The Tumor Suppressor DiRas3 Forms a Complex with H-Ras and C-RAF and Regulates Localization, Dimerization and Kinase Activity of C-RAF*

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Running title: Regulation of Ras/RAF signaling by DiRas3

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Background: The tumor suppressor DiRas3 interferes with the Ras/MAPK mitogenic cascade.

Results: DiRas3 associates with H-Ras and C-RAF and regulates localization, dimerization and kinase activity of C-RAF.

Conclusion: DiRas3 regulates the Ras/MAPK cascade at the level of Ras/RAF signal transmission.

Significance: Learning how DiRas3 affects the Ras/MAPK signaling is crucial for understanding of the molecular mechanisms underlying DiRas3-mediated tumor suppression.

SUMMARY

The maternally imprinted Ras-related tumor suppressor gene DiRas3 is lost or down-regulated in more than 60% of ovarian and breast cancers. The anti-tumorigenic effect of DiRas3 is achieved through several mechanisms including inhibition of cell proliferation, motility and invasion, as well as induction of apoptosis and autophagy. Re-expression of DiRas3 in cancer cells interferes with the signaling through Ras/MAPK and PI3K. Despite intensive research, the mode of DiRas3’s interference with the Ras/RAF/MEK/ERK signal transduction is still a matter of speculation. In this study we show that DiRas3 associates with the H-Ras oncogene and that activation of H-Ras enforces this interaction. Furthermore, while associated with DiRas3, H-Ras is able to bind to its effector protein C-RAF. The resulting multimeric complex consisting of DiRas3, C-RAF and active H-Ras is more stable than the two-protein complexes H-Ras/C-RAF or H-Ras/DiRas3, respectively. The consequence of this complex formation is a DiRas3-mediated recruitment and anchorage of C-RAF to components of the membrane skeleton, suppression of C-RAF/B-RAF heterodimerization and inhibition of C-RAF kinase activity.

The Ras/RAF/MEK/ERK mitogenic cascade is one of the essential signal transduction pathways in the cell, involved in the control of different cellular processes including proliferation, differentiation, transformation, survival, adherence, and motility. This pathway couples extracellular signaling via ligand-bound receptor tyrosine kinases (RTKs), SOS (“son of sevenless”), and activated Ras to the cytoplasmic extracellular signal-regulated kinase (ERK), a multi-substrate kinase, which in turn phosphorylates and activates several proteins in the cytosol and nucleus including various transcription factors such as Elk-1, Ets, and Sp1 (1-2). The interaction of the GTP-loaded Ras with RAF, the key regulator of the pathway, represents the initial and essential step in the activation of the mitogenic cascade.

The family of RAF protein kinases, which comprises A-, B- and C-RAF, shares three highly conserved regions: CR1, CR2, and CR3. The CR3 region represents the catalytic domain, whereas CR1 contains a Ras binding domain (RBD) and a zinc binding domain also called...
cysteine-rich domain (CRD). The serine/threonine-rich CR2 contains a conserved 14-3-3 binding motif. Cytosolic C-RAF exists as a multiprotein complex, consisting of heat shock proteins, 14-3-3 proteins and kinase suppressor of Ras (KSR). Upon stimulation of cell surface receptors, C-RAF translocates to the plasma membrane, associates with Ras-GTP and undergoes a series of activation events including interaction with lipids and regulatory proteins, numerous phosphorylation and dephosphorylation events as well as dimer formation (for review see (3-6)). RAF dimerization proved to be one of the most decisive steps in the regulation of RAF activity, particularly in light of recent findings concerning the paradoxical behavior of some RAF inhibitors in cancer treatment (7-11).

The Ras subfamily of GTP-hydrolyzing oncoproteins is part of a large superfamily of more than 170 evolutionary conserved proteins related to Ras. About 35 members constitute the Ras subfamily, whereby H-, N-, and K-Ras represent the best characterized members (12-13). Ras proteins alternate between GTP- (active “on” state) and GDP-bound (inactive “off” state) conformations. As the “switch-on” and “switch-off” reactions in the cycle of Ras are intrinsically very low, the regulatory input by guanine nucleotide exchange factors (GEFs) and GTPase-activating proteins (GAPs) determines the lifetime of the two states (14-16). Binding of Ras effector proteins to GTP-Ras triggers distinct signaling cascades. RAF kinase was first discovered as Ras effector followed by RafGDS and PI3K. The family of Ras effectors expanded over the past years and includes currently more than ten different proteins (13). Ras proteins are synthesized as cytosolic precursors that undergo posttranslational processing (C-terminal prenylation and palmitoylation) to be able to associate with cellular membranes (12,17-18). Localization of Ras at the cytosolic leaflet of cellular membranes is believed to be required for their biological activity. Ras signaling events previously presumed to be restricted to the plasma membrane have now been observed on intracellular membranes, including endosomes, the endoplasmic reticulum (ER) and the Golgi apparatus (19-24).

The small GTPase DiRas3 (also referred as NOEY2 and ARHI) belongs to the Ras family of proteins and shares 55-62 % homology with Ras and Rap (25-26). Intriguingly, in contrast to most Ras proteins and despite its high degree of GTP-bound state in resting cells, DiRas3 acts as a tumor-suppressor, thus possessing entirely different functional properties compared to H-, N- and K-Ras. DiRas3 gene encodes a 26-kDa protein that is monoallelically expressed and maternally imprinted (25). As a member of the Ras protein family, DiRas3 contains three typical motives: a GTP binding domain, a putative effector domain and the membrane localization motif CAAX (15). However, there are also some unique characteristics, which distinguish DiRas3 from other members of the Ras protein family. It contains a 34-amino-acid extension at the N-terminus and differs from H-Ras in residues critical for GTPase activity and for putative effector function. The substitutions within the GTP binding domain of DiRas3 are consistent with the mutations of Ras responsible for its constitutive activation. Correspondingly, DiRas3 has been found predominantly in its GTP-bound state in cells (27). DiRas3 is lost or down-regulated in more than 60% of ovarian and breast cancers through several different mechanisms including loss of heterozygosity, DNA hypermethylation, transcriptional regulation, and shortened mRNA half-life (26,28). Loss of DiRas3 expression is associated with tumor progression and poor prognosis (29-30). Re-expression of DiRas3 in cancer cells inhibits growth, decreases invasiveness and induces apoptosis (25,31). Signaling alterations caused by introduction of the DiRas3 gene into cancer cells lacking DiRas3 expression range between inhibition of the Ras/MAPK pathway, activation of JNK, inhibition of the STAT3 transcriptional activity, and downregulation of cyclin D1 (25,27,32).

