoc) sp. PCC7120 (Fig. 1B, supplemental Fig. 1, and supplemental Table 2). Spectroscopic studies of assembled An(Nos)-PixJ recently demonstrated that it photo-interconverts between green- and red light- absorbing forms (25), indicating that it has distinct photochemical properties consistent with a different bilin linkage as compared with members of the cyanochrome clade. Consequently, these proteins may represent another photochemically distinct group of chromoproteins within the Phy superfamily.

Assembly studies with Cys-522 and Cys-494 mutants indicate that Te-PixJ can bind PCB by either thioether linkage independently, albeit with much reduced efficiency. In fact, binding of PCB to the C494A mutant generates a biliprotein with absorption and RR spectra nearly identical to more typical Phys such as Syn-Cph1 and Syb-Cph1, which bind PCB via a single thioether linkage involving the C3 carbon (9, 31). The assembly of the C494A mutant is more robust than the C522A mutant, suggesting that binding of PCB to Cys-522 via the A-ring ethylidene side chain is thermodynamically favored as compared with PCB binding to Cys-494 at the C4/5 site. In denaturation experiments, we detected a PCB-like adduct associated with the C522A apoprotein but released upon denaturation, which could reflect either photoisomerized and released PVB or PCB non-covalently bound to the apoprotein in the absence of the Cys-522 ligation site.

By a number of criteria, Te-PixJ and probably other cyanochromes appear to be photochemically similar to canonical Phys despite differences in their bilins, in their binding architectures, and in their GAF domain sequences. In fact, we found that a number of residues demonstrated to be critical for the photochemistry of canonical Phys (8, 42, 43) may play similar roles in cyanochromes. These include Asp-492, Tyr-493, His-523, His-553, and Arg-507 (Fig. 3). According to our alignments and the predicted three-dimensional model of Te-PixJ(Cyc-GAF), Asp-492 appears to be in a similar position to the aspartic acid in the conserved DIP motif in canonical red-absorbing Phys. The DIP aspartate is central to the extensive hydrogen-bond lattice involving the pyrrole nitrogens, the central pyrrole water, and His-523 and appears to be critical for the deprotonation/reprotonation cycle of the chromophore during Pr → Pfr photoconversion (42, 43). Similar to canonical Phys, substitutions of Asp-492 block Pb → Pg photoconversion and generate highly fluorescent variants.

Also parallel canonical Phys, substitution of Tyr-463 for histidine does not affect bilin incorporation or formation of the ground state (Pb) but does impair photoconversion to the activated form (Pg). Although replacements of the equivalent tyrosine in Syn-Cph1 and PhyA and PhyB from the higher plant Arabidopsis thaliana generate intensely fluorescent biliproteins (52, 53, 56), similar replacements in the Te-PixJ and the PAS-less Phy Syb-Cph1 are poorly fluorescent (this report and Ref. 31). This disparity further supports the hypothesis that this tyrosine has different roles among the various Phy clades (57). Arg-507 in Te-PixJ was modeled in a similar position as Arg-101 of Syb-Cph1. In Syb-Cph1, Arg-101 has little impact on the absorption spectrum of Pr and Pfr or photointerconversion but substantially stabilizes the Pfr form against thermal reversion (8). In Te-PixJ(Cyc-GAF), replacement of Arg-507 with an alanine dramatically altered the absorption spectrum of the chromoprotein in the dark-adapted state and blocked its photoconversion, indicating that this residue has a much more pronounced effect in cyanochromes. Finally, our model identified two histidines (His-523 and His-553) near the chromophore of Te-PixJ. In canonical Phys, comparable histidines are essential for proper photochemistry, presumably by participating in the hydrogen-bond lattice involving the pyrrole nitrogens and interacting with the D ring, respectively (4, 5, 42, 43). Although not analyzed here, we expect that these residues play similar roles in Te-PixJ.

