Structural framework for covalent inhibition of Clostridium botulinum neurotoxin A by targeting Cys\textsuperscript{165}

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Running title: inhibition of C. botulinum neurotoxin A

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Background: development of small potent synthetic inhibitors of Clostridium botulinum neurotoxin A remains an unresolved challenge. Result: Small compounds incorporating electrophile moiety can block enzyme activity by covalent modification of Cys\textsuperscript{165}. Conclusion: a structural framework for developing potent covalent inhibitors of Clostridium botulinum neurotoxin A is provided. Significance: this study uncovers a subfamily of zinc proteases containing a conserved cysteine in their active site.

SUMMARY

Clostridium botulinum neurotoxin type A (BoNT/A) is one of the most potent toxin for humans and a major biothreat agent. Despite intense chemical efforts over the ten past years to develop inhibitors of its catalytic domain (catBoNT/A), highly potent and selective inhibitors are still lacking. Recently, small inhibitors were reported to covalently modify catBoNT/A by targeting Cys\textsuperscript{165}, a residue located in the enzyme active site just above the catalytic zinc ion. However, no direct proof of Cys\textsuperscript{165} modification was reported and the poor accessibility of this residue in the X-ray structure of catBoNT/A raises concerns about this proposal. To clarify this issue, the functional role of Cys\textsuperscript{165} was first assessed through a combination of site-directed mutagenesis and structural studies. These data suggested that Cys\textsuperscript{165} is more involved in enzyme catalysis rather than in structural property. Then by peptide-mass fingerprinting and X-ray crystallography, we demonstrated that a small compound containing a sulfonyl group acts as inhibitor of catBoNT/A through covalent modification of Cys\textsuperscript{165}. The crystal structure of this covalent complex offers a structural framework for developing more potent covalent inhibitors catBoNT/A. Other zinc metalloproteases can be found in the protein data base with a cysteine at a similar location, some expressed by major human pathogens, thus this work should find broader applications for developing covalent inhibitors.

The anaerobic spore-forming bacteria Clostridium botulinum produces the most potent neurotoxins known. These toxins, the causative agents of botulism, impair the release of acetylcholine from presynaptic nerve terminals at neuromuscular junctions through specific proteolysis of essential SNARE proteins (1,2). The Centers for Disease Control and prevention have classified these toxins in category A of biowarfare agents. Among the seven serotypes of botulinum neurotoxins so far described, serotype A (BoNT/A) specifically cleaves the synaptosome-associated 25 kDa protein (SNAP-25) (3). BoNT/A is synthesized as a 150 kDa single polypeptide chain, which is then cleaved by endogenous proteases to give the two chains structure consisting of a 50 kDa light chain linked by a single disulfide bond to a 100 kDa...
heavy chain. The heavy chain is responsible for specific cell surface receptor interactions and the light chain is a zinc metalloprotease that cleaves SNAP-25. Disulfide bond reduction is essential for allowing free access of SNAP-25 to the light proteolytic active site (4). Since BoNT/A is the most potent neuron-blocking toxin, major research efforts have focussed on its catalytic domain (catBoNT/A) to design inhibitors able to counter toxin induced post-neuronal internalization. Despite intense efforts over the past ten years, the small inhibitors developed so far are inadequate for in vivo treatment, thus more effective inhibitors are still urgently needed (5). Rather potent peptide-based inhibitors that have been reported, but their metabolic stability may limit their efficacy in blocking BoNT/A within neurons (6-9). Several non-peptidic compounds exhibiting sub-micromolar affinity have also been reported, but evaluation of their in vivo potency needs further improvement (10-14). The flexibility of several loops around the active site of catBoNT/A are believed to be the factor that limits inhibitor development, because of the strong entropic penalty that accompanies inhibitor binding (11,15). Thus, alternative strategies might be necessary for developing more potent inhibitors of BoNT/A.

Covalent inhibitors could be an alternative strategy to the non-covalent ones pursued to date if a nucleophilic amino acid within the active site of catBoNT/A could be found. Looking in the active site of catBoNT/A reveals the presence of a cysteine residue (Cys165) located nearby the catalytic zinc ion (Fig1). Nucleophilic residues involved in catalysis have been successfully targeted by active-site irreversible inhibitors, leading to the development of potent inhibitors, such as serine and cysteine proteases (16). Such inhibitors contain in their structure reactive electrophilic moieties selected for covalent modification of these nucleophilic residues. Targeting non-catalytic nucleophiles present in enzyme cavities is a variation of the same strategy, that can be used to develop irreversible “targeted covalent inhibitors” (17). A large variety of electrophilic groups have been proposed and some of them have been validated for in vivo applications, leading to irreversible inhibitors with potential therapeutic values for a variety of indications (18). Recently, acrylonitrile compounds were reported to inhibit catBoNT/A through covalent modification of Cys165, but no direct proof of this modification was provided (19). This result and the above considerations led us to check if Cys165 in catBoNT/A can be the target of covalent inhibitors and evaluate their potential functional role in enzyme catalysis.