The studies reported on DiRas3 function so far, suggest that the biological activities of DiRas3 GTPase could not only be explained by its effects on a single pathway. Despite considerable progress the molecular mechanisms of the DiRas3 tumor-suppressive activity are not sufficiently elucidated. In particular, the mode of DiRas3 interference with the Ras/MAPK signaling cascade is still a matter of speculation. In the present study, we report that DiRas3 interacts with H-Ras oncogene and that activation of H-Ras enforces its association with DiRas3, indicating that the tumor-suppressive activity of DiRas3 is achieved, at least in part, at the level of Ras signaling. Furthermore, our study reveals that, although associated with DiRas3, H-Ras is able to bind to its effector C-RAF and that the multimeric complex consisting
of DiRas3, C-RAF and active H-Ras is more stable than the two-protein complexes H-Ras/C-RAF or H-Ras/DiRas3, respectively. The consequence of this complex formation is a DiRas3-coordinated translocation and anchorage of C-RAF to components of the membrane skeleton (MSK). In addition, DiRas3 disrupts the H-Ras-induced heterodimerization of C-RAF with B-RAF and suppresses the kinase activity of C-RAF.

**EXPERIMENTAL PROCEDURES**

**Antibodies**—The following antibodies were used: mouse anti-c-myc (9E10), rabbit anti-C-RAF (RAF-1, C-12), mouse anti-HA (12CA5), mouse anti-KDEL (10C3), mouse anti-pERK (E-4), rabbit anti-ERK2 (C-14), rabbit anti-B-RAF (C-19), and mouse anti-vimentin (V9) from Santa Cruz; mouse anti-H-Ras (#R02120) from BD Transduction Laboratories; rabbit anti-phospho-C-RAF-Ser-338 (56A6, was also used for detection of phospho-Ser-446 in B-RAF) from Cell Signaling Technology; mouse anti-M2PK (DF4) from Scheco Biotech; rabbit anti-EEA1 (#E3906) from Sigma; mouse anti-PARP-1 (C-2-10) from Calbiochem; mouse anti-penta-His™ (#34660) from Qiagen. The anti-DiRas3 (6EC.2) antibody (kindly provided by R. Kroschewski) was raised in rabbit against partially purified full length native His-DiRas3. The horseradish peroxidase-labeled (for Western blot) and Cy2- or Cy3-conjugated (for indirect immunofluorescence microscopy) anti-mouse and anti-rabbit secondary antibodies were from Dianova.

**Plasmids**—The following plasmids were used: human C-RAF WT-myc-His in pcDNA3, human HA-C-RAF WT and HA-C-RAF-R89L in pcDNA3, human B-RAF WT-myc-His in pcDNA3, human B-RAF WT in pCMV, human DiRas3-myc and ∆DiRas3-myc in pRK5; human H-Ras12V, H-Ras17N, H-Ras12V/35S, H-Ras12V/37G, H-Ras12V/40C, H-Ras12V/186S, and H-Ras12V/181/4SS in pcDNA3; human K-Ras12V in pEXV3. The plasmids encoding mutants of H-Ras12V were kindly provided by I. Rubio (University of Jena).

**Immunoprecipitation**—The required cDNA plasmids were transfected into COS7 cells under starvation conditions using jetPEI transfection reagent (Polyplus Transfection). Cells were lysed 24 h after transfection with buffer containing 10 mM Tris-HCl (pH 8.0), 50 mM NaCl, 30 mM sodium pyrophosphate, 100 μM Na3VO4, 1% Triton X-100, and standard proteinase inhibitors for 45 min at 4 °C. The lysates were clarified by centrifugation at 27,000 × g for 15 min and incubated for 1 h at 4 °C with the appropriate antibody. After addition of protein G-agarose, the incubation was continued for 2 h at 4 °C. The agarose beads were washed 3 times with lysis buffer containing 0.1% Triton X-100. The immunoprecipitates were supplemented with Laemmli buffer, boiled for 5 min at 100 °C, and applied to SDS-PAGE. After Western blotting the isolated proteins were visualized by appropriate antibodies.

**In Vitro Kinase Assay**—The kinase assay was carried out directly with immunoprecipitated proteins in 25 mM Hepes, pH 7.6, 150 mM NaCl, 25 mM β-glycerophosphate, 10 mM MgCl2, 1 mM dithiothreitol, 1 mM Na3VO4 and 500 μM ATP buffer (50 μl final volume). Recombinant MEK and ERK-2 were used as substrates. After incubation for 30 min at 30 °C, the kinase assay mixtures were supplemented with Laemmli buffer, boiled for 5 min at 100 °C and applied to SDS-PAGE. After Western blotting the extent of ERK phosphorylation was determined by an anti-phospho-ERK antibody.

**Subcellular Fractionation**—Cell fractions were isolated using the ProteoExtract subcellular proteome extraction kit (Calbiochem). COS7 cells were grown on 10-cm Petri dishes and transfected with appropriate cDNA constructs under starvation conditions. The cells were fractionated into four subproteomic fractions (cytosolic and nuclear fractions, fractions of whole membranes, and cytoskeleton) according to the manufacturer’s protocol. Proteins were separated by SDS-PAGE and blotted onto nitrocellulose membrane. The selectivity of subcellular extraction was documented by immunoblotting against marker proteins (M2PK for cytosolic fraction, KDEL (ER retention signal) for membrane fraction, PARP (poly(ADP-ribose) polymerase) for nuclear fraction, and vimentin for cytoskeletal fraction).

**Indirect Immunofluorescence Microscopy**—COS7 cells were grown on coverslips and transfected with required cDNA plasmids. The cell imaging was performed either with whole cells or following cytoplasm and/or membranes extraction by use of ProteoExtract subcellular proteome extraction kit (Calbiochem). For the whole cell imaging, cells were fixed with 4% paraformaldehyde in 1xPBS (Morphisto) for 15 min and permeabilized with 0.1% v/v Triton X-
100 in 1xPBS for 5 min. For the imaging of the cytoplasm/membranes depleted cells, fixation (15 min) with 4% paraformaldehyde in 1xPBS was performed immediately after extraction. In all cases fixation was stopped by incubation with 50 mM NH₄Cl in 1xPBS for 15 min. Immunostaining of the proteins was performed with specific antibodies and fluorescently labeled secondary antibodies that were diluted in 1xPBS containing 0.5% BSA. After three washes with 1xPBS and brief wash with deionized water the coverslips were mounted using Mowiol® mounting medium. Samples were analyzed with a LAS AF software controlled Leica TCS SPE confocal microscope. TIFF images were processed with the Adobe Photoshop 7.0 software.

