The deposition of amyloid-like filaments in brain is the central event in the pathogenesis of neurodegenerative diseases. Here we report cellular models of intracytoplasmic inclusions of \( \alpha \)-synuclein, generated by introducing nucleation seeds into SH-SY5Y cells with a transfection reagent. Upon introduction of preformed seeds into cells over-expressing \( \alpha \)-synuclein, abundant, highly filamentous \( \alpha \)-synuclein-positive inclusions, which are extensively phosphorylated and ubiquitinated and partially thioflavin-positive, were formed within the cells. SH-SY5Y cells that formed such inclusions underwent cell death, which was blocked by small-molecular compounds that inhibit \( \beta \)-sheet formation. Similar seed-dependent aggregation was observed in cells expressing 4-repeat tau by introducing 4-repeat tau fibrils, but not 3-repeat tau fibrils or \( \alpha \)-synuclein fibrils. No aggregate formation was observed in cells overexpressing 3-repeat tau upon treatment with 4-repeat tau fibrils.

Our cellular models thus provide evidence of nucleation-dependent and protein-specific polymerization of intracellular amyloid-like proteins in cultured cells.

The conversion of certain soluble peptides and proteins into insoluble filaments or misfolded amyloid proteins is believed to be the central event in the etiology of a majority of neurodegenerative diseases (1-4). Alzheimer’s disease (AD) is characterized by the deposition of two kinds of filamentous aggregates, extracellular deposits of \( \beta \)-amyloid plaques composed of amyloid \( \beta \) peptides (A\( \beta \)), and intracellular neurofibrillary lesions consisting of hyperphosphorylated tau. In Parkinson’s disease (PD) and dementia with Lewy bodies, filamentous inclusions consisting of hyperphosphorylated \( \alpha \)-synuclein (\( \alpha \)-syn) are accumulated in degenerating neurons (5). The deposition of prion proteins in synapses and extracellular spaces is the defining characteristic of Creutzfeldt-Jakob disease (CJD) and other prion diseases (3). The identification of genetic defects associated with early-onset AD, familial PD, frontotemporal dementia and parkinsonism linked to chromosome 17 (caused by tau mutation and deposition), and familial CJD has led to the hypothesis that the production and aggregation of these proteins are central to the development of neurodegeneration. Fibrils formed of A\( \beta \) display a prototypical cross-\( \beta \) structure characteristic of amyloid (6), as do many other types of filaments deposited in the extracellular space in systemic or organ-specific amyloidoses (7), including prion protein deposits (8). Filaments assembled from \( \alpha \)-syn (9) and from tau filaments (10) were also shown to possess cross-\( \beta \) structure, as were synthetic filaments derived from exon 1 of huntingtin with 51 glutamines (11). It therefore seems appropriate to consider neurodegenerative disorders developing intracellular deposits of amyloid-like proteins as brain amyloidosis. The accumulation and propagation of extracellular amyloid proteins are believed to occur through nucleation-dependent polymerization (12,13). However, it has been difficult to establish the relevance of this process in the \textit{in vivo} situation, because of the lack of a suitable cell culture model or method to effectively introduce seeds.
into cells. For example, it has not yet been possible to generate bona-fide fibrous inclusions reminiscent of Lewy bodies as a model of PD by overexpressing α-syn in neurons of transgenic animals. Here, we describe a novel method for introducing amyloid seeds into cultured cells using lipofection, and we present experimental evidence of seed-dependent polymerization of α-syn, leading to the formation of filamentous protein deposits and cell death. This was also clearly demonstrated in cells expressing different tau isoforms by introducing the corresponding tau fibril seeds.

**EXPERIMENTAL PROCEDURES**

**Chemicals and antibodies** - A phosphorylation-independent antibody Syn102 and monoclonal and polyclonal antibodies against a synthetic phospho-peptide of α-syn (PSer129) were used as described previously (5). Polyclonal anti-ubiquitin antibody was obtained from Dako. Polyclonal anti-tau PS396 was obtained from CALBIOCHEM. Monoclonal anti-alpha-tubulin and anti-HA clone HA-7 were obtained from Sigma. Lipofectamine was purchased from Invitrogen. Monoclonal anti-tau T46 was from Zymed. AT100 and HT7 antibodies were obtained from Innogenetics.

**Preparation of α-syn seed, oligomers, and tau fibrils** - Human α-syn cDNA in bacterial expression plasmid pRK172 was used to produce recombinant protein (14). Wild-type (WT) or C-terminally HA-tagged α-syn was expressed in E. coli BL21 (DE3) and purified as described (15). To obtain α-syn fibrils, α-syn (5–10 mg/ml) was incubated at 37°C for 4 days with continuous shaking. The samples were diluted with 5 volumes of 30 mM Tris-HCl buffer (pH 7.5) and ultracentrifuged at 110,000 x g for 20 min at 25°C. The pellets were resuspended in 30 mM Tris-HCl buffer (pH 7.5) and sonicated for 5 sec x 2. The protein concentration was determined as described, and this preparation was used as Seed αS. In the case of α-syn oligomers, α-syn (10 mg/mL) was incubated at 37°C for 3 days in the presence of 10 mM exifone. After incubation, the mixture was ultracentrifuged at 110,000 x g for 20 min at 25°C. The supernatant was desalted by Sephadex G-25 (Pharmacia) column chromatography and eluted fractions (α-syn oligomers) were analysed by reversed-phase HPLC, SDS-PAGE and immunoblot analysis. Recombinant human 3-repeat tau isoform with one amino-terminal insert (3R1N) and 4-repeat tau isoform with one amino-terminal insert (4R1N) monomer and corresponding fibrils were prepared as described previously (16,17).

**Introduction of proteins into cells** - Human neuroblastoma SH-SY5Y cells obtained from ATCC were cultured in DMEM/F12 medium with 10 % FCS. Cells at ~30-50 % confluence in 6-well plates were treated with 200 μl of opti-MEM containing 2 μg of the seed α-syn WT (Seed αS), HA-tagged α-syn (Seed-HA), α-syn monomers, oligomers or tau 3R1N or 4R1N fibrils and 5 μl of Lipofectamine (LA) for 3 hr at 37°C. The medium was changed to DMEM/F12 and culture was continued for 14 hr. The cells were collected by treatment with 0.5 ml of 0.25% trypsin for 10 min at 37°C, followed by centrifugation (1,800 x g, 5 min) and washing with PBS. The cellular proteins were extracted with 100 μl of homogenization buffer containing 50 mM Tris-HCl, pH 7.5, 0.15 M NaCl, 5 mM EDTA, and a cocktail of protease inhibitors by sonication. After ultracentrifugation at 290,000 x g for 20 min at 4 °C, the supernatant was collected as Tris-soluble fraction and the protein concentration was determined by BCA assay. The pellet was solubilized in 100 μl of SDS-sample buffer. Both Tris-soluble and insoluble fractions were analyzed by immunoblotting with appropriate antibodies as indicated (15,18).

