

Endoplasmic Reticulum Stress-induced Degradation of DNAJB12 Stimulates BOK Accumulation and Primes Cancer Cells for Apoptosis

Pattarawut Sopha^{#*^}, Hong Yu Ren^{*}, Diane E. Grove^{*}, and Douglas M. Cyr^{*^}

^{*}Department of Cell Biology and Physiology, University of North Carolina at Chapel Hill, Chapel Hill, NC 27599, USA

[#]Program of Applied Biological Sciences, Chulabhorn Graduate Institute
54 Kamphaeng Phet 6, Talat Bang Khen, Lak Si, Bangkok 10210, Thailand

[^]Address correspondence to: Douglas M. Cyr (dmcyr@med.unc.edu) or Pattarawut Sopha (pattarawut@cgi.ac.th)

Abbreviations: BOK: BCL-2 ovarian killer; Bort: bortezomib; DTT: Dithiothreitol; ER: endoplasmic reticulum; ERQC: ER protein quality control; ERAD: ER-associated protein degradation; Tg: Thapsigargin; Tm: Tunicamycin; UPR: unfolded protein response

Abstract

DNAJB12 (JB12) is an endoplasmic reticulum (ER)-associated Hsp40 family protein that recruits Hsp70 to the ER surface to coordinate the function of ER-associated and cytosolic chaperone systems in protein quality control. Hsp70 is stress inducible, but paradoxically, we report here that JB12 was degraded by the proteasome during severe ER stress. Destabilized JB12 was degraded by ER-associated degradation (ERAD) complexes that contained HERP, Sel1L, and gp78. JB12 was the only ER-associated chaperone that was destabilized by reductive stress. JB12 knockdown by siRNA led to the induction of Caspase processing, but not the unfolded protein response. ER stress-induced apoptosis is regulated by the highly labile and ER associated BCL-2 family member BOK, which is controlled at the level of protein stability by ERAD components. We found that JB12 was required in Huh-7 liver cancer cells to maintain BOK at low levels and BOK was detected in complexes with JB12 and gp78. Depletion of JB12 during reductive stress or by shRNA from Huh-7 cells was associated with accumulation of BOK, and activation of Caspase 3, 7, and 9. Absence of JB12 sensitized Huh-7 to death caused by proteotoxic agents and the proapoptotic chemotherapeutic LCL-161. In summary, JB12 is a stress sensitive Hsp40 whose degradation during severe ER stress provides a mechanism to promote BOK accumulation and induction of apoptosis.

Cell viability is dependent upon maintenance of the proteome in a pristine state with defects in protein homeostasis being associated with neurodegenerative diseases, cancers and diabetes (1). Approximately 30% of total protein synthesis occurs on ER-associated ribosomes and protein biogenesis in the ER is coupled with lipid biosynthesis, the secretory pathway, and energy metabolism (2,3). Defects in ER homeostasis are sensed in part by the unfolded protein response (UPR) signaling pathway which controls rates of protein synthesis and expression of ER associated molecular chaperones (4). ER-transmembrane Hsp40s containing a cytosolic J-domain direct cytosolic Hsp70 to function on the ER surface for action in facilitating the folding and/or degradation of polytopic membrane proteins (5-8).

Hsp40s that attract Hsp70 to the cytosolic face of the ER include HDJ-2, which is farnesylated (6), and ER transmembrane Hsp40s such as DNAJB12 (JB12) (7,9) and DNAJB14 (JB14) (10). These Hsp40s employ a conserved J-domain to attract Hsp70 to the cytosolic face of the ER and mediate the localized regulation of Hsp70's polypeptide binding and release cycle (11-13). HDJ-2 is implicated in the folding of proteins, whereas JB12 is multifunctional. JB12 is a component of ERQC complexes and triages ERAD sensitive and resistant forms of misfolded membrane proteins between pathways for proteasomal degradation or ERQC autophagy (8,14). JB12 also functions in a pathway with JB14 to facilitate the assembly of homotetrameric K⁺ channels. Notably JB12 and

JB14 appear to facilitate different steps required for conversion of K⁺ channel monomers to tetramers (15). Likewise, requirements for JB12 in ERQC-autophagy are not shared by JB14 (8). JB12 and JB14 help specify actions of Hsp70 in ER homeostasis, and contain divergent ER luminal DUF1977 (domain of unknown function) domains, could confer the observed specificity of their actions (10).