RESULTS

DiRas3 Binds Preferentially to the Active and Non-farnesylated H-Ras—Several reports documented that the growth of the ovarian and breast cancer cell lines could be inhibited by exogenous expression of DiRas3 (25,27,32). The DiRas3-induced growth inhibition has been proved by use of various cell lines, such as HeLa, Saos-1, NIH3T3, 293, and COS7 (25). Although much effort has been devoted to the study of this phenomenon, the mechanism by which DiRas3 inhibits cell proliferation is still not fully understood. Yu et al. (25) and Luo et al. (27) have reported that introduction of DiRas3 gene into cancer cells triggers apoptosis, downregulates cyclin D1, activates JNK, and impairs signaling through Ras/MAPK pathway. As Ras/RAF/MEK/ERK cascade is a central pathway coordinating cell proliferation, differentiation, migration, and apoptosis, it appears very likely that DiRas3 may interfere with signal transduction between the modules of this cascade.

The Ras/RAF interaction represents the initial and crucial step in the signal transduction of the Ras/RAF/MEK/ERK cascade. Regarding the mode of Ras coupling to RAF the formation of Ras homodimers has been reported to be essential for C-RAF activation (33-34). Also numerous other GTPases are regulated by dimerization (33,35). Therefore, we hypothesized that DiRas3 may regulate the Ras/RAF/MEK/ERK cascade by formation of heterodimers with H-Ras. To test this issue, we performed coimmunoprecipitation of the constitutively active H-Ras12V or dominant-negative H-Ras17N with the myc-tagged DiRas3. To this end, the proteins of interest were expressed in COS7 cells. The results of this assay revealed that DiRas3 indeed formed a stable complex with H-Ras in vivo (Fig. 1A). Moreover, DiRas3 bound preferentially to the H-Ras12V mutant suggesting that activation of H-Ras enforced its association with DiRas3. Interestingly, DiRas3 appeared to interact with both, farnesylated (the lower band in the anti-H-Ras blot (36)) and non-farnesylated (the upper band in the anti-H-Ras blot (36)) forms of the inactive H-Ras17N. On the contrary, in the case that the active H-Ras12V was assayed, DiRas3 bound exclusively to its non-farnesylated form (Fig. 1A). As the unique N-terminal extension of DiRas3 has been shown to be important for its inhibitory effect on cell growth (27), we also tested the DiRas3 mutant lacking the 34 amino-acid N-terminal extension (∆NDiRas3) for its ability to interact with H-Ras. As shown in Fig. 1A, N-terminal deletion reduced the binding affinity of DiRas3 to H-Ras.

H-Ras Lipidation Sites are not Required for the DiRas3/H-Ras Interaction—As demonstrated above, the lipidation status of H-Ras plays a role in the interaction with DiRas3. To examine, whether lipidation sites or lipidation of H-Ras in general are required for the interaction with DiRas3, we tested the binding of DiRas3 to farnesyl-deficient mutant H-Ras12V/186S and palmitoyl-deficient mutant H-Ras12V/181/4SS (13,15). As a control for the experimental conditions used in our coimmunoprecipitation assay, we made use of the interaction between the C-RAF and H-Ras. As reported previously, in contrast to B-RAF, which associates effectively with both farnesylated and non-farnesylated H-Ras, C-RAF requires farnesylated H-Ras (37). As expected, the loss of the palmitoylation sites did not severely impair the association of H-Ras with C-RAF, but this binding was completely abolished, if the farnesylation site of H-Ras was mutated (Fig. 1B). These results are in line with published data (37). In contrast, loss of the palmitoylation or farnesylation sites in H-Ras did not affect its interaction with DiRas3 under the same experimental conditions (Fig. 1B). This finding is in agreement with the binding selectivity of DiRas3 toward the active, non-farnesylated H-Ras (Fig. 1A).

H-Ras Effector Domain is not Essential for the H-Ras/DiRas3 Interaction—Because the coimmunoprecipitation experiments did not
provide information on whether the interaction between H-Ras and DiRas3 takes place directly or indirectly, the possibility that this association might be mediated by an effector protein of H-Ras could not be excluded. Therefore, we investigated the binding properties of DiRas3 toward the H-Ras mutants that were impaired in their binding to effector proteins. For that purpose, we made use of three different H-Ras effector domain mutants: H-Ras12V/35S, H-Ras12V/37G and H-Ras12V/40C, which are altered in activation of effector proteins RAF, RafGDS and PI3K (38). Again, as a control for the experimental settings of our coimmunoprecipitation assay, we used the interaction between C-RAF and the H-Ras mutants. All H-Ras effector domain mutants tested were indeed severely impaired in their binding to C-RAF (Fig. 1C). In contrast, the interaction between DiRas3 and H-Ras was not compromised by the mutations within the effector domain (Fig. 1C). These findings exclude the possibility, that effector proteins mediate the association between DiRas3 and H-Ras. Furthermore, the data of this experiment indicate that the effector domain of H-Ras is not involved in the DiRas3/H-Ras interaction.

C-RAF Associates with the H-Ras/DiRas3 Complex—As the effector domain of H-Ras is not involved in the DiRas3/H-Ras interaction, H-Ras might still be able to bind to its effector proteins, while associated with DiRas3. To explore this issue, we used C-RAF as the best characterized representative of the H-Ras effector proteins. To this end, we expressed myc-DiRas3 and H-Ras12V together with C-RAF kinase in COS7 cells. The DiRas3-bound proteins were isolated by the anti-myc immunoprecipitation and tested for the presence of H-Ras and/or C-RAF. Indeed, as shown in Fig. 2A, C-RAF WT coprecipitated with DiRas3 in the presence of H-Ras12V, suggesting that all three proteins form a multimeric complex, given that H-Ras is present in its active state. The association of C-RAF with the H-Ras/DiRas3 complex was disrupted by the R89L mutation within the RBD of C-RAF (Fig. 2A). This single amino acid exchange in C-RAF has been shown to disrupt H-Ras binding (39). Therefore, the data obtained with the C-RAF-R89L mutant suggest, that the association of C-RAF with the H-Ras/DiRas3 complex occurs through the C-RAF/H-Ras interaction. This conclusion is also supported by the previously published results, demonstrating that DiRas3 does not bind to the RBD of C-RAF (27). Furthermore, data presented in Fig. 2A indicate that binding of C-RAF to the H-Ras/DiRas3 complex enforces the interaction between these two GTPases, as significantly more H-Ras coprecipitated with DiRas3, if C-RAF WT was coexpressed. In contrast, expression of the C-RAF-R89L mutant did not support H-Ras/DiRas3 interaction (Fig. 2A).