**Cell culture model of seed-dependent polymerization of α-syn or tau** - α-Syn or tau 3R1N or 4R1N was transiently overexpressed in SH-SY5Y cells by transfection of 1 μg of wild-type human α-syn cDNA in pcDNA3 (pcDNA3-α-syn) or human tau cDNA in pcDNA3 (pcDNA3-tau 3R1N or 4R1N) with 3 μl of FuGENE6 (Roche Diagnostics) in 100 μl of opti-MEM, followed by culture for 14 hr. Under our experimental conditions, the efficiency of transfection with pEGFP-C1 vector was 20-30%. The cells were washed with PBS once and then Seed αS, Seed-HA, Seed 3R1N or Seed 4R1N was introduced with Lipofectamine as described above. The medium was changed to DMEM/F12
and culture was continued for ~2-3 days. Cells were harvested in the presence of trypsin to digest extracellular cell-associated α-syn fibrils. The cellular proteins were differentially extracted and immunoblotted with the indicated antibodies, as described (18).

Confocal microscopy- SH-SY5Y cells on cover slips were transfected with pcDNA3-α-syn and cultured for 14 hr as described above, then Seed αS was introduced, and culture was continued for ~1-2 days. After fixation with 4% paraformaldehyde, the cells were stained with appropriate primary and secondary antibodies as described previously (18). For thioflavin S staining, the cells were incubated with 0.05% thioflavin S at room temperature for 5 min. Fluorescence was analyzed with a laser scanning confocal fluorescence microscope (LSM5Pascal, Carl Zeiss).

Immunoelectron microscopy- For electron microscopy, cells overexpressing α-syn were transfected with Seed αS, cultured for 2 days, fixed in 0.1 M phosphate buffer containing 4% glutaraldehyde for 12 hrs, then processed and embedded in LR White resin (London Resin, Reading, UK). Ultrathin sections were stained with uranyl acetate for investigation. Immunolabelling of the inclusions was performed by means of an immunogold-based post-embedding procedure. Sections were blocked with 10% calf serum, incubated overnight on grids with anti-PSer129 antibody at a dilution of 1: 100, rinsed, then reacted with secondary antibody conjugated to 10 nm gold particles (E-Y Laboratories, San Mateo, CA) (1:10), rinsed again and stained with uranyl acetate.

Immunoelectron microscopic analysis of α-syn or tau filaments extracted from cells was performed as follows. Cells overexpressing α-syn or tau were transfected with Seed αS or Seed tau, respectively. After incubation for 3 days, they were harvested, suspended in 200 μL of 10 mM Tris-HCl, pH 7.4/1 mM EGTA/10% sucrose/0.8 M NaCl) and sonicated. The lysates were centrifuged at 20,400 g for 20 min at 4°C. The supernatant was recovered and Sarkosyl was added (final 1% v/v). The mixtures were incubated at room temperature for 30 min, and then centrifuged at 113,000 g for 20 min. The resulting pellets were suspended in 30 mM Tris-HCl, pH 7.5, placed on collodion-coated 300-mesh copper grids, and stained with the indicated antibodies and 2% (v/v) phosphotungstate. Micrographs were recorded on a JEOL 1200EX electron microscope.

Cell death assay- Cell death assay was performed using a CytoTox 96 Non-Radioactive Cytotoxicity Assay Kit (Promega). TUNEL staining was performed using an In Situ Cell Death Detection Kit (Roche).

Assay of proteasome activity- SH-SY5Y cells transfected with pcDNA3-α-syn and Seed αS were cultured for 3 days or treated with 20 μM MG132 for 4 h. Cells were harvested and cytosolic fraction was prepared as follows. Cells were resuspended in 100 μL of phosphate-buffered saline (PBS) and disrupted by sonication, then insoluble material was removed by ultracentrifugation at 290,000 x g for 20 min at 4°C. The supernatant was assayed for proteasome activity by using a fluorescent peptide substrate, benzoyloxycarbonyl-Leu-Leu-Glu-7-amido-4-methylcoumarin (z-LLE-MCA, Peptide Institute, Inc.). 7-amino-4-methylcoumarin (AMC) release was measured fluorometrically (excitation at 365 nm; emission at 460 nm). In green fluorescent protein (GFP)-reporter assay of proteasome activity in living cells by confocal laser microscopy, SH-SY5Y cells transfected with pcDNA3-α-syn (1 μg) and GFP-CL1 (0.3 μg) using FuGENE6 and then transfected with Seed αS were grown on coverslips for 2 days or treated with 20 μM MG132 for 6 h (19). These cells were analyzed using a laser scanning confocal fluorescence microscope (LSM5Pascal, Carl Zeiss).

Statistical analysis- The P-values for the description of the statistical significance of differences were calculated by means of the unpaired, two-tailed Student's t-test using GraphPad Prism 4 software (Graphpad Software).

RESULTS

Introduction of seed α-syn into cultured cells using Lipofectamine reagent. Cellular overexpression of α-syn by itself does not lead
to fibrillization of α-syn in a form that resembles Lewy bodies. This prompted us to examine whether or not introduction of preformed aggregation seeds of α-syn (Seed αS) would elicit fibril formation. To introduce Seed αS into SH-SY5Y cells in a non-invasive manner, we tried several reagents used for transporting proteins or plasmid DNA into cells, and found that Lipofectamine (LA), a cationic gene introducer, enables the introduction of Seed αS into SH-SY5Y cells. We were not able to detect any introduced α-syn monomer or fibrils following simple addition of protein preparations to the culture medium, notwithstanding a previous report on this approach (20). The insoluble α-syn formed following LA-mediated Seed αS introduction was detected as buffer-insoluble α-syn in cell lysates (Fig. 1A). The insoluble α-syn was phosphorylated at Ser129 upon introduction into cells (Fig. 1A), indicating that Seed αS was incorporated in cells and phosphorylated intracellularly. Cells were harvested in the presence of trypsin to digest extracellular cell-associated α-syn fibrils. The optimal ratio of LA to Seed αS was about 5 μL to 2 μg protein in 6-well plates. This treatment effectively introduced Seed αS not only into SH-SY5Y cells, but also into several other types of cells examined, including CHO cells and human embryonic kidney 293T cells (data not shown). In sharp contrast, soluble α-syn (either monomeric or oligomeric forms) was not introduced into the cells by the same treatment (Fig. 1A and Fig. 4), suggesting that the LA treatment works exclusively for the internalization of insoluble α-syn aggregates.