**EXPERIMENTAL PROCEDURES**

**Expression and purification of BoNT/A catalytic domain.** Gene encoding the catalytic domain of the BoNT/A (Met1–Phe425) was obtained from the Pasteur Institute (Dr. M. Popoff, Pasteur Institute Paris). This gene was inserted into the pET28a+ vector (Novagen), between the NdeI and SalI site, generating a thrombin cleavable N-terminal 6-His affinity tag. C134Ser mutant was produced from 5'-gttattgatactaattgaAttaatgtgatacaaccagatgg-3' and 5'-caataactatgattaaGataattacactatgttggtctacc-3' oligonucleotides, and C165S was produced from 5'-cagctgatattatacagtttgaatCtaaaagctttggacatgaagtttt-3' and 5'-aaaacttcatgtccaaagcttttaGattcaaactgtaatacatcgctg-3' using the Site-Directed Mutagenesis Kit (Quick change II). All plasmids were propagated in the Escherichia coli strain XL1-Blue at 37 °C, and all constructions were verified by DNA sequencing using the ABI PRISM 310 Genetic analyzer (Applied Biosystems). Recombinant proteins were expressed in E. coli BL21 (DE3 star) cells carrying the BoNT/A catalytic domain-encoding plasmids. C134S/C165S double mutant was produced with the same procedure, introducing the C165S mutation on the C134S mutant.

E. coli BL-21 (DE3) containing BoNT/A expression plasmids were grown overnight on Luria Broth agar (LB) with 50µg/ml kanamycin. Cells were inoculated into fresh LB medium containing antibiotic, grown at 28°C for 4h at 175 rpm, at absorbance (600nm) of 0.6. Protein expression was induced by addition of 1 mM IPTG (isopropyl-β-thiogalactopyranoside), and then cell cultured at 175 rpm overnight at 16°C. Cells were grown in 2 or 4 L of LB, and harvested cells were then passed through a cell disruption system at 4°C (Constant Systems Ltd., Daventry Northants, England) in 40 ml ice-cold buffer A (10 mM imidazole, 500 mM NaCl, and 20 mM Tris-HCl, pH 7.9) for 1L the culture
containing PMSF, benzonase, 10 mM MgCl$_2$ and 10 mg/ml lysozyme. The lysate was clarified by centrifugation at 18,000 rpm for 45 min at 4°C and subsequently passed through a 0.22 µm filter. The filtered lysate was loaded onto a column of His Trap HP 5 ml that had been equilibrated with 25 ml buffer A. The column was washed with 10% buffer B (500 mM imidazole, 500 mM NaCl, and 20 mM Tris-Cl, pH 7.9) and eluted with 50% buffer B. The filtered lysate was loaded onto a column of His Trap HP 5 ml that had been equilibrated with 25 ml buffer A. The column was washed with 10% buffer B (500 mM imidazole, 500 mM NaCl, and 20 mM Tris-Cl, pH 7.9) and eluted with 50% buffer B. Peak fractions were pooled and dialyzed overnight in 5L buffer (10 mM Tris-HCl, 20 mM NaCl, pH 7.8) and clarified by centrifugation at 18,000 rpm for 30 min at 4°C. The solution dialyzed, proteins were loaded onto a column His Trap Q 5ml, that equilibrated with buffer C (10 mM Tris-Cl, 20 mM NaCl, pH 7.8) and eluted with a linear gradient of 0.02-1M NaCl. Fractions of 2 ml, were analysed by 12% SDS-PAGE. Purified proteins were aliquoted and then stored at -80°C. Purified proteins were aliquoted and then stored at -80 °C. Full form of protein containing the His tag and the thrombin cleavage was observed to display better activity when conserved at 4°C, than protein form in which the His-tag was removed by thrombin cleavage, we thus decided to perform all studies with Histag containing protein.