As is the presumed case for canonical Phys (1, 4, 5, 7), the experimental RR spectra of Te-PixJ are consistent with a ZZSSsa to ZZESsa isomerization at the C15=C16 methine bridge of PVB during photoconversion. Concomitant changes in the environment of the D ring are supported by NMR spectra for Te-PixJ(Cyc-GAF) labeled at the C19 carbon with 13C or at the pyrrole nitrogens with 15N. The 13C spectra clearly show that the carbonyl of this pyrrole ring undergoes a chemical shift change upon photoconversion, whereas the 15N spectra identify a single pyrrole as changing its environment, which by default we assign to the D ring based on the 13C data. The large 1H-15N chemical shift change assigned to the D-ring amide could be due to either a hydrogen bond breaking in Pb and re-forming in Pg or possibly a substantial geometric strain of a hydrogen bond that results in simultaneous 15N shielding and 1H deshielding. Both scenarios are compatible with the predicted Z to E rotation of the C15=C16 methine bridge for Phys, but this rotation remains to be experimentally confirmed.

Two-dimensional NMR with the Te-PixJ(Cyc-GAF) domain labeled with 15N revealed numerous rearrangements of the polypeptide during Pb → Pg photoconversion similar to that
seen for the GAF domain from the PAS-less Phy SyB-Cph1 during Pr → Pfr photoconversion (31). Although these rearrangements could reflect conformational changes in its backbone and secondary structure upon photoconversion, IR data imply that changes in amino acid side chain conformations predominate.

The distinct absorption spectra of Pb and Pg suggested that Pb → Pg photoconversion of Te-PixJ is nearly complete with little residual Pb present after saturating irradiation with blue light. This was confirmed by comparison of the 1H,15N HSQC NMR spectra for dark adapted and blue light-irradiated samples. Many of the chemical shifts seen in the Pb samples were absent in the Pg samples, indicating that little or no Pb remained. Thus, cyanochromes offer the unique opportunity to examine the two photochromic states of a Phy in isolation without substantial contributions from the other spectral form. An exciting possibility is that cyanochromes could be used to generate crystallographic, NMR, or single particle electron microscopic structures of the photoactivated wild-type form of a Phy, which has been hampered in canonical Phys by the residual dark-adapted Pr form substantially contaminating the preparations.

Like canonical Phys, we and Rockwell et al. (24) found that cyanochromes become highly fluorescent when photochemically impaired by various mutations affecting the bilin-binding pocket. Here, we found that several Asp-492 variants are intensely green/yellow fluorescent with peak emissions around 578 nm, whereas the Cys-494 substitutions are intensely orange/red fluorescent with peak emissions around 650 nm. In addition to being potentially useful fluorophores by themselves, this overlap in excitation and emission spectra offers the attractive scenario where these fluorophores could be combined for resonance energy transfer approaches. For example, the emission spectra of the D492H and C522A mutations of Te-PixJ overlap well with the excitation spectra of the various Cys-494 mutations or with the D86H mutation of SyB-Cph1, which have excitation and emission spectra at 620 and 650 nm, respectively (31) (Fig. 8C), suggesting that efficient energy transfer from one to the other is possible. The attractiveness of the Te-PixJ-type fluorophores is also enhanced by their presumed heat stability and their small size, which for Te-PixJ requires just the 177-amino acid GAF domain of itself for efficient bilin assembly and fluorescence. Cyanochromes in combination with the recently discovered IR-emitting Phys should enhance the repertoire of research tools in the growing field of bilin-based fluorescence reporters (31, 58, 59).

Exhaustive searches of the various microbial genome data bases with the Cyc-GAF sequence of Syn-PixJ and Te-PixJ found a number of cyanochrome representatives in cyanobacteria but none outside of this phylum. In evolutionary context, this restriction suggests that cyanochromes evolved from Cphs after the divergence of cyanobacteria from other bacterial phyla, likely to meet the special photobiological needs of these organisms. Their closest relatives are the PAS-less Phys that includes SyB-Cph1, a clade that also appears to be restricted to cyanobacteria (supplemental Fig. 1) (31). Although the PAS-less Phys are a priori absent an N-terminal PAS domain, only a portion of the cyanochromes appears to be without this domain. One consistent difference between canonical/PAS-less Phys and cyanochromes is the absence in cyanochromes of the insertion between the β3 strand and α3. In canonical Phys, this sequence forms the lasso portion of the knot that connects the PAS and GAF domains (4–7, 9). Without the PAS domain, this region forms a solvent-exposed flexible domain in PAS-less Phys (8).