These results strongly suggest that α-syn fibrils are incorporated with the aid of LA, but do not exclude the possibility that extracellular α-syn fibrils may induce aggregation of endogenous α-syn without incorporation. To confirm that the extracellular α-syn fibril seeds are internalized into cells by LA, we performed the transduction of preformed C-terminally HA-tagged α-syn fibril seeds (Seed-HA) instead of non-tagged α-syn seeds. As shown in Figure 1B and 1C, time-course experiments revealed that Seed-HA was also incorporated into cells in the presence of LA and could be detected with both anti-HA antibody and a phospho-α-syn specific antibody (anti-PS129), even 3 days after infection. Confocal microscopic analyses also indicated that Seed-HA was phosphorylated at Ser129 intracellularly. All anti-PSer129 positive dot-like structures were also stained with anti-HA, indicating that no endogenously phosphorylated α-syn aggregates are present in the cells (Fig. 1D and S1C).

Establishment of a cell culture model for nucleation-dependent polymerization of α-syn. Although introduction of the seed α-syn into cells was accompanied with phosphorylation, no further dramatic change was observed. Since the level of endogenous α-syn was relatively low in SH-SY5Y cells, we introduced non-tagged or HA-tagged seeds into cells transiently overexpressing α-syn. After 3 days of culture, immunocytochemistry for α-syn revealed a diffuse (Fig. 2B) or dot-like (Fig. 2C) pattern of cytoplasmic labeling by anti-PSer129 in cells transfected with wild-type α-syn without seeds, or in non-overexpressing cells with Seed αS, respectively. Surprisingly, however, in cells transfected with both pcDNA3-α-syn and Seed αS we observed abundant round inclusions that occupied the cytoplasm and displaced the nucleus, with morphology highly reminiscent of cortical-type Lewy bodies observed in human brain (Fig. 2D). The size of the α-syn-positive inclusions was approximately 10 μm in diameter (Fig. 2D), which is similar to that of the Lewy bodies detected in the brains of patients with dementia with Lewy bodies. Similarly, when cells expressing α-syn were transfected with Seed-HA, abundant phosphorylated α-syn-positive cells were also detected (Fig. S1D).

We next examined the status of ubiquitin, which is positive in most types of intracellular filamentous inclusions, including Lewy bodies, in neurodegenerative disease brains. As shown in Fig. 2E, we found that almost all intracellular inclusions labeled with anti-PSer129 were also positive for ubiquitin, as is the case for Lewy bodies in the cortex of human DLB brain (Fig. 2F). Furthermore, the juxtanuclear PSer129-positive, Lewy body-like inclusions were also positively labeled with thioflavin S, a fluorescent dye that specifically
intercalates within structures rich in β-pleated sheet conformation (Fig. 2G and 2H), indicating that the inclusions contain β-sheet-rich filamentous aggregates. Electron microscopic analysis of cells transfected with both wild-type α-syn and the seeds revealed that the inclusions are composed of filamentous structures ~10 nm in diameter, that are often covered with granular materials (Fig 2I and 2J). The filamentous structures were randomly oriented within the cytoplasm of these cells, forming a meshwork-like profile, and were frequently intermingled with mitochondria (Fig. 2I and 2J), being highly reminiscent of human cortical Lewy bodies. Immunoelectron microscopy showed that the filaments were densely decorated with anti-PSer129 (Fig. 2K), demonstrating that they were composed of phosphorylated α-syn.

To biochemically validate this cellular model and to investigate further the molecular mechanisms underlying nucleation-dependent aggregation within cells, we differentially extracted α-syn from these cells using detergents of various strengths and analyzed the extracts by immunoblotting with anti-Syn102 and PSer129 antibodies. The levels of α-syn in the sarkosyl-soluble and insoluble fractions (total α-syn and α-syn phosphorylated at Ser129, respectively) were dramatically increased in cells transfected with both wild-type α-syn and the seeds (WT+Seed αS in Fig. 3A and 3B). To distinguish endogenous α-syn from exogenous α-syn fibrils, we used LA to transduce Seed-HA into cells overexpressing α-syn. Immunoblot analyses of these cells showed that HA-tagged α-syn with slower mobility than non-tagged α-syn was detected in the Sar-insoluble pellets as phosphorylated forms by anti-HA and anti-PSer129 antibodies in cells treated with Seed-HA+LA (Fig. 3C, 3D, 3E). Interestingly, in cells expressing α-syn (WT) treated with Seed-HA+LA, much more abundant non-tagged α-syn was detected in the TX- and Sar-insoluble fractions as phosphorylated forms with a smaller amount of the HA-α-syn. We also performed a dose-dependence experiment with Seed-HA in cells expressing α-syn. As shown in Fig. S2, immunoreactive levels of TX-insoluble phosphorylated α-syn increased in parallel with increase in the amount of Seed-HA. Furthermore, we tested whether tau protein forms intracellular aggregates in the presence of α-syn seeds, instead of tau seeds. We found that tau was not aggregated with Seed-HA, confirming that intracellular aggregate formation of soluble α-syn is specific to and dependent on fibril seeds of the same protein (Fig. S3). This nucleation-dependent polymerization of α-syn in cells was greater at 3 days than at 1 day after transduction of the seeds (Fig. 3F).

Negative-stain electron microscopic observation of sarkosyl-insoluble fractions of the cells harboring inclusions revealed anti-Syn102 and PSer129-positive filaments of ~5 to 10 nm width (Fig. 3G and 3H), that are highly reminiscent of those derived from human α-synucleinopathy brains (21). Such filaments were never detected in the sarkosyl-insoluble fraction of cells solely overexpressing α-syn (data not shown). These results indicated that the biochemical characteristics of α-syn accumulated in cells forming the Lewy body-like inclusions were very similar to those of α-syn deposited in the brains of patients with α-synucleinopathies, including PD and DLB.