**Enzyme kinetics.** Enzyme catalytic efficiency assays were performed in in 40mM Hepes, pH 7.4 at 37°C. Substrate concentration (Mca-S-N-K-T-R-I-D-E-A-N-Q-R-A-T-K-Nle-Dap(Dnp)-NH$_2$) was determined by amino acid analysis. Substrate was prepared as 10 mM stock solution in dimethylsulfoxide. Enzyme concentration was determined spectrophotometrically using $\varepsilon_{280nm} = 43527$ M$^{-1}$ cm$^{-1}$. Individual kinetic parameters $k_{cat}$ and $K_m$ were obtained from analysis of fluorescence progress curves, under steady-state rate conditions, over 0.2 to 5 $K_m$ substrate concentration ranges. The experiments were carried out in black 96-well plates (nonbinding-surface plates 3650; Corning Costar). Progress curves were monitored by following the increase in fluorescence at 405 nm ($\lambda_{ex} = 320$ nm), induced by the cleavage of the synthetic substrate by BoNT/A enzyme, using a photon counter spectrophotometer (Fluoroskan Ascent; Thermo-Labsystems). $K_m$ and $k_{cat}$ values were determined by progress curves fitting using DYNAFIT Program (20).

**Enzyme inhibition.** The rate of irreversible inhibition of BoNT/A by MTSEA and MTSPA was followed by withdrawing samples for assay at several time intervals after the mixing of enzyme and inhibitor (HEPES buffer 40mM at pH7, 37°C), with $[I] >> $ to enzyme concentration. A plot in logarithm of residual activity against time gives the observed rate of inactivation $k_{obs}$ using the equation (1). Assuming that the inactivation proceeds by the rapid formation of the non covalent enzyme:inhibitor complex, followed by a slower step leading to covalent enzyme modification, the Kitz and Wilson equation was used in its linearized form (2) to deduce $K_i$ and $k_{inact}$ values (21). Enzyme assays were performed as described for enzyme kinetic studies.

1. $\ln(\text{residual. act}) = -k_{\text{inact}} t$

2. $1/(k_{obs}) = (K_i/k_{\text{inact}}) 1/[I] + 1/(k_{\text{inact}})$

**Cleavage of SNAP-25.** Cleavage of recombinant full length human SNAP-25 (isoform B, 206aa, 23 kDa from Abcam, Ref ab74529) (5µM) was initiated by adding BoNT/A or serine mutants (5 or 10 nM) in 40 mM Hepes, pH 7.4 at 37°C. After 1h, the reaction was stopped by adding 1µL TFA 50% water and SDS-PAGE sample buffer. The products were resolved by 12% SDS-PAGE gel and protein bands revealed by silver staining. The extent of SNAP-25 cleavage was determined by densitometric scanning of the band corresponding to the product (ImageJ program).

**CD- spectroscopy.** CD spectra were recorded at 25°C on a JACSO J-810 spectropolarimeter equipped with a Peltier type temperature controlled system. Proteins were equilibrated in sodium phosphate buffer 10 mM, pH 7.5. UV spectra were recorded to compare CD spectra of wild type and mutants at similar protein concentration (5 µM and 15 µM for far-UV and near-UV region, respectively)(Figure 1S and 2S). Samples were prepared exchanging conservation buffer by sodium phosphate buffer 10mM pH 7.5 with PD-spin trap G-25 and diluting the sample in sodium phosphate buffer until the desired protein concentration was obtained. Thermal denaturation was performed by monitoring the CD signal at 222 nm from 20
to 60 °C.

**MTSEA and MTSPA adduct formation by mass-spectrometry analysis.** 1ml of a solution containing BoNT/A 16 µM and 32 mM of MTSEA or MTSPA (HEPES buffer 40mM at pH7, 37°C) was diluted in 19 mL of a 50mM ammonium bicarbonate solution to which was added 0.6 mL of trypsin 400 ng/ml. Samples were heated at 50 °C for 2h. 2 µL of TFA were added to stop the enzymatic reaction. The resulting digest (0.5 µL) was manually spotted on MALDI plate with an equal volume of HCCA (R-cyano-4-hydroxycinnamic acid) matrix solution prepared at 10 mg/mL in H2O/CH3CN/TFA (50/50/0.4). MS spectra were recorded from crystallized samples. The DataExplorer software (4.9) from ABI was used to generate ASCII peak lists from peptide mass fingerprinting MS analyses. Each peak list was manually applied for searches using MASCOT software (www.matrixscience.com) in the NCBInr database updated on Dec 2008. The parameters used for the search were as follows: a taxonomy restriction was placed to Homo sapiens (human), one missed trypsin cleavage was allowed, a maximum mass tolerance was set at 20 ppm because of an internal calibration made on fragments resulting from trypsin autolysis for all MS analyses and methionine oxidation was set as a variable modification. MASCOT protein sift Mowse score greater than 66 (assuming p < 0.05) were considered significant.