Interestingly, JB12 and JB14 expression is not induced by the UPR or heat stress (10), and the processes that are facilitated by them in ERQC are dictated in part by the conformation of the clients to which each respective Hsp40 binds (8,15). The study of JB12 has received more attention than JB14, and JB12 is the primary focus of this study. Depletion of JB12 from mammalian cells did not cause a strong induction of the UPR, but reduced cell growth rates and also sensitized cells to death caused by challenge of the ER with misfolded membrane proteins (8). JB12 therefore appears to participate in an ER stress signaling circuit that senses fluctuations in loads of misfolded proteins to regulate induction of ER stress-induced apoptosis via a circuit that may lie outside of the UPR network. However, how JB12 functions to protect cells from ER stress-induced cell death is not clear.

A basic understanding of how JB12 facilitates cross-talk between the UPR and heat-shock response systems to suppress ER stress-induced apoptosis requires information about how its activity is regulated. The study of this question was initiated through examination of the impact that acute ER stress caused by proteotoxic chemical stressors had on JB12, and for comparison JB14, expression and function. Remarkably, JB12, but not JB14, was found to be stress sensitive, as it was rapidly destabilized during challenge of the ER with the reducing agent dithiothreitol (DTT). Reductive stress caused by DTT appeared to alter the conformation of the DUF1977 domain on JB12 and cause its selection for degradation by the ERQC factors HERP, Sel1L, and the ERAD ubiquitin ligase gp78. Regions in the DUF1977 domain of JB12 and JB14 are not well conserved, and this may account for the differential sensitivity of them to DTT. JB12 is an ER associated Hsp40 that is selectively degraded in response to ER stress.

Huh-7 liver carcinoma cells experience chronic ER stress as indicated by detection of high level expression of the glycoprotein chaperone calnexin, the ERQC E3 ligase RMA1, and the ER associated BCL-2 protein BOK. BOK has an extremely short half-life, is constitutively degraded by ERAD, and its stress-dependent accumulation helps trigger the

induction of ER stress-induced apoptosis (16-18). JB12 is detected in ERQC complexes that control BOK levels and stress-dependent depletion of JB12 from Huh-7 caused the accumulation of BOK, induction of Caspase-9, -3 and -7 processing, and increased sensitivity to proteotoxic agents. JB12 is a multifunctional and ER stress sensitive Hsp40 that functions to suppress BOK activity during normal growth. During severe bouts of ER stress the inactivation of JB12 appears to contribute to the elevation of BOK and the induction of ER stress-mediated apoptosis. Mechanisms by which JB12 functions to facilitate protein homeostasis and suppress ER stress-induced apoptosis will be discussed.

Results

JB12 is Selectively Degraded in Response to Reductive ER Stress

To identify mechanisms for post-translational regulation of JB12 and JB14 in response to acute proteotoxic stress, COS-7 cells were challenged with chemicals that disrupt protein homeostasis in the ER membrane system, dithiothreitol (DTT), thapsagargin (Tg), or tunicamycin (Tm), or in the cytosol, bortezomib (Bort). Then, stress-dependent changes in levels of JB12, JB14 and different ERAD factors and cytosolic Hsp70 and Hsp40s were measured by western blot (Figure 1A). Under conditions where Bort inhibited the proteasome and increased the level of cytosolic Hsc70, there was no change in the level of JB12 or JB14 (Figure 1A). Likewise, disruption of glycoprotein folding with Tm, which blocks N-linked glycosylation, had no effect on JB12 or JB14 levels. Yet, Tm clearly stressed cells because it caused an increase in the levels of the ER luminal Hsp70 BiP, ER transmembrane calnexin, and the ERAD factors Derlin-1 and Derlin-3 (Figure 1A). In stark contrast, a dramatic and selective decrease in levels of JB12, but not JB14, occurred when cells were challenged with the reducing agent DTT (Figure 1A). Tg, an inhibitor of Serca2A that depletes the ER of Ca²⁺, caused a reduction in JB12 levels during the 6 hr challenge. Yet, the effect of Tg was modest when compared to the impact of DTT on steady-state levels of JB12.