As the idea has been gaining ground that transient oligomers, rather than mature fibrils, are responsible for cytotoxicity, we examined whether soluble oligomers could be introduced into cells in the same manner as fibril seeds by means of LA treatment, and whether they could function as seeds for intracellular α-syn aggregate formation. As shown in Fig. 4A and 4B, we purified stable α-syn oligomers from recombinant α-syn treated with exifone, an inhibitor of in vitro α-syn aggregation, which is thought to inhibit filament formation of α-syn by stabilizing SDS-resistant soluble oligomers (22,23). Then, cells expressing α-syn or mock plasmid were treated with a mixture of the oligomer fraction (5 μg) and LA and incubated for 3 days. Immunoblot analyses of lysates of these cells did not detect any SDS-resistant soluble oligomeric α-syn, and the levels of phosphorylated α-syn in the Sar-soluble and insoluble fractions showed no increase (Fig. 4C and 4D). On the other hand, we observed phosphorylated and deposited α-syn in the Sar-soluble and insoluble fractions in cells expressing α-syn treated with Seed αS (Fig. 4C and 4D).
and 4D). These results showed that SDS-resistant soluble oligomer of α-syn could not be introduced into cultured cells in the same manner as monomeric α-syn, and/or could not function as seeds for intracellular α-syn aggregation.

**Mutagenic analysis of nucleation-dependent assembly of α-syn.** To investigate further the nucleation-dependent polymerization of α-syn, we analyzed the polymerization of α-syn mutated or truncated at various residues or subdomains that are believed to be crucial for its aggregation. Overexpression of A53T familial Parkinson mutant α-syn, which is readily fibrillogenic in vitro, in the presence of Seed αS moderately increased the accumulation and phosphorylation of α-syn in the sarkosyl-soluble and insoluble fractions compared with those in cells with wild-type α-syn expression and Seed αS (Fig. 5A and 5B). In contrast, overexpression of Δ11 mutant α-syn, an assembly-incompetent mutant lacking residues 73-83, which have been shown to be essential for fibril formation of α-syn (24), elicited neither deposition nor phosphorylation of α-syn. We next introduced α-syn into SH-SY5Y cells expressing S129A mutant α-syn, and observed slightly lower levels of sarkosyl-insoluble α-syn compared with those in cells with wild-type α-syn expression and Seed αS. However, the frequency of inclusion bodies observed in seed-transduced cells expressing S129A was similar to that in seed-transfected cells expressing wild-type α-syn (data not shown), suggesting that phosphorylation at Ser129 is not required for the nucleation-dependent polymerization of α-syn within cells.

**Nucleation-dependent intracellular polymerization of α-syn elicits neurotoxicity and cell death.** SH-SY5Y cells overexpressing α-syn started to show marked clumping suggestive of cellular degeneration and death by ~48 hr after introduction of seeds (Fig. 6B). Quantitative analysis of cell death by lactate dehydrogenase (LDH) release assay at 72 hr after introduction of Seed αS showed that cells overexpressing wild-type, A30P, A53T or S129A α-syn released ~30% of total LDH from total cell lysate, whereas only ~12% of LDH was released from cells expressing Δ11 mutant α-syn, which lacks polymerization ability. In control cells transfected with empty vector or pcDNA3-α-syn followed by treatment with Lipofectamine without seeds, only ~7 % of LDH was released (Fig. 6C). These results suggest a close correlation between the seed-dependent aggregation of α-syn and cell death. However, the dying cells transfected with fibrilization-competent α-syn and seeds did not show typical morphological changes of apoptosis, e.g., nuclear fragmentation, positive TUNEL staining (Fig. S4A) or activation of caspase-3 (Fig. S4B), suggesting that they did not undergo typical apoptotic cell death, despite a previous report that exposure to neuron-derived extracellular α-syn may cause apoptosis (25).

**Impairment of proteasome activity in cells with intracellular aggregates of α-syn.** Since α-syn is ubiquitinated in the brains of patients with α-synucleinopathies (26) and inhibition of ubiquitin-proteasome systems by aggregates of proteins with expanded polyglutamine tracts has been reported (27), we analyzed the ubiquitination state of cellular proteins in α-syn aggregate-forming cells and compared the pattern with that in cells treated with a proteasome inhibitor, MG132. Sarkosyl-soluble fraction of seed-transduced cells expressing wild-type α-syn, and harboring abundant inclusions, showed increased levels of ubiquitin-positive staining, which was similar in pattern to that observed in cells treated with MG132 (Fig. 6D). Because this pattern suggested an impairment of the ubiquitin-proteasome system, we directly analyzed the proteasome activity of α-syn inclusion-forming cells using a specific fluorescent peptide substrate, z-LLE-MCA, that emits fluorescence following proteasomal digestion, and confirmed that proteasome activity was significantly reduced in these cells, as well as in cells treated with 20 μM MG132 for 4 h (Fig. 6E). We further examined the suppression of proteasome activity using CL1, a short degron that has been reported to be an effective proteasome degradation signal (28), and whose fusion protein with green fluorescent protein (GFP-CL1) has been used as a reporter for inhibition of proteasomal activity by intracellular polyglutamine aggregates (27) and intracellular...
α-syn (19). To examine if intracellular α-syn inclusions affected proteasomal activity, SH-SY5Y cells were transfected with both wild-type α-syn and GFP-CL1, followed by the introduction of Seed αS. Fluorescent signals of GFP were scarcely detected in control cells transfected with GFP-CL1 alone (Fig. 6F, none), but were markedly increased upon treatment with proteasome inhibitor MG132 (Fig. 6F, MG132), confirming that GFP-CL1 was effectively degraded by proteasome. Strikingly elevated GFP signals were detected in cells forming α-syn inclusions (Fig. 6F, WT+Seed αS) compared with those in control cells (Fig. 6F, none or WT), and GFP-CL1 and deposits of phosphorylated α-syn were co-localized within these cells (arrowheads). These results strongly suggest that proteasome activity is impaired in cells harboring α-syn inclusions elicited by the introduction of Seed αS.

Small-molecular inhibitors of amyloid filament formation protect against cell death induced by seed-dependent α-syn polymerization. We have previously shown that several classes of small-molecular compounds inhibit amyloid filament formation of α-syn, tau and Aβ in vitro (17,23). These observations prompted us to test whether these inhibitors exert a protective effect against death of SH-SY5Y cells mediated by the nucleation-dependent polymerization of α-syn. Figure 7A shows the effects of three polyphenol compounds, exifone, gossypetin, and quercetin, and a rifamycin compound, rifampicin, added to the culture media at a final concentration of 20 or 60 μM. Remarkably, all these compounds blocked cell death, with gossypetin being the most effective. Our previous in vitro studies elucidated that several polyphenols, including gossypetin and exifone, inhibit α-syn assembly, and that SDS-stable, noncytotoxic soluble α-syn oligomers are formed in their presence (23), suggesting that such polyphenols may inhibit filament formation of α-syn by stabilizing soluble, prefibrillar intermediates. Gossypetin or exifone might suppress intracellular α-syn aggregate formation by stabilizing such soluble intermediates in cultured cells as well. Immunoblot analysis revealed that the levels of sarkosyl-insoluble α-syn in cells transfected with both α-syn and seeds were reduced by treatment with exifone or gossypetin compared with those in untreated cells (Fig. 7B), supporting the notion that these compounds entered the cytoplasm and blocked cell death by suppressing the seed-dependent polymerization of α-syn.