Binding of DiRas3 to H-Ras Supports the Interaction Between H-Ras and its Effector Protein C-RAF—The finding that C-RAF supports the H-Ras/DiRas3 interaction raises the question, whether it works also vice versa and DiRas3 might facilitate binding of H-Ras to its effector C-RAF. To answer this question we expressed H-Ras12V and HA-tagged C-RAF together with or without DiRas3 in COS7 cells. The C-RAF-bound proteins were isolated by the anti-HA immunoprecipitation and tested for the presence of H-Ras. The C-RAF-R89L mutant was used as a negative control. The results of this experiment revealed that the amount of the coprecipitated H-Ras was two times higher, if DiRas3 was coexposed, suggesting that the binding of DiRas3 to H-Ras indeed supports the interaction between H-Ras and C-RAF (Fig. 2B). Of note, as shown in Fig. 2B, in the absence of DiRas3 coexpression, the signal of the coprecipitated H-Ras appeared as a doublet of two bands with equal intensity, indicating that both, farnesylated and non-farnesylated H-Ras12V were bound to C-RAF WT. In contrast, upon co-expression with DiRas3, the upper band of the doublet, which represents the non-farnesylated H-Ras, clearly dominated. This observation suggests that DiRas3 stabilizes selectively the complex between C-RAF and the non-farnesylated H-Ras. These data are in line with the finding that DiRas3 binds preferentially to the non-farnesylated H-Ras12V (Fig. 1A). Collectively, the results of the experiments presented in Fig. 2 lead to the conclusion that the multimeric complex consisting of DiRas3, C-RAF and active H-Ras is more stable than the two-protein complexes consisting of H-Ras/C-RAF or H-Ras/DiRas3, respectively.

DiRas3 Induces Accumulation of C-RAF within the Cytoskeletal Fraction—As the coimmunoprecipitation experiments revealed that C-RAF associates with the H-Ras/DiRas3 complex (see also Fig. 2A), we asked whether this association may affect the subcellular distribution of C-RAF. To address this question we expressed different combinations of C-RAF, DiRas3, H-Ras12V, and H-Ras17N in COS7
After membranes extraction, the staining of H-Ras12V disappeared (data not shown). As it has been previously shown that H-Ras localizes to the Golgi (19-20), we concluded that the juxtanuclear localization of H-Ras12V shown in Fig. 5A could be assigned to the association of H-Ras12V with the Golgi. The netlike localization of H-Ras12V resembles the MSK web, the portion of the cytoskeleton that is closely associated with the cytoplasmic surface of the plasma membrane (41). We propose that the observed netlike pattern of H-Ras12V localization is formed by the collapse of the plasma membrane caused by cytoplasm extraction, resulting in decoration of the MSK by the plasma membrane-associated and -associated proteins. Indeed, similar localization pattern has been observed by Cole et al. (42) and Lallemand et al. (43) for the membrane-associated protein merlin. Under conditions, where permeabilization was carried out after fixation, an enrichment of the merlin protein at the plasma membrane was found. When fixation and permeabilization were performed simultaneously, localization pattern of the merlin protein appeared as a network that resembled the cortical actin web.

Regarding DiRas3, our results revealed that after cytoplasm extraction this GTPase localized to the EEA1 (Early Endosome Antigen 1 protein) positive endosomes and to the netlike structures (Fig. 4). The netlike pattern of DiRas3 localization suggests that, similar to H-Ras, DiRas3 is associated with the plasma membrane, which decorates MSK upon cytoplasm extraction. However, in contrast to H-Ras, the netlike pattern of the DiRas3 localization persisted after membranes extraction (Figs. 4A and B), whereas endosomal localization disappeared. These data suggest that DiRas3 located at the plasma membrane is connected to the cortical components of the cytoskeleton. If DiRas3 was coexpressed with H-Ras12V, both proteins colocalized at the plasma membrane-decorated MSK, but not at the Golgi (Fig. 5B). The fact that DiRas3 does not associate with the Golgi is explained by the absence of the palmitoylation sites in DiRas3, which are necessary for plasma membrane targeting of Ras proteins through the Golgi pathway (21,44).

Because the association of H-Ras with the plasma membrane and Golgi requires palmitoylation and farnesylation of H-Ras, respectively, (22) mutation of the corresponding
lipidation sites in H-Ras should alter its subcellular localization. Indeed, the palmitoylation deficient mutant H-Ras12V/181/4SS revealed dramatic redistribution of the staining after cytoplasm extraction (compare Fig. 5A with 5C). The H-Ras12V/181/4SS mutant did not localize to the Golgi and only a very small fraction of the protein accumulated at the plasma membrane-decorated MSK. Most of the H-Ras12V/181/4SS was found in small vesicles distributed throughout the cell, probably representing the collapsed endoplasmic reticulum (Fig. 5C). As expected, the farnesylation deficient mutant H-Ras12V/186S did not associate with the plasma membrane and the endomembranes of the cell and the staining of the H-Ras12V/186S disappeared almost completely after cytoplasm extraction (compare Fig. 5A with 5E). In contrast, when coexpressed with DiRas3, both lipidation mutants of H-Ras12V revealed accumulation at the plasma membrane-decorated MSK where they colocalize with DiRas3 (see Figs. 5D and F). These findings are in line with our coimmunoprecipitation results suggesting that the lipidation deficient mutants of H-Ras were targeted to the plasma membrane by the recruitment through DiRas3.