**Crystallization.** The crystallization of the catalytic domain of neurotoxin wild type BoNT/A and C134S/C165S double mutant were screened under sitting drop vapor diffusion using ChemCryst Plates (Hampton Research) with a limited number of conditions from the “Stura” screens (22) (Molecular Dimensions) with a protein solution consisting of 7.8 mg/mL in 10mM Tris-HCl, 150 mM NaCl pH 7.8 for the Ser134/Ser165 double mutant and 2.88 mg/mL in 0.01 M Tris-HCl, 0.15 M NaCl, pH 7.8 for the wild type at 20° C in a cooled incubator. Good results were obtained under different polyethylene glycol (PEG) conditions and various pH. The crystal optimization strategy was to use a working condition (methodology described in (23)) consisting of 36% mono-methyl PEG 2000 (MPEG-2K), 50 mM Li2SO4 to which various buffers at different pH were applied. Optimized conditions consisted of 80% working solution with 100 mM imidazole-HCl pH 6.0 in the reservoir. Drops consisted of 1µL of protein solution to which 1µL of reservoir solution was added. Seed crystals were added with a capillary-syringe combination to the drop and allowed to grow (methodology described in (24)). For flash-cooling 18% MPEG-2K, 22% MPD, 10% DMSO, 50 mM Bicine, pH 7.5 was used as the cryoprotectant. After a short solvent exchange step, the crystals were cryo-cooled in liquid ethane. The purified MTSEA-modified catalytic domain of C134S mutant of catBoNT/A was at 3.0 mg/ml in 10 mM Tris-HCl pH 7.9, 50 mM Li2SO4. Crystallization of the MTSEA-modified protein was more complicated, thus larger 3 µl drops were used as well as two separate working solutions for addition to the protein drop and for use in the reservoir. This was part of the strategy to achieve a higher degree of protein concentration by vapor diffusion. Thus 3 µl of 24% MPEG-550, 50 mM Li2SO4, imidazole malate pH 5.5 was added to the protein drop which was then equilibrated with a reservoir consisting of 42% PEG MPEG-550, 100 mM Li2SO4, 200 mM imidazole malate, pH 5.5. Seeding was used as for the double mutant. For flash-cooling, the same cryoprotectant was used, but after a short solvent exchange step the crystals were cryo-cooled in liquid nitrogen. Crystallization details are summarized briefly in Table S1.

**Data collection and crystal structure determination.** The cryo-cooled crystals were transported to the European Synchrotron Radiation Facility (ESRF) in Grenoble in accordance with the safety requirements of the Agence Française de Sécurité Sanitaire des Produits de Santé (afssaps). Data for crystals of the catalytic domain of neurotoxin BoNT/A and C134S/C165S double mutant were collected on beamline ID14-4 at 100 °K from single crystals to 1.87 and 1.2 Å resolution respectively. The crystal was found to belong to the space group P21 with cell parameters 49.3Å, 66.3Å, 64.9Å, β=99.1° (Table S1). For the MTSEA-modified single mutant data were collected on the same beamline at a later date. The crystals also belong to the tetragonal space group P4_2_2 with cell parameters a=b=65.3 Å, c=200.9 Å diffracting to 1.8 Å resolution. In both cases there is only one
molecule in the asymmetric unit. The data were processed using MOSFLM (25) and reduced using programs from the CCP4 suite of programs. Molecular replacement was carried out with MOLREp (26) using as the model an inhibitor complex of the same protein with similar crystal parameters (PDB code=2ILP (11)) as the starting model. After restrained refinement using REFMAC (27) the amino acid differences between the two loops were corrected according to the sequence. Density fitting and refinement were carried out with the aid of electron density maps (omit σA-weighted 2Fo- Fc and Fo-Fc) calculated and displayed using the XtalView (28 suite of programs and COOT (Emsley, #42). Stereochemical analysis of the final refined model was checked with the validation tools in COOT. Waters were checked with phenix.refine (29). Refinement statistics are detailed in Table S1. The figures were made with PYMOL from DeLano Scientific LLC (The pyMOL Molecular Graphics System. DeLano Scientific, San Carlos, CA, USA).