Cellular models for nucleation-dependent polymerization of tau. Beside α-syn, tau is another major pathogenic protein that is deposited in degenerating neurons or glial cells in various neurodegenerative diseases, and aggregation of distinct tau isoforms has been found in different diseases, i.e., deposition of 3-repeat tau isoforms in Pick’s disease, 4-repeat tau isoforms in progressive supranuclear palsy and corticobasal degeneration, and both 3- and 4-repeat tau isoforms in AD. It is unknown why distinct tau isoforms deposit in different diseases. Thus, we also tried to establish a cellular model of intracellular tau aggregate formation by transduction of tau fibril seeds into cultured cells.

First, we confirmed that expression of 3R1N or 4R1N by itself induced phosphorylation of Ser396, but no aggregated form was detected in detergent-insoluble fractions (Fig. 8 and S5). Next, we tested whether introduced tau 4R1N or 3R1N fibril seed (Seed 4R1N or 3R1N, respectively) is detectable by immunoblot analysis using anti-T46, anti-HT7 or anti-PS396 antibody. However, we could not detect any band in TX-insoluble fractions of cells treated with Seed tau 4R1N or 3R1N in the presence of LA with any of these antibodies (data not shown). It seems likely that the efficiency of introduction of tau 4R1N and 3R1N fibrils by LA treatment is very low, as compared with that of Seed αS. Then, we checked whether treatment with recombinant tau fibrils causes intracellular tau aggregate formation in a LA-dependent manner. As shown in Fig. S5, LA treatment itself did not cause intracellular tau deposition in cells expressing tau 4R1N without Seed 4R1N. Recombinant tau 4R1N monomer in the presence of LA did not elicit the formation of intracellular tau aggregates in these cells. On the other hand, when Seed 4R1N was added to cells expressing tau 4R1N with LA, aggregated and phosphorylated tau was detected in Sarkosyl-insoluble fractions by immunoblot analyses of
these cell lysates using anti-HT7 or anti-PS396 antibody (Fig. S5 and 8). In the case of intracellular tau 3R1N aggregate formation, the results were similar to those in the experiments using tau 4R1N described above (data not shown).

Intracellular aggregated 4- or 3-repeat tau was also found to be detected with not only anti-PS396 but also anti-AT100 antibody in the Sar-insoluble fraction (Fig. 8B and 8C). Phosphorylated and deposited tau was not found in the TX-insoluble fraction of tau-expressing cells without tau seed treatment or mock plasmid-expressing cells treated with tau seed. In accordance with findings described earlier in this paper, these results suggested that soluble 4- or 3-repeat tau expressed from the plasmid was accumulated into intracellular inclusions in the presence of small amounts of Seed 4R1N or 3R1N.

We also found that hyperphosphorylated and aggregated tau was not detected in 3-repeat tau-expressing cells treated with Seed 4R1N (Fig. 8B and 8C). On the other hand, aggregated form of 3-repeat tau was detected in TX-insoluble fractions of 3-repeat tau-expressing cells treated with Seed 3R1N, and hyperphosphorylation at Ser396 and Ser212/Thr214 was observed in fractionated samples of these cells, whereas no such bands were detected in 4-repeat tau-expressing cells treated with Seed 3R1N (Fig. 8B and 8C). These results clearly showed that 4-repeat tau fibrils can be seeds for polymerization of 4-repeat tau and 3-repeat tau fibrils can be seeds for polymerization of 3-repeat tau. Tau does not polymerize (cross-seed) in the presence of seeds of a different isoform. Similarly, no tau aggregation was detected in tau-expressing cells treated with α-syn fibril seeds (Fig. S3C and S3D), and no α-syn aggregation was detected in α-syn-expressing cells transduced with tau fibril seeds (data not shown). Furthermore, we observed anti-AT100 and anti-PS396-positive tau 4R1N or 3R1N filaments of ~15 nm width by negative-stain electron microscopic analyses of Sarkosyl-insoluble fractions of cells transfected with both tau plasmid and the seeds (Fig. 9A-9D).

Confocal microscopic analyses also showed that GFP-tagged tau 4R1N (GFP-tau 4R1N) is aggregated into round inclusions in the presence of Seed 4R1N together with LA (Fig. 8E). No inclusion-like structures were found in cells expressing GFP-tau 4R1N (Fig. 8D) or in cells expressing GFP-tau 4R1N after treatment with Seed 3R1N (data not shown). The ratio of the round aggregates to all GFP-positive transfectants was calculated to be 5.8% ± 0.8602 (P=0.0002 by Student's t-test against the value of cells expressing GFP-4R1N, n=5). Significant cell death was not observed in cells containing intracellular 3R1N or 4R1N aggregates (data not shown). These results strongly suggest that proteins assemble easily into amyloid fibrils in the presence of amyloid seeds derived from the same protein, but not a different protein.

**DISCUSSION**

Nucleation-dependent protein polymerization occurs in many well-characterized physiological processes, e.g., microtubule assembly and actin polymerization. It is also the mechanism of amyloid fibril formation in various pathological conditions, and has been confirmed to occur in vitro for a wide variety of extracellular amyloids, such as Aβ peptides and prion proteins (12,13), as well as intracellular proteins, such as α-syn and tau (29,30). Both extra- and intracellular amyloids have been well studied in vitro, but much less is known about the mechanisms of assembly in vivo. Here we report a simple and effective method to introduce polymerization seeds into cells using Lipofectamine, a widely used transfection reagent. This method enabled us to evaluate the nucleation-dependent polymerization of α-synuclein, and to establish a cellular model of the neurodegeneration seen in Parkinson’s disease.