**DiRas3 Recruits C-RAF to Cytoskeletal Components in H-Ras-dependent Manner.**–Our results of cell fractionation reveal an accumulation of C-RAF within the cytoskeletal fraction upon coexpression with DiRas3 and H-Ras12V. To further investigate these findings, we analyzed the subcellular localization of the C-RAF WT and C-RAF-R89L mutant in the presence and absence of these GTPases by confocal microscopy. When the whole cells were fixed, most of the C-RAF WT localized in the cytosol (Fig. 6A and B, left panels). After the cytoplasm extraction, it became evident that a small portion of C-RAF WT localized at the plasma membrane, whereas most of the membranes-associated C-RAF WT staining was found associated with the EEA1-positive endosomes, in the case that C-RAF WT was expressed alone (Fig. 6C). Exactly the same subcellular localization was observed for C-RAF-R89L mutant (data not shown). Coexpression of C-RAF WT or C-RAF-R89L with DiRas3 did not change the pattern of C-RAF staining (Fig. 6D, the data for C-RAF-R89L are not shown), suggesting that DiRas3 alone does not alter the subcellular localization of C-RAF. In contrast, when the constitutively active H-Ras12V mutant was coexpressed with C-RAF WT, a dramatic redistribution of C-RAF WT from the EEA1-positive endosomes to the plasma membrane-decorated MSK was observed following cytoplasm extraction (Fig. 6E). This observation, together with the fact that the C-RAF-R89L mutant was not redistributed by coexpression with H-Ras12V (data not shown) suggests that the observed relocation of C-RAF and accumulation at the plasma membrane is driven by binding of C-RAF to activated H-Ras. Importantly, upon coexpression with both, DiRas3 and H-Ras12V, C-RAF WT showed a complete colocalization with DiRas3 at the plasma membrane-decorated MSK after cytoplasm extraction (Fig. 6F, left panel). This netlike C-RAF staining and colocalization with DiRas3 persisted even after removal of membranes (Fig. 6F, right panel), revealing an association of both proteins with components of the cytoskeleton. In contrast, the fluorescence signal of C-RAF almost completely disappeared following membranes extraction, if coexpressed with H-Ras12V alone (data not shown). The findings presented in Fig. 6 support our data obtained by immunoprecipitation and fractionation assays (see Figs. 1-3) and indicate that the DiRas3-mediated recruitment of C-RAF to the cytoskeletal components occurs in an H-Ras-dependent manner.

**DiRas3-mediated Regulation of Ras/RAF Signaling is not Limited to H-Ras and C-RAF Only.**–We next addressed the question, whether the DiRas3-mediated regulatory mechanism presented above is limited to H-Ras and C-RAF or it is also valid for other members of the Ras protein subfamily and for other RAF isoforms. Regarding Ras proteins, we tested the validity of the proposed mechanism for K-Ras, as this GTPase differs from H-Ras and N-Ras in its membrane targeting motive lacking the palmitoylation site but containing a polybasic sequence (12,23). The results of coimmunoprecipitation revealed that similar to H-Ras12V (Fig. 2A), K-Ras12V supports the association between C-RAF and DiRas3, as much more C-RAF coprecipitated with DiRas3 in the presence of K-Ras12V (Fig. S1A). Consistently, we observed accumulation of C-RAF within the cytoskeletal fraction upon coexpression with DiRas3 and K-Ras12V (Fig. S1B).

Regarding RAF protein family, we examined the validity of the proposed
mechanism for B-RAF isoform, whose regulation differs significantly from that of C-RAF (4-5). The results presented in Fig. 2A revealed that similar to C-RAF, B-RAF associates with DiRas3 as well, and that this interaction strongly depends on the presence of activated H-Ras. In accordance with this finding, B-RAF accumulated to a high degree within the cytoskeletal fraction, if coexpressed with H-Ras12V and DiRas3 (Fig. S2B). Taken together, these data suggest that the DiRas3-mediated regulation of Ras/RAF signaling is not limited to H-Ras and C-RAF, but is valid for other members of the Ras and RAF protein families.

**DiRas3 Impairs the Dimer Formation of RAF Proteins and Suppresses the Kinase Activity of C-RAF**—As the catalytic activity of RAF proteins is required for the signal transmission from Ras to MEK/ERK, we asked, whether the complex formation between Ras, RAF and DiRas3 may affect the activity of RAF proteins. To address this question we expressed C-RAF-myc-His or B-RAF-myc-His constructs either alone or in combination with H-Ras12V (double transfection) and H-Ras12V/DiRas3 (triple transfection). RAF proteins were isolated by anti-His immunoprecipitation and subjected to an *in vitro* kinase assay using recombinant MEK and ERK as substrates. Surprisingly, the results of this experiment revealed that DiRas3 strongly suppressed the H-Ras12V-induced kinase activity of C-RAF, but not that of B-RAF (Fig. 7A and B). Similar results were obtained for the K-Ras12V-mediated activation of RAF (see Fig. S3). Importantly, the activating phosphorylation of C-RAF at the Ser338 (in B-RAF Ser446) within the regulatory N-region was not impaired by DiRas3 (Fig. 7A and B), thus excluding the possibility that the DiRas3-mediated inhibition of C-RAF activity may occur through the interference with the phosphorylation within the N-region. However, an unexpected effect of DiRas3 expression on the electrophoretic mobility of B-RAF has been observed. As shown in Fig. 7C, the intensity of the shifted B-RAF bands was significantly reduced upon coexpression with DiRas3. This finding suggests that DiRas3 impairs Ras-induced phosphorylation of B-RAF on one or several sites different from Ser446.

These data raised the question: What is the molecular mechanism of the DiRas3-mediated suppression of C-RAF kinase activity? RAF dimerization proved to be one of the most decisive steps in the regulation of RAF activity, especially with respect to C-RAF activation. It has been previously shown that C-RAF is activated by B-RAF through the mechanism involving heterodimerization (45). Therefore, we investigated, whether DiRas3 may affect the dimerization of RAF proteins. To monitor the dimer formation between C-RAF and B-RAF, either C-RAF-myc-His was expressed together with untagged B-RAF (see Fig. 7D) or B-RAF-myc-His was expressed together with HA-tagged C-RAF (see Fig. 7E) in COS7 cells. As Weber et al. (46) reported that dimer formation of RAF proteins is induced by active Ras and the present study revealed that DiRas3 forms a complex with H-Ras, we additionally cotransfected the cells with H-Ras12V and DiRas3. The RAF dimers were isolated by anti-His immunoprecipitation. As shown in Fig. 7D-F, C-RAF dimerized effectively with B-RAF in the presence of active H-Ras12V mutant. In contrast, the Ras-induced dimerization of RAF was strongly impaired (approximately up to 50% reduction, see Fig. 7F) upon coexpression with DiRas3. These data suggest that DiRas3 inhibits the kinase activity of C-RAF through suppression of heterodimer formation between C-RAF and B-RAF.

**DISCUSSION**

Almost 10 years ago Luo et al. (27) reported that the small GTPase DiRas3 interferes with the Ras/MAPK signaling. However, the underlying molecular mechanism has not been further investigated. Data presented in this study reveal that the tumor suppressor protein DiRas3 associates with H-Ras and that activation of H-Ras enforces this interaction (Fig. 1), indicating that the tumor-suppressive activity of DiRas3 may be achieved, at least in part, at the level of Ras effector proteins. Interestingly, our experiments document that DiRas3 interacts with the non-processed form of activated H-Ras, indicating the association of DiRas3 with nascent H-Ras, thus, before the post-translational lipidation process of H-Ras began (see Fig. 1).