Another main difference is the apparent absence of a PHY domain downstream of the GAF domain in cyanochromes. The PHY domain is essential for full Pr to Pfr photoconversion and the thermal stability of Pfr in canonical Phys (1, 3). Intriguingly, the two PAS-less Phys studied thus far, SyA-Cph1 and SyB-Cph1, are less reliant on the PHY domain for Pr → Pfr photoconversion as compared with canonical Phys (31), suggesting that PAS-less Phys deemphasized the need for the PHY domain that proceeded further in cyanochromes, culminating with the complete loss of this domain. Taken together, some cyanochromes can bind bilins and generate their photochromic behavior with only the GAF domain, making their photosensory module the smallest known in the Phy superfamily.

Two well studied Phy-like proteins cluster phylogenetically in the cyanochrome clade: Syn-PixJ (TaxD1) that regulates phototaxis toward blue light (26, 27) and S. elongatus CikA that is involved in blue light entrainment of the circadian rhythm (47). At least for RcaE, its blue-shifted absorption better overlaps with the absorption of the photosynthetic pigments phycocyanin and phycocerythrin and thus better adjust the abundances of these two photosynthetic accessory complexes to the ambient light environment. Like Te-PixJ, both the RcaE and the CikA apoproteins display a strong preference of PCB over BV (60, 61). Similarly, they also assemble poorly with purified PCB in vitro, suggesting that in vivo assembly systems should be used when attempting to generate large quantities of fully assembled cyanochromes.

Also in the cyanochrome clade is Syn-Etr1, previously shown to bind ethylene via a binding pocket sandwiched between two predicted transmembrane domains (40). Using our recombinant assembly system, we found that Syn-Etr1 readily bound PCB to generate a blue/green light photochromic biliprotein. Based on the structural organization of this receptor, it is plausible that its histidine kinase output domain is controlled by both ethylene and light as a way to integrate chemical and light signals. Interestingly, it has been reported that ethylene is produced naturally in seawater by photooxidation of dissolved organic carbons (62). Thus, it is possible that abiotically derived ethylene provides a second indirect sensory input for cyanobacteria seeking to optimize light capture.

Acknowledgments—We thank Dr. Sarah Paterson for the Syn-Etr1-containing plasmid and Dr. Katrina Forest for helpful discussions. This study made use of the NMRFAM, which is supported by National Institutes of Health Grants P41RR02301 (Biomedical Research Technology Program/National Center for Research Resources (BRTP/NCCR)) and P41GM66326 (NIGMS). Additional equipment was purchased with funds from the University of Wisconsin-Madison, the National Institutes of Health (Grants RR02781 and RR08438), the National Science Foundation (Grants DMB-8415048, OIA-9977486, and BIR-9214394), and the United States Department of Agriculture.
Cyanochromes Are Blue/Green Light Photoreversible Photoreceptors Defined by a Stable Double Cysteine Linkage to a Phycoviolobilin-type Chromophore

Andrew T. Ulijasz, Gabriel Cornilescu, David von Stetten, Claudia Cornilescu, Francisco Velazquez Escobar, Junrui Zhang, Robert J. Stankey, Mario Rivera, Peter Hildebrandt and Richard D. Vierstra

doi: 10.1074/jbc.M109.038513 originally published online August 11, 2009

Access the most updated version of this article at doi: 10.1074/jbc.M109.038513

Alerts:
- When this article is cited
- When a correction for this article is posted

Click here to choose from all of JBC's e-mail alerts

Supplemental material:
http://www.jbc.org/content/suppl/2009/08/11/M109.038513.DC1

Read an Author Profile for this article at
http://www.jbc.org/content/suppl/2009/10/02/284.43.29757.DC1

This article cites 60 references, 26 of which can be accessed free at
http://www.jbc.org/content/284/43/29757.full.html#ref-list-1