Lipofectamine is a reagent widely used for the transfection of DNA into eukaryotic cells through the formation of liposomes of polycationic and neutral lipids in water, based on the principle of cell fusion. Various methods, including microinjection, the calcium phosphate method, the DEAE-dextran method, electroporation, and viral transfer, have been
employed to introduce substances that are not normally incorporated into eukaryotic cells under physiological conditions. Microinjection is versatile, but is not efficient in experiments involving large numbers of cells, and the traumatic damage to cells hampers evaluation of cytotoxic effects. Here, we have successfully employed lipofection to introduce protein aggregates as seeds for amyloid fibril formation (patent pending for US [12/086124], for EU [06834541.2], and for Japan [2007-549210]). The reason why Lipofectamine could specifically incorporate Seed αS, but not soluble α-syn, into cells is unknown. However, one possibility is that aggregated α-syn with an ordered filamentous structure was preferentially bound to Lipofectamine and formed a complex that could be more effectively transported into cells compared with soluble α-syn, which has a random-coil structure. In line with this idea, it has been reported that yeast prion fibrils can be introduced into yeast cells (31). Recently, Luk et al. have also reported that α-syn monomers and fibrils, but not oligomers were introduced into cells by Bioporter, a cationic-liposomal protein transduction reagent (32).

We confirmed the incorporation of insoluble α-syn seeds into cells by detecting phosphorylation of α-syn, as has been seen in intracellular aggregates of α-syn in various neurodegenerative conditions referred to as synucleinopathies. This suggests that Seed αS introduced into cells is a good target for phosphorylation at Ser129. In contrast to our results, a recent report suggested that α-syn fibrils were not phosphorylated after internalization (32). It is possible that this specific phosphorylation represents an active attempt by cells to maintain the intracellular milieu by sequestering protein species that are harmful to cells. Notably, the phosphorylation of α-syn was dramatically increased when Seed αS was introduced into cells overexpressing α-syn (Fig. 3, S1D and S2). The possibility therefore arises that widespread propagation of hyperphosphorylation of α-syn throughout the cytoplasm reflects the activation of a certain kinase(s) associated with conversion of soluble α-syn into fibrillar form in the presence of Seed αS. However, further investigation is needed to elucidate the importance of phosphorylation for protein aggregation.

The significance of intracellular and extracellular protein aggregates in neurodegeneration is still a matter of debate. The present results clearly show that nucleation-dependent polymerization of amyloid proteins is closely related to neuronal degeneration leading to cell death. According to the seeding theory, amyloid fibrils grow rapidly, without a time-lag, when seeds are exposed to an amount of amyloidogenic soluble protein that exceeds the critical concentration. Our experiments with seed-transfected SH-SY5Y cells overexpressing α-syn clearly demonstrated that this is the case in the intracellular environment. We have unequivocally demonstrated that nucleation-dependent polymerization of amyloid-like fibrils can occur inside cells, and the intracellular filament formation elicits a variety of cellular reactions, including hyperphosphorylation and compromise of the ubiquitin proteasome system. We also showed that α-syn oligomers were not introduced into cells by LA and did not function as seeds for α-syn aggregate formation in cultured cells. It has been speculated that protein fibrils, not oligomers, are spread or transmitted in recently reported in vivo models (25,33).

Our study also revealed that intracellular protein aggregation is highly dependent on the species of protein fibril seeds. This important finding may explain why only a single tau isoform is deposited in several tauopathies, including Pick's disease, progressive supranuclear palsy and corticobasal degeneration. In this study, α-syn fibrils were shown to be unable to seed intracellular tau aggregation, which is consistent with neuropathological reports that deposited α-syn is not markedly co-localized with tau aggregates. Our observations strongly support a seed-dependent mechanism for the formation of the intracellular protein aggregates.

Importantly, we showed that seed α-syn or tau, an insoluble aggregate prepared from α-syn or tau filaments, is effectively incorporated into cells by lipofection. This, in turn, suggests that high-molecular-weight protein aggregates or amyloid seeds shed from one cell may easily be propagated to others (e.g., neurons or glial cells).
under pathological conditions, e.g. alteration in membrane permeability owing to aging or virus infection, impairment of membrane function as a result of physical interaction with extracellular amyloid deposits, or abnormal membrane depolarization, that favor intracellular deposition of protein fibrils.

It remains to be clarified whether the incorporation of amyloid seeds into neurons or glial cells, as shown in this study, also occurs in vivo. However, some observations in AD or in transgenic animals support this possibility: apolipoprotein E (apoE) is involved in lipoprotein particle uptake mediated by cell surface receptors, and the E4 allele is the strongest genetic risk factor for AD. The apoE polypeptide has also been shown to bind Aβ (34), tau (35) and NAC region of α-syn (36) and to be localized in amyloid plaques and neurofibrillary tangles in AD, and prion plaques (37) in CJD. ApoE and low-density lipoprotein receptor-related protein (LRP) facilitate intraneuronal Aβ42 accumulation in transgenic mice (38). Furthermore, activation of both endocytic uptake and recycling of these proteins at a preclinical stage has been reported in sporadic AD and Down’s syndrome (39), Thus, it is strongly suggested that extracellular amyloid may be taken up into neurons by apoE and LRP-mediated endocytosis. Therefore, intracellular amyloid seeds composed of α-syn or tau may also be incorporated into neurons by similar mechanisms when these seeds are released to the extracellular space after neuronal death.

It is well established that tau protein starts to accumulate in the entorhinal region and spreads to the neocortices, closely correlating with the progression of AD (40). Similarly, accumulation of phosphorylated α-syn has been shown to start in vulnerable regions (i.e., limbic cortices) and to spread to the neocortices in PD or DLB. However, the mechanism of propagation of abnormal protein deposition remains unknown. This study strongly supports a seed-dependent mechanism for the formation of the intracellular protein aggregates. In the context of our propagation hypothesis, it will be crucial to inhibit not only the production of intracellular amyloid seeds, but also their spread into the extracellular space. Vaccination against the intracellular amyloid proteins, such as α-syn (41) or tau may be an effective approach, together with inhibition of intracellular amyloid filament formation by small-molecular inhibitors, for the therapy of these diseases.

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FOOTNOTES

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The abbreviations used are: AD, Alzheimer's disease; PD, Parkinson's disease; DLB, dementia with Levy bodies; α-syn, alpha-synuclein; CJD, Creutzfeldt-jakob disease; WT, wild-type; tau 3R1N, 3-repeat tau isoform with one amino-terminal insert; tau 4R1N, 4-repeat tau isoform with one amino-terminal insert; LA, Lipofectamine; HPLC, high performance liquid chromatography; LDH, lactate dehydrogenase; TUNEL, terminal deoxynucleotidyl transferase-mediated dUTP-biotin nick end labeling; z-LLE-MCA, benzyloxycarbonyl-Leu-Leu-Glu-7-amido-4-methylcoumarin.