Prenylation of the wild type H-Ras is required for its efficient activation by the guanine nucleotide exchange factor SOS, whereas the constitutive active H-Ras12V mutant is independent of SOS-mediated activation and therefore is also active in its non-processed state (17). Moreover, B-RAF has been shown to associate effectively with both farnesylated and non-farnesylated H-Ras and it
has been proposed that activation of B-RAF may take place both at the plasma membrane and in the cytosol (37). Considering these facts, it appears plausible that in order to ensure its tumor suppressive impact at all stages of H-Ras signaling, DiRas3 interacts with the non-processed and active H-Ras. In addition, capturing of the non-processed H-Ras by DiRas3 before the attachment of the farnesyl isoprenoid took place prevents the further palmitoylation of H-Ras and targeting to the plasma membrane microdomains, where the activity of H-Ras is normally required to promote processes such as cell survival and proliferation. Instead, DiRas3 may target H-Ras to the signaling platforms involved in induction of autophagy and apoptosis. There is indeed increasing appreciation that Ras, and other oncogenes, paradoxically induce both pro- and anti-apoptotic signaling and that the balance of positive and negative signals may differ according to kinetics, stoichiometry, availability of different binding partners, and activation of other similar or countervailing forces (for review see (47)). Our results support the view that DiRas3 may relocate H-Ras to the signaling platforms involved in control of the anti-tumorigenic processes. In fact, we observed that DiRas3 was able to recruit the lipidation deficient mutants of H-Ras to the plasma membrane (Fig. 5C-F). DiRas3 itself is post-translationally processed and prenylation of the CAAX box is essential for DiRas3’s membrane association (27). However, the mechanism of the DiRas3’s trafficking to the plasma membrane is still unknown. Considering the fact that there is no palmitoylation site in DiRas3, the translocation of this GTPase to the plasma membrane cannot occur through the Golgi pathway. Our results revealing that DiRas3 does not colocalize with the Golgi-associated H-Ras (Fig. 5B) support this view. It is conceivable that the N-terminal domain of DiRas3, which is unique for this member of the Ras proteins, may contribute to the plasma membrane localization of DiRas3 and may target this GTPase to specific microdomains. The presence of the conserved myristoylation site (see Fig. 2 in (12)) within the N-terminal extension of DiRas3 and the fact that deletion of this DiRas3-specific extension nearly abolishes its inhibitory effect on cell growth (27) corroborate this assumption. In addition, in our study we found that the N-terminal extension of DiRas3 supports its association with H-Ras (Fig. 1A). Concerning the possible binding of DiRas3 to the fully lipidated form of H-Ras, this interaction cannot be excluded so far. The fact that the processed H-Ras12V was not coprecipitated with DiRas3 could be due to the insolubility of the DiRas3 fraction, which may be associated with the lipidated H-Ras at the plasma membrane. Our data exhibiting that most of the plasma membrane located DiRas3 protein is associated with the insoluble cytoskeletal fraction even after membranes extraction (Figs. 3 and 4) might support this assumption.

In addition to the discussed role of DiRas3 in the targeting of H-Ras to particular plasma membrane microdomains, DiRas3 interferes with the effector proteins of H-Ras. In the present study we focused our attention on the Raf kinase, as this effector protein connects H-Ras to the MAPK pathway via direct activation of MEK. Of particular importance are experiments indicating that the effector domain of H-Ras is not involved in the DiRas3/H-Ras interaction (Fig. 1C) and that H-Ras is able to bind to C-RAF while it is associated with DiRas3 (Fig. 2). Moreover, our data document that the complex consisting of DiRas3, H-Ras and C-RAF is more stable than the complexes between H-Ras and C-RAF or H-Ras and DiRas3 (Fig. 2). Presently, it cannot be excluded that also other proteins support the formation of the multimeric DiRas3/H-Ras/C-RAF complex. The formation of large Ras-signaling complexes, which may work as platforms for transducing the Ras signal to effector molecules, has been assumed by several groups. Based on the results of the single-molecule imaging analysis of Ras activation in living cells, Murakoshi et al. (48) proposed a model, in which activated Ras molecules may be bound by Ras-specific scaffolding proteins, which might initiate the cooperative formation of transient signaling complexes including the effector molecules like C-RAF, and deactivating proteins for Ras. This group also reported that the activated Ras molecules (perhaps temporarily) become immobile in the plasma membrane. They concluded that immobilization may be induced by the formation of such a large signaling complex on the plasma membrane, which become connected to the actin-based membrane skeleton mesh (48). This view is supported by our results presented in this study. Our data reveal that DiRas3 colocalizes with H-Ras at the plasma membrane-decorated MSK (Fig. 5B). However, whereas the H-Ras staining almost
disappears, DiRas3 localization at the MSK network structures persists after membranes extraction (Fig. 4A and B). This observation suggests that DiRas3, but not H-Ras, is either directly or indirectly connected to the components of the cytoskeleton. Moreover, our data reveal that DiRas3 induces massive recruitment of C-RAF to the MSK mesh in an H-Ras-dependent manner (Figs. 3 and 6). The recruitment of C-RAF to the cytoskeletal elements has been first reported by Stokoe et al. (40). These authors found that C-RAF, which has been induced to translocate to the plasma membrane by activated Ras, cannot be solubilized with a buffer containing 1% of the detergent Nonidet P-40. They concluded that, once recruited to the plasma membrane, C-RAF becomes tightly associated with the cytoskeletal elements underlying the plasma membrane, but not with the lipid bilayer (40). Our data are in agreement with this model and suggest that the tumor suppressor DiRas3 supports the anchorage of C-RAF to the MSK.