Protein data bank accession numbers. The coordinates and structure factors for the catalytic domain of neurotoxin BoNT/A C134S/C165S double mutant and single mutant C134S-MTSEA covalent adduct have been deposited in the RCSB Protein Data Bank with the following ID codes: 4EJ5, 4EL4 and 4ELC, respectively.

RESULTS

Functional role of Cys^{165} in catBoNT/A.

To assess the potential functional role of Cys^{165}, we expressed a recombinant version of the wild type and serine-substituted catBoNT/A. This catalytic domain possesses two free cysteines, one close to the catalytic zinc ion (Cys^{165}) and the other (Cys^{134}) far away from the active site (Fig. 1). Cys^{134} is located inside an exosite involved in the binding of SNAP-25, the natural protein substrate of BoNT/A. Impact of the replacement of these cysteine residues by serine was first evaluated by comparing the efficiency of wild type enzyme and single cysteine mutants 134S/C165S double mutant at 5 and 10 nM (see experimental section). As compared to wild type enzyme under the same conditions, the serine mutants performed clearly worse (Table 1, Fig. S1). To obtain more quantitative data, the catalytic efficiency of wild type and mutant enzymes was assessed with a synthetic fluorogenic peptidic substrate. Both the wild type and the C134S mutant displayed similar catalytic efficiencies (k_{cat}/K_{M}) in cleaving the synthetic substrate (Table 2). This is in agreement with the fact that this synthetic substrate is much more shorter than SNAP-25, thus its binding to catBoNT/A does not involve the exosite containing Cys^{134} (19). In contrast, a 50-fold reduction in catalytic efficiency was observed for the C165S mutant relative to the wild type. This substantial reduction in activity can be imputed to a marked decrease in k_{cat} value (0.007 s^{-1}), when compared with the wild type enzyme (k_{cat} = 0.43 s^{-1}). Similar values were obtained for the C134S/C165S double mutant (Table 2).

The above data suggest that Cys^{165} plays a key role in the catalytic efficiency of catBoNT/A when cleaving both natural and synthetic substrates. To gain more insights into these findings, X-ray structures of the wild type and the C134S/C165S double mutant were determined at 1.8 and 1.2 Å, respectively. Overlay of both structures shows no significant change at the active site or the exosite (Fig. 2). Consistently, CD spectra recorded in the near and far UV regions for the wild type enzyme and the mutants were similar (Fig. S2, S3). The thermal stability was also probed by monitoring changes in ellipticity at 222 nm as a function of temperature to assess the thermal denaturation pattern. All proteins exhibited similar single sigmoidal transitions with identical slopes, with a mid point of thermal transition Tm equal to 42 °C (Fig. S4). Taken together, these structural data revealed that the mutation of Cys^{165} did not induce detectable changes in the enzyme structure or active site, suggesting rather a direct role of Cys^{165} in catalysis.

Implication for inhibitor design

The crystal structure of BoNT/A shows that the side chain of Cys^{165} is poorly accessible to solvent (Fig. 1), thus potentially limiting its ability to be covalently modified by electrophiles. To test this, the C134S mutant, which still contains a cysteine at position 165, was incubated with (2-aminoethyl) methanethiosulfonate (MTSEA). MTSEA was selected because it contains a highly reactive sulfonyl moiety and displays a high reactivity...
and selectivity towards cysteine (30) (scheme 1).

\[
\text{MTSEA} \quad \text{MTSPA} \\
\text{O} \quad \text{O} \quad \text{O} \quad \text{O} \\
\text{S} \quad \text{S} \quad \text{NH}_2 \quad \text{NH}_2 \\
\text{Enz-SH} \quad \text{Enz-SH} \\
\text{Enz-S-S-NH}_2 \quad \text{Enz-S-S-NH}_2 \\
\text{Scheme 1}: \text{chemical structure of MTSEA and MTSPA compounds and chemical reaction with cysteine.}
\]

After incubation catBoNT/A with MTSEA, the enzyme activity was drastically reduced. Exhaustive dialysis did not rescue enzyme activity, suggesting that the enzyme has been irreversibly inactivated through covalent modification of Cys 165. A similar experiment with the C165S mutant revealed no inhibition, supporting this proposal. Monitoring the time-dependent loss of catBoNT/A when incubated in presence of different MTSEA concentrations for various time allows to determine the rate of inactivation \( k_{\text{obs}} \) as function of MTSEA concentrations (see experimental procedures). From these data, using equation 2, the kinetic parameters for the inactivation of catBoNT/A by MTSEA can be derived (Figure 3 and Table 3).