ER stress caused by proteotoxic agents occurs in a time and dose dependent fashion, so the impact of Tg on the apparent stability of JB12 was further explored in a time course experiment in which the Tg concentration of 3 μ M used in panel 1A was increased to 6 μ M and changes in JB12 levels were

monitored over a longer 24 hr period of challenge (Figure 1B). The higher dose of Tg caused a detectable reduction in JB12 levels between 2 and 4 hrs of challenge, and near complete depletion occurred between 8 and 24 hrs. Time dependent reductions in JB12 caused by Tg correlated with increasing levels of ER stress as indicated by increased accumulation of BiP.

JB12 levels were reduced by DTT and Tg, but not Tm. JB12 is not predicted to contain a Ca^{2+} binding domain, but its ER luminal DUF1977 domain contains conserved cysteine residues (7), which could be sensitive to changes in oxidative/reductive state of the ER lumen. DTT might therefore have a direct effect on the conformation of JB12, whereas impacts of Tg, which occur over a longer time period, could be an indirect result of the interference with function of Ca^{2+} dependent chaperones such as calnexin that participate in oxidative protein folding in the ER lumen (3). Tm may not impact the stability of JB12 because it interferes with N-linked glycosylation and JB12 is not a glycoprotein. For these reasons, the challenge of cells with DTT was employed as a model for ER stress to study the mechanism for stress-dependent degradation of JB12.

If DTT was having a direct impact on the conformation of JB12, the destabilization of JB12 should occur before the ER becomes severely stressed (Figure 1C). Indeed, in cells challenged with the low dose of 0.6 mM DTT for 2 hrs the destabilization of JB12 occurs, while at the same time the phosphorylation of the translation initiation factor 2 alpha ($\text{eIF2}\alpha$) was not stimulated. The phosphorylation of $\text{eIF2}\alpha$ is mediated by the ER transmembrane kinase PERK in response to the ER stress-induced sequestration of BiP into complexes with misfolded proteins (3). Thus, DTT can rapidly destabilize JB12 in the absence of severe ER stress.

In cycloheximide chase studies, DTT reduced the half-life of JB12 from 6 hrs to around 1.5 hrs (Figure 1D). In these same cells, the half-life of the short-lived RMA1 (19), which is an ERQC ubiquitin ligase that interacts with JB12, was increased from 2 hrs to greater than 6 hrs, whereas the half-life of Derlin-1, an ERQC factor present in complexes with JB12, was not detectably altered. Destabilization of JB12 occurs soon after challenge of cells with low doses of DTT and happens while its normally short-lived interaction partner RMA1 is stabilized. JB12 was identified as the first Hsp40 family member to be selectively destabilized in response to acute ER stress.

The ER Luminal DUF1977 Domain Permits the Stress Sensitive Degradation of JB12

JB12 contains an N-terminal J-domain, a transmembrane span, and an ER luminal DUF1977 domain, and each could be sensitive to ER stress (Figure 2A). Yet, DTT prevents oxidative folding within the ER lumen, so the logical ER stress sensitive domain of JB12 is DUF1977. It is not clear what role the DUF1977 plays in JB12 function, but it is just over 110 amino acid residues long (Figure 2A), and is conserved in ER transmembrane Hsp40s that are expressed in organisms from yeast to man (7,20). JB12 and JB14 both contain a DUF1977 domain, but the stability of JB12 is sensitive to stress, whereas JB14 is resistant to challenges that destabilize JB12. The differential stress sensitivity of JB12 and JB14 is consistent with observations that these Hsp40s share some overlapping functions, but also have specific non-overlapping roles in ER homeostasis (15). Sequence alignment of the JB12 and JB14 DUF1977 showed a high degree of identity in amino-terminal regions, but there was a low degree of identity between their C-termini (Figure 2A). Such differences might explain why JB14 appears resistant to ER stress that destabilizes JB12.