Furthermore, data presented here revealed that additionally to subcellular localization DiRas3 regulates the kinase activity of C-RAF. Although DiRas3 associates in a Ras-dependent manner with both, C-RAF (Fig. 2) and B-RAF (Fig. S2), the results of the in vitro kinase assay clearly show that DiRas3 suppresses specifically the catalytic activity of C-RAF, but not that of B-RAF (Figs. 7 and S3). How could this C-RAF-specific inhibition be explained? The answer to this question is delivered by the results of RAF dimerization assay. We show here, that DiRas3 disrupts the Ras-induced heterodimerization between C-RAF and B-RAF (Fig. 7D-F). Previously, Garnett et al. (45) reported that B-RAF activates C-RAF through the mechanism involving heterodimerization. They have shown that B-RAF can activate C-RAF downstream of Ras, but that C-RAF does not activate B-RAF. In light of these data we propose that DiRas3 inhibits specifically the kinase activity of C-RAF through suppression of heterodimer formation between C-RAF and B-RAF. We also found that expression of DiRas3 affects the phosphorylation status of B-RAF (Fig. 7C). At present, we cannot definitely state, whether the impaired B-RAF phosphorylation is a result of the disrupted C-RAF/B-RAF dimerization or, vice versa, the disrupted C-RAF/B-RAF dimerization is a result of impaired B-RAF phosphorylation. However, we propose that there is interplay between the RAF heterodimerization and the B-RAF phosphorylation status. This suggestion is supported by the studies of Rushworth et al. (49) and Ritt et al. (50), who reported that RAF dimerization is coupled to the ERK-induced phosphorylation of B-RAF. In addition, Heidorn et al. (8) previously showed that B-RAF undergoes a mobility shift in cells treated with the dimerization-inducing RAF inhibitors. Moreover, they suggested that the B-RAF bound to C-RAF is hyperphosphorylated through MEK-ERK-dependent and MEK-ERK-independent mechanisms, but that this phosphorylation is not required for B-RAF binding to C-RAF (8). Considering all these findings, the possibility that DiRas3 reduces hyperphosphorylation of B-RAF by suppressing B-RAF/C-RAF dimerization appears more likely.

Taken together, our considerations raise the question, what is the physiological output of the DiRas3/Ras/RAF interaction. The tumor suppressor DiRas3 has been shown to play a role in the control of cell proliferation, apoptosis and cell motility (31,51), the cellular processes that are also regulated by C-RAF (52-54). One outcome could be that DiRas3 can modify MEK/ERK activity specifically through C-RAF to provide fine tuning of signaling intensity or duration. Alternatively, DiRas3 may utilize the C-RAF pathway to signal to other C-RAF effectors. One common linker that connects C-RAF to the regulation of the quite different processes, such as apoptosis and cell motility, is Rok-α, the kinase involved in the Rho-GTPase-induced rearrangement of the cytoskeleton in migrating cells (55). Furthermore, through its effect on the cytoskeleton, Rok-α regulates clustering and internalization of the death domain-containing receptor Fas, which has a central role in the regulation of the programmed cell death (56). Recently, it has been shown that, in the open state, the C-RAF regulatory domain binds to the kinase domain of Rok-α and inhibits its enzymatic activity directly and independently of C-RAF kinase activity (52,57). Thus, restraining Rok-α is a common molecular basis of the essential function of C-RAF in apoptosis and cell migration. Binding of DiRas3 to the H-Ras/C-RAF complex may interfere with the inhibition of Rok-α by C-RAF, which may
result, depending on physiological context, in altered cell motility and/or induction of apoptosis. Indeed, Badgwell et al. (51) reported that DiRas3 suppresses ovarian cancer cell migration inter alia through inhibition of the FAK/RhoA pathway. However, the underlying molecular mechanism is still unknown. Our study may provide the missing link between DiRas3 and the Rho signaling. DiRas3, which is recruited to C-RAF by active H-Ras, may bind directly to the regulatory domain of C-RAF and prevent its interaction with the kinase domain of Rok-α. A DiRas3-mediated recruitment of other yet unknown protein, which would compete with Rok-α for the binding to C-RAF is also conceivable. The anchorage of DiRas3 to the components of the MSK may deliver this protein to the specific sites of action and avoid undesirable effects.

In conclusion, our data establish a new insight into regulation of Ras/RAF signaling. The finding that DiRas3 forms a complex with the activated Ras and its effector RAF has important biochemical implication. Tumor suppressor functions of DiRas3 are not limited to its antiproliferative activity, but also rely on its combined effects on apoptosis, autophagy and cell migration (27,31,58). Our data suggest Ras/RAF signaling as a common target for the tumor suppressive activity of DiRas3 in all these cellular processes. However, several questions remain to be answered and future studies will be necessary to elucidate in more detail the physiological output of the DiRas3-mediated regulation of Ras/RAF signal transduction.

REFERENCES

Regulation of Ras/RAF signaling by DiRas3


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**FOOTNOTES**

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2The abbreviations used are: CR, conserved region; RBD, Ras binding domain; CRD, cysteine-rich domain; GTP, guanosine 5'-triphosphates; GDP, guanosine 5'-diphosphates; GEF, guanine nucleotide exchange factor; GAP, GTPase-activating protein; ER, endoplasmic reticulum; MSK, membrane skeleton; COS7, African green monkey SV40-transfected kidney fibroblast cell line; WT, wild type; SDS-PAGE, sodium dodecyl sulfate polyacrylamide gel electrophoresis; IB, immunoblot; IP, immunoprecipitation.

**FIGURE LEGENDS**

**FIGURE 1. Binding of H-Ras to DiRas3 does not require H-Ras lipidation or the H-Ras effector domain.** A, myc-tagged DiRas3 wild type or ∆NDiRas3 mutant were transfected into COS7 cells together with either constitutive active H-Ras12V or dominant negative H-Ras17N. H-Ras/DiRas3 protein complexes were isolated from cell lysates by anti-myc immunoprecipitation. The doublet of two bands in the anti-H-Ras immunoblots represents non-farnesylated H-Ras (upper band, indicated as Ras) and farnesylated H-Ras (lower band, indicated as F-Ras) (36). B, H-Ras12V or the indicated H-Ras lipidation mutants were transfected into COS7 cells together with either myc-tagged C-RAF or myc-tagged DiRas3. H-Ras/DiRas3 and H-Ras/C-RAF protein complexes were isolated from cell lysates by anti-myc immunoprecipitation. NM is for non-muted H-Ras12V. C, H-Ras12V, H-Ras17N or H-Ras effector domain mutants were transfected into COS7 cells together with either myc-tagged C-RAF or myc-tagged DiRas3. The protein complexes were isolated as described in B.