FIGURE LEGENDS

Fig 1. Introduction of seed α-syn into cultured cells with Lipofectamine reagent. (A) Purified recombinant α-syn (soluble form; 2 μg) and filaments (2 μg) were sonicated and then incubated with Lipofectamine (LA). The protein-LA complexes were dispersed in opti-MEM and added to SH-SY5Y cells. After 14 hr of culture, the cells were collected and sonicated in SDS sample buffer. After boiling, the samples were analyzed by immunoblotting with a phosphorylation-dependent anti-α-syn PSer129 (right) or a phosphorylation-independent antibody Syn102 (left). (B-C) C-terminally HA-tagged α-syn fibril seeds (Seed-HA) were transduced into cells by the use of LA. After incubation for 1 day or 3 days, cells were harvested with or without trypsin and proteins were differentially extracted from the cells with Tris-HCl (TS), Triton X-100 (TX), and Sarkosyl (Sar), leaving the pellet (ppt). Immunoblot analyses of lysates using anti-HA and anti-PSer129 are shown. The immunoreactive band positive for anti-HA or anti-PSer129 in the TX-insoluble fraction was quantified. (D) Confocal laser microscopic analysis of cells treated with Seed-HA in the presence of LA. Cells were transduced with 2 μg of Seed-HA using 5 μL of LA. After 48 h incubation, cells were fixed and immunostained with anti-PSer129 (green) and anti-HA (red), and counterstained with TO-PRO-3 (blue).

Fig 2. Confocal laser and electron microscopic analyses of α-syn inclusions in plasmid-derived α-syn-expressing cells treated with seed α-syn. (A-D) Confocal laser microscopic analyses of control SH-SY5Y cells transfected with pcDNA3 vector and Lipofectamine alone (A), cells transfected with pcDNA3-α-syn (WT) (B), cells transduced with the seed α-syn (Seed αS) (C), and cells transfected with both pcDNA3-α-syn and Seed αS (WT+Seed αS) (D) immunostained with anti-PSer129 (green), and
counterstained with TO-PRO-3 (blue). Arrows indicate cytoplasmic round inclusions stained with anti-PSer129. Scale bars, 20 μm.

(E-F) Comparison of confocal images of cells transfected with both α-syn plasmid and Seed αS (E) and tissue sections from DLB brains (F) using anti-PSer129 (green) and anti-ubiquitin antibodies (red). Cytoplasmic inclusions in transfected cells (arrows) are positive for ubiquitin, like Lewy bodies (arrowheads) in DLB brains. Scale bars, 20 μm.

(G-H) Confocal microscopic images of cells transfected with both pcDNA3-α-syn and Seed αS. Cells were stained with 0.05% Thioflavin S (green) and anti-PSer129 antibody (red). The boxed area on the left is shown in the right panel. Scale bars, 20 μm on the left and 10 μm on the right.

(I-J) Electron microscopic analyses of cells transfected with both pcDNA3-α-syn and Seed αS. High magnification of the boxed area in (I) is shown in (J). An asterisk or arrow indicates a nucleus or mitochondrion, respectively. Scale bars, 2 μm in (I) and 200 nm in (J).

(K) Immunoelectron microscopic observation of cells transfected with both pcDNA3-α-syn and Seed αS using a polyclonal antibody against phosphorylated Ser129 of α-syn. Scale bar, 200 nm.

Fig 3. Immunoblot and immunoelectron microscopic analyses of intracellular α-syn aggregates in cultured cells.

(A-B) Immunoblot analysis of α-syn in cells treated with Seed αS alone (Seed αS), pcDNA3-α-syn alone (WT), or both WT and Seed αS (WT+Seed αS). Proteins were differentially extracted from the cells with Tris-HCl (TS), Triton X-100 (TX), and Sarkosyl (Sar), leaving the pellet (ppt). Blots were probed using anti-α-syn (Syn102) (A) and anti-PSer129 (B).

(C-F) Immunoblot analysis of proteins differentially extracted from mock (none) or cells transfected with pcDNA3-α-syn (WT), cells transduced with Seed-HA with (Seed-HA+LA) or without LA treatment (Seed-HA-LA), and cells overexpressing α-syn treated with Seed-HA with (WT+Seed-HA+LA) or without LA treatment (WT+Seed-HA-LA). Immunoreactivity of phosphorylated α-syn in the TX-insoluble fraction was quantified using anti-PSer129 and the results are shown in (F).

(G-H) Immunoelectron microscopy of α-syn filaments extracted from transfected cells. SH-SY5Y cells were transfected with both pcDNA3-α-syn and Seed αS. Sarkosyl-insoluble fraction was prepared from the cells, and the filaments were immunolabelled with anti-Syn102 (G) or PSer129 (H) antibody. Scale bar, 200 nm.

Fig 4. α-Syn oligomers were not introduced into cultured cells.

(A–B) α-Syn oligomers were prepared as described in Experimental Procedures. Oligomeric α-syn protein incubated with (47.8 μg protein) or without exifone (30 μg protein) was analysed by reversed-phase HPLC (Aquapore RP-300 column) (A). These samples (0.2 μg protein of each) were also analyzed by SDS-PAGE and immunoblotted with anti-Syn102 (B).

(C-D) Cells were transfected with empty plasmid (none) or pcDNA3-α-syn (αsyn) and then treated with or without α-syn oligomer (Oligomer αS, 5 μg) or fibrils (Seed αS, 2 μg). After incubation for 3 days, cells were harvested and immunoblot analyses were performed. Proteins differentially extracted from the cells with Tris-HCl (TS), Triton X-100 (TX), and Sarkosyl (Sar), and the pellet (ppt), were probed using anti-Syn102 (C) and anti-PSer129 (D).

Fig 5. Effects of α-syn mutations on intracellular deposition.

Immunoblot analysis of α-syn in cells transfected with pcDNA3-α-syn alone (WT), Seed αS alone (Seed αS), both WT and Seed αS (WT+Seed αS), and non-treated control cells (none). Cells overexpressing familial PD-linked A30P or A53T polymerization-deficient Δ11 mutant α-syn followed by transfection with Seed αS were also analysed. Proteins were extracted...
differentially with Tris-HCl (TS), Triton-X (TX), and Sarkosyl (Sar), leaving the pellet (ppt), and immunoblotting was done with anti-Syn102 and PSer129. The PSer129-immunoreactive bands detected in sarkosyl-soluble and insoluble fractions from each cell type shown in (A) were quantified (B). The results are expressed as means + S.E.M. (n=3).