To gain experimental support that the DUF1977 of JB12 is sensitive to ER stress, N- and C-terminal epitope tags were placed on JB12. Then the sensitivity to DTT of the different forms of JB12 was determined (Figure 2B). Overexpressed forms of wild-type and N-terminally FLAG tagged JB12 were destabilized by DTT in a manner similar to endogenous JB12. However, the presence of a MYC tag on the C-terminus abrogated the sensitivity of JB12 to DTT. The C-terminus of JB12 is highly charged, so it may be disordered. The amino acid sequence of a MYC tag (Glu-Gln-Leu-Ile-Ser-Glu-Glu-Asp-Leu) contains a number of polar and charged amino acids. Therefore, interactions between residues within DUF1977 and the MYC tag have the potential to promote conformational changes that confer resistance of JB12 to DTT.

JB12 contains two conserved cysteine residues that could be sensitive to DTT that are located at position 329 and 363 of DUF1977 (Figure 2A). The mutant C329A JB12 accumulates at a moderately lower level than JB12, but C329A JB12 remains sensitive to DTT. In contrast, accumulation of the mutant C363A JB12 was markedly reduced and it was not sensitive to DTT. Therefore, C363 appears to be important for folding of JB12 into stable conformation that is sensitive to DTT.

Why JB14, which contains the same two conserved cysteine residues as JB12, is not sensitive to DTT, is not entirely clear. Yet, C363 is located near the end of JB12, and the last 40 amino acids of the DUF1977 in JB12 and JB14 are highly divergent (Figure 2A). These differences in amino acid composition may permit JB14, but not JB12, to adopt a stress resistant state.

Interference with BiP Function in the ER Lumen Destabilizes JB12

The DUF1977 of JB12 might be metastable and therefore undergo a conformational change that leads to its degradation during ER stress. If so, then the action of ER luminal chaperones should be required for maintenance of JB12 during normal growth, and forms of JB12 that are destabilized by stress might become substrates of BiP. Indeed, in unstressed cells, a small pool of endogenous BiP was detected in native immunoprecipitates with FLAG-JB12 (Figure 3A). Upon challenge of cells with DTT, a 4-fold increase in the association of endogenous BiP with immunoprecipitated FLAG-JB12 occurred (Figure 3A). In addition, the depletion of BiP by siRNA caused a reduction in JB12 levels and the absence of BiP also increased the sensitivity of JB12 to DTT (Figure 3B). Thus, depletion of BiP partially destabilizes JB12 and forms of JB12 that are destabilized by DTT are bound by BiP.

The depletion of BiP by siRNA is nearly complete, and is similar to challenge with DTT in that it caused the strong induction of ER stress (Figure 3B). Levels of ER stress caused by the depletion of BiP were monitored by assaying decreases in the mobility of the kinase PERK on SDS-PAGE, which occur when PERK becomes activated through autophosphorylation (3). PERK appeared to be fully activate when BiP was depleted because BiP was no longer present to bind the ER luminal regions of PERK and maintain it in an inactive state. Therefore, a concern about the evaluation of data from BiP siRNA knockdown studies is that the near complete loss of BiP from cells causes a stress that is severe enough to non-specifically destabilize JB12. Hence, it was asked if JB12 could also be destabilized under conditions where BiP was only partially depleted, and the ER experienced low to moderate stress (Figure 3C).

The partial reduction of BiP in cells was achieved through the ectopic expression the cytotoxic endoprotease Subtilase A, which accumulates in the ER lumen and cleaves BiP (21). Cells were transfected with a Subtilase A expression

plasmid. Then at 24, 48, and 72 hrs post-transfection the levels of JB12, BiP, HERP, CHOP, and cleaved caspase 3 and 7 were measured by western blot. After 24 hrs, JB12 levels were relatively unchanged and levels of BiP were about 50% below normal. Yet, at the 48 hr time point, JB12 levels were 60% lower and BiP was reduced by 70%. Notably, depletion of JB12 and BiP detected 48 hrs after transfection of the cells with the Subtilase A expression plasmid, was not associated with a strong induction ER stress. This is the case because pools of active and lower mobility forms of PERK were not strongly increased. In addition, induction of CHOP, an ER stress responsive transcription factor that controls ER chaperone levels (3), and processing of Caspase 3 and 7 in Subtilase A expressing cells were well below the levels observed in DTT treated cells. In addition, HERP expression is induced by the severe ER stress caused by DTT, but HERP levels were not increased when BiP was partially depleted by Subtilase A (Figure 3C).