**FIGURE 2. C-RAF associates with the H-Ras/DiRas3 complex.** HA-tagged C-RAF wild type (WT) or C-RAF-R89L mutant were transfected into COS7 cells together with H-Ras12V and myc-tagged DiRas3 as indicated in A and B. Protein complexes were isolated from cell lysates either by anti-myc (A) or anti-HA (B) immunoprecipitation. For the bar diagram in B, data from three independent experiments were quantified by optical densitometry. The quantification results are expressed in terms of -fold complex formation, where 1-fold represents the amount of immunoprecipitated H-Ras/C-RAF complexes without DiRas3 co-expression.
FIGURE 3. **C-RAF accumulates within the cytoskeletal fraction upon coexpression with DiRas3.**

A, Western blot analysis of the subcellular distribution of C-RAF, DiRas3 and H-Ras. C-RAF was transfected either alone or together with DiRas3 and indicated H-Ras mutants into COS7 cells. 24 h after transfection, cytosolic (F1), membrane (F2), nuclear (F3), and cytoskeletal (F4) fractions were collected. Anti-M2PK, anti-KDEL, anti-PARP, and anti-vimentin immunodetection was used as fractionation controls for cytosolic, membrane, nuclear, and cytoskeletal fractions, respectively. B, to demonstrate the differences more clearly, the samples from A were separated again by electrophoresis using alternative loading scheme. C, shown is the subcellular distribution of C-RAF-R89L compared to C-RAF WT. D, to demonstrate the differences more clearly, the samples from C were separated again by electrophoresis using alternative loading scheme.

FIGURE 4. **DiRas3 is associated with components of the cytoskeleton.** Myc-tagged DiRas3 was transiently expressed in COS7 cells. After sequential cytoplasm and membranes extraction, cells were fixed and DiRas3 was stained with either anti-DiRas3 (A) or anti-myc antibody (B). Colocalization of DiRas3 with EEA1-positive endosomes (Early Endosome Antigen 1 protein) is shown in C. Arrowheads indicate the endosome-associated DiRas3.

FIGURE 5. **DiRas3 recruits the lipidation deficient mutants of H-Ras12V to the plasma membrane.** H-Ras12V and the indicated lipidation deficient mutants were expressed either alone (A, C, E) or together with DiRas3 (B, D, F) in COS7 cells. After cytoplasm extraction, cells were fixed and the proteins were stained with anti-H-Ras and anti-DiRas3 antibodies as described in Experimental Procedures. Arrowheads indicate the Golgi-associated H-Ras12V.

FIGURE 6. **C-RAF colocalizes with DiRas3 at cytoskeletal components in H-Ras-dependent manner.** Myc-His-tagged C-RAF WT was transiently expressed either alone or together with H-Ras12V and DiRas3 in COS7 cells. After sequential cytoplasm and membranes extraction, cells were fixed and C-RAF was stained with either anti-myc (A, C, E) or anti-His antibody (B, D, F) as described in Experimental Procedures. A and B, C-RAF WT was expressed alone and the localization was proved by two different antibodies as indicated. C, colocalization of C-RAF WT with the EEA1-positive endosomes (Early Endosome Antigen 1 protein) after cytoplasm extraction. D, localization of C-RAF WT in the cytoplasm depleted cells upon coexpression with DiRas3 (the latter was stained by anti-DiRas3 antibody). Arrowheads in A-D indicate the endosomes-associated C-RAF. E, localization of C-RAF WT in the cytoplasm depleted cells upon coexpression with H-Ras12V (the latter was not stained). F, C-RAF WT was expressed together with H-Ras12V (unstained) and DiRas3 (stained with anti-DiRas3 antibody).

FIGURE 7. **DiRas3 disrupts the C-RAF/B-RAF heterodimer formation and suppresses the kinase activity of C-RAF.** Myc-His-tagged C-RAF (A) or B-RAF (B) was expressed either alone or together with H-Ras12V and DiRas3 in COS7 cells. The RAF kinase was immunoprecipitated by anti-His antibody and catalytic activity was analyzed in the in vitro kinase assay using purified MEK and ERK as substrates. Phosphorylation status of C-RAF at Ser338 and B-RAF at Ser446 was analyzed by use of an appropriate phosphospecific antibody. C, B-RAF was expressed in COS7 cells together with H-Ras12V and DiRas3 as indicated. 24 h after transfection, cells were lysed and the proteins separated on the 8% polyacrylamide gel. To examine the impact of DiRas3 on dimer formation between B- and C-RAF, either C-RAF-myc-His, B-RAF and DiRas3-myc (D) or B-RAF-myc-His, HA-C-RAF and DiRas3-myc (E) were transfected into COS7 cells as indicated. Dimerization of RAF proteins was induced by H-Ras12V. RAF dimers were isolated from cell lysates by anti-His immunoprecipitation. F, data from four independent experiments (representative blots are shown in D and E) were quantified by optical densitometry. The quantification results are expressed in terms of % dimer formation, where 100% represents the amount of immunoprecipitated RAF dimer complexes from the cells lacking DiRas3 cotransfection.
Figure 2

A

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B

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Graph showing H-Ras12V/HA-C-RAF(WT) A.U. for Vector and DiRas3 conditions.
Figure 5

A. H-Ras12V
   - Whole cell
   - After cytoplasm extraction

B. H-Ras12V
   - Overview
   - Merge

C. H-Ras12V/181/4SS
   - Whole cell
   - After cytoplasm extraction

D. H-Ras12V/181/4SS
   - Overview
   - Merge

E. H-Ras12V/186S
   - Whole cell
   - After cytoplasm extraction

F. H-Ras12V/186S
   - Overview
   - Merge
Figure 6

A whole cell after cytoplasm extraction
C-RAF (anti-myc) overview close-up view

C-RAF (anti-His) overview close-up view

C-RAF overview close-up view DiRas3 overview close-up view
after cytoplasm extraction after membranes extraction

merge merge
Figure 7

A

DiRas3-myc
H-Ras12V
C-RAF-myc-His

pERK IB
ERK IB
pS338 IB
myc IB
H-Ras IB
DiRas3 IB

kinase assay
with anti-His IP

D

DiRas3-myc
C-RAF-myc-His
H-Ras12V
B-RAF

myc (C-RAF) IB
B-RAF IB
myc (C-RAF) IB
H-Ras IB
DiRas3 IB

anti-His IP

B

DiRas3-myc
H-Ras12V
B-RAF-myc-His

pERK IB
ERK IB
pS446 IB
myc IB
H-Ras IB
DiRas3 IB

kinase assay
with anti-His IP

E

DiRas3-myc
B-RAF-myc-His
H-Ras12V
HA-C-RAF

myc (B-RAF) IB
C-RAF IB
myc (B-RAF) IB
H-Ras IB
DiRas3 IB

anti-His IP

C

DiRas3-myc
H-Ras12V

B-RAF IB
H-Ras IB
DiRas3 IB

cell lysate

F

RAF dimerization [%]

vector DiRas3
The tumor suppressor DiRas3 forms a complex with H-Ras and C-RAF and regulates localization, dimerization and kinase activity of C-RAF
Angela Baljuls, Matthias Beck, Ayla Oenel, Armin Robubi, Ruth Kroschewski, Mirko Hekman, Thomas Rudel and Ulf R. Rapp

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