After trypsin digestion of the C134S mutant pre-incubated with MTSEA, mass spectrometry analysis of the resulting peptides revealed a mass increment of 74 Da for the fragment of catBoNT/A containing Cys 165, which is consistent with a Cys-MTSEA disulphide adduct (Fig. S5A). The X-ray structure of the C134S mutant modified with MTSEA determined at 1.72 Å resolution confirms this finding. Interestingly, this structure revealed that the amino group of MTSEA was pointing toward Glu 224 and that a malate molecule from the crystallization buffer was chelating the zinc ion (Fig. 4). From this structure, we speculated that the longer MTSPA compound (scheme 1), with an additional methylene, might increase inhibitor potency by allowing its NH\(_2\) group to better interact with Glu 224 or the zinc ion. MTSPA was also found to irreversibly inhibit catBoNT/A (Figure 3) and kinetic analyses confirmed that MTSPA is a better inhibitor than MTSEA (Table 3), mostly due to increase in affinity (MTSPA, Ki value of 7.7 \( \mu \)M; MTSEA, Ki= 625 \( \mu \)M).

MTSPA is a rather potent of BoNT/A, as after 2h of incubation it displays an \( IC_{50} \) value of 260 nM. Mass spectrometry analyses confirmed that MTSPA covalently modified Cys 165 (Fig S5B). Similar results were also obtained with the wild type enzyme, with the two compounds displaying \( IC_{50} \) values comparable to those obtained for the C134S mutant.

**Metallopeptidases with cysteine residue in their active site.**

Systematic analysis of the protein databank revealed that other zinc proteases likewise have a cysteine in a similar position to BoNT/A in their active sites. In the majority of zinc peptidases, a HEXXH signature is found, in which the two histidines bind the active site metal ion and the glutamate acts as the general base/acid during catalysis. These key active site residues WERE USED to perform the superimposition of available 3D-structures (Fig. 5). Of the seven proteases identified in the PDB, six contained a zinc ion in their active sites and one a cobalt. The superposition confirmed not only the presence but also the topological equivalence of the cysteine site in all these structures, thus pointing to a common functional role. These peptidases are expressed by different kingdoms of life: eukaryotes with mammalian thimet oligopeptidase (PDB, 1S4B) and neurolysin (PDB, 1I1I); prokaryotes (Clostridium botulinum neurotoxins type A (PDB, 1XTG) and type F (PDB, 2A97) and oligopeptidase F (PDB, 2H1J) from Bacillus steatoroathermophilus); and protists (Trypanosoma cruzi metallocarboxypeptidase (PDB, 3DWC)). All these metallopeptidases belong to the “cowrin family” according to the proposed nomenclature of Gomis-Ruth et al (31). However, not all members of the cowrin family possess this cysteine residue. For example, in mammalian angiotensin-converting enzyme, structurally related to thimet oligopeptidase and neurolysin, a serine is observed at the topological equivalent position to the cysteine and in T. thermophilus carboxypeptidase (PDB, 1WGZ), closely related to Trypanosoma cruzi metallocarboxypeptidase (32), a glutamate residue is observed at this site.
DISCUSSION

The presence of a cysteine close to the active-site metal ion in a subset of metalloenzymes whose side chain is not pointing toward the solvent from crystal structure is intriguing. The present study reveals that in catBoNT/A Cys\textsuperscript{165} is not playing a key structural role but contributes to in enzyme activity. The 50-fold reduction in catalytic efficiency of the C165S mutant in cleaving a synthetic substrate is comparable to that reported for other mutants of catBoNT/A, Arg\textsuperscript{231} (33), Arg\textsuperscript{362} and Tyr\textsuperscript{365} (34), three residues involved in the stabilization of the transition-state. These mutations rendered a reduction in $k_{\text{cat}}$ (65-fold for Arg->Ala\textsuperscript{231}, 84-fold for Arg->Ala\textsuperscript{363} and 35-fold for Tyr->Phe\textsuperscript{365}), close to the 60-fold reduction in $k_{\text{cat}}$ observed for the C165S mutant of catBoNT/A. This mutation also greatly affects the cleavage of a natural substrate, suggesting a functional relevance for this result. Combining experimental approaches, we conclude that the effect of the Cys\textsuperscript{165}->Ser mutation on enzyme activity does not result from structural perturbations but rather from the suppression of sulfur chemistry. Without information on the structure of reaction-intermediates involved in substrate cleavage, the precise role of Cys\textsuperscript{165} in enzyme activity is difficult to assess. However, it should be kept in mind that a cysteine residue, particularly one located at the protein surface like Cys\textsuperscript{165}, is highly polarisable and carries a terminal dipole with a pronounced partial negative charge. These are properties that render cysteine able to perform multiple interactions and to change its reactivity when the local pH is altered within its micro-environment (35). In this respect, the short distance between Cys\textsuperscript{165} and Arg\textsuperscript{231} (Cys\textsuperscript{165} Sg – Arg\textsuperscript{231} Ne, 4.9 Å), a residue involved in enzyme catalysis, may suggest a possible concerted action between these two residues during catalysis. To further assess the possible concerted action between these two residues during catalysis. From crystal structure of catBoNT/A, we retrieved all sequences for serotype A and F orthologues found in different clostridial strains to assess conservation. For BoNT/A, all 18 sequences identified contained the cysteine, while only 6 out of 16 BoNT/F sequences exhibited a cysteine (FigS6). Obviously, better understanding the exact role of Cys\textsuperscript{165} in enzyme catalysis will require additional studies.