Partial depletion of BiP, under conditions where PERK is not strongly activated, leads to destabilization of JB12. JB12 has features of a metastable protein that requires BiP to maintain its steady-state levels in the ER. In addition, destabilized JB12 appears to misfold and be bound by BiP prior to its delivery to ERAD for degradation.

Degradation of Destabilized JB12 Requires the ERAD Factors HERP, Sel1L and gp78

In unstressed cells, JB12 functions in complexes with the ERQC E3 ligases RMA1 and gp78, where it acts with Hsp70 to triage non-native membrane proteins between folding and degradation pathways (7). Therefore, we asked if destabilized JB12 becomes a substrate of the ERAD machinery with which its stable forms normally interact. Depletion of RMA1, CHIP or HRD1 by siRNA did not have a remarkable impact on the DTT dependent degradation of JB12 (Figure 4A). In contrast, the depletion of gp78, Sel1L, and HERP suppressed the degradation of JB12 caused by DTT.

HERP, and Sel1L act in the ER in heterooligomeric E3 Ubiquitin ligase complexes that contain either HRD1 or gp78 that degrade both soluble and membrane inserted conformers of misfolded proteins (22,23). HERP functions as an organizing factor (24,25), whereas Sel1L acts in the selection of misfolded proteins for ubiquitination (23). JB12 was detected in immunoprecipitable complexes with both Sel1L and HERP (Figure 4B). In pull-downs where HERP-FLAG was the bait, Sel1L and JB12 were both detected. In pull-downs

where MYC-Sel1L was the bait, JB12 was also detected in immunoprecipitates (Figure 4C). Notably, the siRNA mediated depletion of endogenous HERP caused an approximately 60% reduction in the association of JB12 with Sel1L (Figure 4C). This result may explain why the knockdown of HERP hindered the degradation of JB12 caused by DTT. Treatment of cells with DTT reduced JB12 association with HERP and Sel1L, so damaged forms of JB12 appeared to be rapidly cleared from ERAD machinery.

Sel1L/gp78 has been reported to form an E3 complex that has clients that are distinct from the HRD1/Sel1L complex (26), and this helps explain why the degradation of JB12 caused by DTT is less sensitive to depletion of HRD1 than gp78. These data suggest that the stress damaged forms of JB12 are selected for ERAD by the ERQC machinery with which native JB12 normally interacts during maintenance of ER homeostasis.

JB12 Limits BOK Accumulation and Suppresses ER Stress-induced Apoptosis in Huh-7 Cells

Cancer cells experience chronic ER stress due in part to constant demands for oxidative protein folding in the ER lumen and in some cancers Hsp70 molecular chaperones are essential for malignancy (27,28). Therefore, we asked if the human hepatoma cell line 7 (Huh-7) (29) experiences chronic ER stress. The role played by JB12 in supporting the cancerous growth of Huh-7 was also examined. Huh-7 experiences a form of chronic ER stress because levels of RMA1 and calnexin were several fold higher in Huh-7 versus COS-7 (Figure 5A). HEK293 cells are multinucleate and express high levels of molecular chaperones, and levels of BiP, Derlin-1 and JB12 were similar in Huh-7, COS-7 and HEK293 (Figure 5A).

The acute challenge of Huh-7 with DTT caused JB12 levels to plummet, BiP levels to rise, and induced cleavage of proCaspase-9, -3 and -7 (Figure 5B). JB12 was also found to be important for viability of unstressed Huh-7 cells because its depletion via shRNA was accompanied by an apparent induction of apoptosis that was indicated by the dramatic increase in processed Caspase-9, -3 and -7 (Figure 5B). Notably, depletion of JB12 by shRNA resulted in a much larger increase in Caspase processing than treatment of Huh-7 with DTT.

In the absence of JB12, acute treatment of Huh-7 with DTT, further stimulated accumulation of cCasp-9, -3 and -7 (Figure 5B). Whereas, challenge of JB12 depleted Huh-7 with Tg or Bort, for the same time period, only increased the accumulation

of cCasp-7. JB12 plays an important role in growth of Huh-7, and loss of JB12 leads to the induction of apoptosis.