Fig 6. Cell death caused by formation of intracellular α-syn inclusions.
(A-B) Phase-contrast microscopy of the control cells (A) and cells transfected with both pcDNA3-α-syn and Seed αS (B) 3 days after treatment with Seed αS (20 x objective).
(C) The extent of cell death of transfected cells was quantified using LDH release assay. Cells transfected with α-syn plasmid alone (WT, A30P, A53T, S129A or Δ11) or with both wild-type or several mutants and Seed αS were incubated, and cell death assay was performed 3 days thereafter. The results are expressed as means + S.E.M. (n=5). * Not significant; **P<0.01; ***P<0.0005 by Student's t-test against the value of Seed αS.
(D-F) Impairment of proteasome activity caused by intracellular aggregates of α-syn. (D) Immunoblot analysis of proteasome activity by intracellular aggregates of α-syn. Cytosol fractions of non-treated control cells (none), cells transfected with wild-type α-syn plasmid alone (WT) or with both pcDNA3-α-syn and Seed αS (WT+Seed αS), and cells treated with 1 μM MG132 for 16 h (MG132) using anti-ubiquitin antibody. An arrow indicates monomeric ubiquitin. Poly-ubiquitinated proteins, reflecting impairment of the proteasome activity, are observed in the sarkosyl-soluble fraction. TS, Tris-soluble; TX, 1% Triton X-100-soluble; Sar, 1% sarkosyl-soluble; ppt, sarkosyl-insoluble and SDS-soluble. (E) Peptide hydrolysis activity of proteasome. Cytosol fractions of non-treated control cells (none), cells transfected with wild-type α-syn plasmid alone (WT) or with WT and Seed αS (WT+Seed αS), and cells treated with 20 μM MG132 for 4 h (MG132) were prepared and assayed using z-LLE-MCA as a substrate. The results are expressed as means + S.E.M. (n=3). *P<0.05; **P<0.01; ***P<0.0005 by Student's t-test against the value of none.
(F) Proteasome activity in cells having intracellular aggregates of α-syn. SH-SY5Y cells transfected with both GFP-CL1 and WT were treated with Seed αS for 2 days, fixed, and stained with anti-PSer129. In the staining of cells transfected with wild-type α-syn plasmid alone (WT), anti-Syn102 was used. As a control, untreated or MG132-treated cells were also stained and analyzed. In untreated control cells, the fluorescence of GFP was poorly detected, because GFP-CL1 could be degraded by proteasome in cells. In cells treated with MG132, fluorescence was markedly increased as compared with that in untreated cells because of the inhibition of proteasome activity by MG132. Co-localized images (arrowheads) with both increased intensities of GFP (green) and the fluorescence of anti-PSer129 (red) were detected in cells transfected with both WT and Seed αS (WT+Seed αS), indicating that the proteasome activity in these cells was inhibited.

Fig 7. Small-molecular inhibitors of amyloid filament formation protect against cell death caused by intracellular α-syn aggregates.
(A) The cell death of cells transfected with Seed αS and with both α-syn plasmid (WT) and Seed αS in the presence or absence of 20 or 60 μM gossypetin, 20 or 60 μM exifone, 60 μM quercetin, or 60 μM rifampicin was quantified by LDH release assay. The results are expressed as means + S.E.M. (n=4). * Not significant; **P<0.05; ***P<0.0005 by Student's t-test against the value of none. (B) Immunoblot analyses of the sarkosyl-insoluble fraction prepared from cells transfected with Seed αS and with both WT and Seed αS in the absence or presence of exifone or gossypetin, with anti-Syn102 and anti-PSer129 antibodies. Doubly transfected cells were treated with 20 or 60 μM exifone or gossypetin 2 h after transfection of Seed αS, and cultured for 3 days in the presence of polyphenols. Tubulin-alpha loading controls are also shown.
Fig. 8. Immunoblot analyses of intracellular tau aggregates. 
(A-C) Immunoblot analysis of tau in cells treated with tau fibrils alone (Seed 3R1N or Seed 4R1N), pcDNA3-tau alone (3R1N or 4R1N), or both Seed tau and pcDNA3-tau. Tau proteins differentially extracted from the cells with Tris-HCl (TS), Triton X-100 (TX) and Sarkosyl (Sar), and the pellet (ppt), were probed with anti-T46 (A), anti-P396 (B) and anti-AT100 (C).

Fig. 9. Cellular models for intracellular tau aggregation. 
(A-D) Immunoelectron microscopy of tau filaments extracted from transfected cells. SH-SY5Y cells were transfected with both pcDNA3-tau 3R1N and Seed 3R1N (A, B) or pcDNA3-tau 4R1N and Seed 4R1N (C, D). Sarkosyl-insoluble fraction was prepared from the cells, and the filaments were immunolabelled with anti-AT100 (A, C) or PS396 (B, D) antibody. Scale bar, 200 nm.

(E-G) Confocal laser microscopic analyses of SH-SY5Y cells transfected with pEGFP empty vector (E), pEGFP-tau 4R1N (F) and cells transfected with both pEGFP-tau 4R1N and Seed 4R1N (G), immunostained with anti-PS396 (red), and counterstained with TO-PRO-3 (blue). Scale bars, 10 μm.
Fig. 2 Nonaka et al

A. none
B. pcDNA3-αsyn (WT)
C. Seed αS
D. WT+Seed αS

E. WT+Seed αS
   anti-Ubiquitin  anti-PSer129
   merge
   Thioflavin S  merge

F. DLB brain sections
   anti-Ubiquitin  anti-PSer129
   merge
   anti-PSer129  Phase
   merge

G. anti-PSer129  Phase
   merge

H. anti-PSer129  Phase
   merge

I.

J.

K.

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Fig. 3 Nonaka et al
Fig. 4 Nonaka et al

A

B

C

anti-Syn102

(kDa)

37 25 15

TS TX Sar ppt TS TX Sar ppt TS TX Sar ppt

none αsyn αsyn + Oligomer αS

D

anti-PSer129

(kDa)

37 25 15

TS TX Sar ppt TS TX Sar ppt TS TX Sar ppt

none αsyn αsyn + Oligomer αS

TS TX Sar ppt

αsyn + Seed αS
Fig. 9 Nonaka et al

A  anti-AT100  B  anti-PS396  C  anti-AT100  D  anti-PS396

3R1N + Seed 3R1N  4R1N + Seed 4R1N

E  TO-PRO-3  anti-PS396

GFP  merge

F  TO-PRO-3  anti-PS396

GFP  merge

GFP-tau 4R1N  GFP-tau 4R1N + Seed 4R1N
Seeded aggregation and toxicity of α-synuclein and tau: cellular models of neurodegenerative diseases
Takashi Nonaka, Sayuri T. Watanabe, Takeshi Iwatsubo and Masato Hasegawa

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