From crystal structure of catBoNT/A, the reduced accessibility of the Cys\textsuperscript{165} sulfur atom could have limited its covalent modification by electrophilic groups. The successful covalent modification of Cys\textsuperscript{165} thus suggests the existence of some local “flexibility” permitting the sulfonate reactive group of MTSEA and MTSPA to reach and react with the Cys\textsuperscript{165} thiol group. The present work also supports the covalent modification of Cys\textsuperscript{165} by acrylonitrile as recently speculated(19). The most potent inhibitor of catBoNT/A reported to date is a pseudo-heptapeptide, which exhibits a $K_{i}$ value of 41 nM (7); in addition, many low-molecular-weight inhibitors have been reported to evalue sub-micromolar potency (5). Instead of increasing the size of the non-covalent inhibitors to further optimize interactions with enzyme active site, the present study reveals on a structural basis how small covalent inhibitors of catBoNT/A can be developed. To design irreversible inhibitors, the availability of detailed structural information is critical for positioning the electrophilic moiety of the inhibitor correctly, so that it can attack the nucleophile present in the enzyme active site efficiently and yield the desired covalent modification. Compared to non-covalent inhibitors, the potency and selectivity of irreversible inhibitors can be optimized in two ways, by modulating the binding capacity of the non-reactive part of the inhibitor and by modifying the reaction rate between the nucleophile and the electrophile. When properly designed, irreversible inhibitors can be exceptionally potent and selective because part of the binding energy is derived from protein-inhibitor covalent bond (36) (17). Thus, in a context where programs based on non-covalent inhibitor development have been disappointing, developing of covalent inhibitors may represent a new avenue. The crystal structure of BoNT/A-MTSEA complex is a very good starting point for designing libraries of small compounds, containing on one side various electrophilic groups and on the other different zinc-chelating groups. In this structure, an L-malic acid molecule is making interactions with the amino group of MTSEA, the zinc ion and residues Glu\textsuperscript{224} and Tyr\textsuperscript{366}. From these observations, inhibitors can also be designed by “connecting” the L-malic acid to electrophiles, a classical approach in fragment-based design (37). Using similar structure-based design, a very potent covalent inhibitor of HCV protease has been recently reported (38). For the purpose of this
study, we have selected as electrophile a sulfonyl group which is too reactive in context of complex proteome or cell systems, thus in the future less reactive electrophiles like acrylonitrile would have to be chosen for developing useful covalent inhibitors of BoNT/A.

Analysis of the PDB reveals that other zinc peptidases likewise exhibit a cysteine in their active sites and that the topology of these cysteine residues is very well conserved (Fig. 5). While in catBoNT/A, Cys165 is close to Arg230, a structural context that may influence the reactivity or function of Cys165, in other analysed proteins there is not an arginine-equivalent that would lead to a modulated cysteine reactivity. Whether the conserved cysteines in these proteases are also involved in enzyme catalysis needs to be clarified. Whatever the exact roles of these cysteines are, their use to develop targeted covalent inhibitors for these will only depend on their reactivity and flexibility. Among the most interesting potential targets are a family of zinc carboxypeptidases expressed by pathogens like Vibrio cholerae, Yersinia pseudotuberculosis and peptis, Leismania major and Trypanosoma cruzi. Thus, beyond BoNT/A inhibitors, the results reported here should stimulate the development of targeted covalent inhibitors of these carboxypeptidases.
REFERENCES


**Footnotes**

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**Figure Legends**

Figure 1: details of the active site of the catalytic domain of BoNT/A; residues chelating the zinc ion are displayed in cyan, residues reported to play a role in enzyme catalytic efficiency are displayed in green, cysteine residues in yellow and the catalytic zinc ion as magenta sphere.

Figure 2: superimposition of crystal structure between wild type catBoNT/A catalytic domain (1.8 Å) and C134S, C165S double mutant (1.2 Å). A closer view of the peptide segment bearing Cys^{165} in wild type (yellow) or C134S, C165S in double mutant (cyan) is shown in stick representation.

Figure 3: Kitz-Wilson plot for the inactivation of catBoNT/A by MTSEA (cross) and by MTSPA (closed circle). HEPES buffer 40mM at pH7, 37°C.

Figure 4: crystal structure of catBoNT/covalently modified by MTSEA. This structure reveals the NH$_3$ group of MTSEA is surrounded by Glu$^{220}$, the zinc ion and a malate molecule from the crystallisation buffer, interacting itself with the zinc ion.

Figure 5: Superimposition of the crystal structure of clostridium botulinum neurotoxin type A (PDB, 1XTG) with that of clostridium botulinum neurotoxin type F (PDB, 2A97), thimet oligopeptidase (PDB, 1S4B), neurolysin (PDB, 1IIl), oligopeptidase F (PDB, 2H1J) and trypanosoma cruzi metallocarboxypeptidase (PDB, 3DWC). Structure overlay was based on the best fit between the zinc ligand (His, His and Glu) in these proteases.

**Table 1:** % of SNAP-25 cleavage by wild type catBoNT/A catalytic domain and serine mutants. Experiments were performed in Hepes buffer 40mM, pH 7.4 at 37°C.

**Table 2:** kinetic parameters for the cleavage of the fluorogenic substrate (5µM) by wild type catBoNT/A catalytic domain, C134S mutant and C165S mutant (20 nM). Assays were carried out in Hepes 40mM, pH 7.4 at 37°C.

**Table 3:** kinetic constant for the inactivation of catBoNT/A catalytic domain by MTSEA and MTSPA. Experiments were performed in Hepes 40mM, pH 7.4 at 37°C and the kinetic constants were determined from plots reported in figure 3.
Figure 3
Figure 4
Table 1

<table>
<thead>
<tr>
<th></th>
<th>% of SNAP-25 cleavage</th>
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<tbody>
<tr>
<td></td>
<td>5 nM of catBoNT/A</td>
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<tr>
<td>Wild type</td>
<td>50%</td>
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<tr>
<td>C134S</td>
<td>10%</td>
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<tr>
<td>C165S</td>
<td>&lt; 10%</td>
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Table 2

<table>
<thead>
<tr>
<th>enzyme</th>
<th>$K_m (10^{-6}$M)</th>
<th>$k_{cat}$ (s$^{-1}$)</th>
<th>$k_{cat}/K_m (M^{-1}.s^{-1})$</th>
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<tbody>
<tr>
<td>Wild type</td>
<td>68 ± 13</td>
<td>0.54 ± 0.07</td>
<td>7948</td>
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<tr>
<td>C134S</td>
<td>85 ± 11</td>
<td>0.54 ± 0.05</td>
<td>6430</td>
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<tr>
<td>C165S</td>
<td>76 ± 15</td>
<td>0.014 ± 0.001</td>
<td>190</td>
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<tr>
<td>C134S/C165S</td>
<td>35 ± 7</td>
<td>0.007 ± 0.008</td>
<td>220</td>
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</table>

Table 3

<table>
<thead>
<tr>
<th>inhibitor</th>
<th>$k_{max}$ (s$^{-1}$)</th>
<th>$K_i$ (mM)</th>
<th>$k_{max}/K_i (M^{-1}.s^{-1})$</th>
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</thead>
<tbody>
<tr>
<td>MTSEA</td>
<td>0.041</td>
<td>625</td>
<td>65</td>
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<tr>
<td>MTSPA</td>
<td>0.0034</td>
<td>7.7</td>
<td>440</td>
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Structural framework for covalent inhibition of Clostridium botulinum neurotoxin A by targeting Cys165
Enrico A. Stura, Laura Le Roux, Karine Guittot, Sandra Garcia, Sarah Bregant, Fabrice Beau, Laura Vera, Guillaume Collet, Denis Ptchelkine, Huseyin Bakirci and Vincent Dive

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