ER stress-induced apoptosis is regulated in part by mechanisms that control the steady-state level of the ER associated BCL-2 protein BOK. BOK induces Caspase processing by stimulating the release of cytochrome C from mitochondria (16). Remarkably, BOK is expressed, but not readily detected in unstressed cells, because it is constitutively degraded and has a short half-life of 15 minutes (16). However, during proteotoxic stress that compromises function of the proteasome, ERQC E3 ligases such as gp78, which mediate BOK degradation, become saturated with misfolded proteins, and BOK accumulates (16).

To define the mechanism by which JB12 suppresses the induction of apoptosis, the impact that its presence or absence in Huh-7 had on BOK levels was examined (Figure 6A). Consistent with Huh-7 experiencing chronic ER stress, endogenous BOK was readily detected in western blots of cell extracts (Figure 6A). Treatment of Huh-7 with DTT was accompanied by a greater than 2.5-fold increase in BOK and 60% decrease in JB12 levels. The shRNA knockdown of JB12 to levels below detection by western blot also resulted in a 3.5-fold increase in BOK (Figure 6A). BOK levels were increased up to 5-fold above controls upon challenge of JB12 depleted Huh-7 with DTT. Thus, endogenous BOK is readily detected in Huh-7, and its levels increase several fold upon depletion of JB12. The dramatic increase in BOK levels that accompanies depletion of JB12 from Huh-7 helps explain why Caspase processing was activated in Huh-7 cells that lacked JB12 (Figure 5B).

Next, we evaluated whether or not JB12 played a direct or indirect role in influencing the steady-state levels of BOK. To accomplish this goal, the impact on BOK of elevating the activity of WT JB12 or inhibiting JB12 function through overexpression of a dominant negative form of JB12 (QPD JB12) was measured (Figure 6B). For these studies FLAG-BOK was expressed at low levels in HEK293. Since JB12 is an abundant protein, its overexpression did not reduce BOK accumulation (Figure 6B). In contrast, QPD JB12, a form of JB12 that can bind ERQC factors, but fails to interact with Hsp70 (7), caused a dose-dependent and several fold increase in FLAG-BOK (Figure 6B). Interference with the function of endogenous JB12 appears to block the BOK turnover.

If JB12 directly influences BOK accumulation, QPD JB12 would be expected to block BOK turnover at an intermediate stage where BOK was

associated with components of the ERAD system. Indeed, QPD JB12 was detected at 4-fold higher levels than WT JB12 in immunoprecipitates with FLAG-BOK (Figure 6C). Likewise, when cells were treated with Bort to inhibit proteasomal degradation of BOK, both gp78 and QPD JB12 were found to accumulate in complexes with FLAG-BOK (Figure 6D).

JB12 is physically present in complexes that contain BOK and gp78. Failure of JB12 to interact with Hsp70 causes BOK levels to rise because BOK appears to become trapped in complexes with gp78. These findings suggest that JB12 and Hsp70 mediate protein assembly/disassembly events that enable gp78 to facilitate steps in BOK degradation. It is plausible that inactivation of JB12 during ER stress provides a mechanism to promote the accumulation of BOK and trigger downstream induction of Caspase processing.

JB12 Confers Resistance of Huh-7 to Chemotherapeutics that Stimulate Apoptosis

To further test the concept that JB12 protects cells from ER stress-induced death, the requirement for JB12 in the survival of Huh-7 to challenge with proteotoxic agents was examined (Figure 7A). Huh-7 tolerated treatment with 2 mM DTT for 36 hrs. Yet, 50% of JB12 depleted Huh-7 lost viability within 24 hrs of exposure to DTT (Figure 7A). Huh-7 tolerated treatment with Bort for 12 hrs, but 50% lost viability after exposure for 24 hrs. Whereas, 50% of JB12 depleted Huh-7 lost viability within 12 hrs of Bort challenge. JB12 protects Huh-7 from toxic stress caused by perturbation of reductive protein folding and the global interference with proteasome function.

To further assess the function of JB12 in protecting cells from stress, we asked if reduction of its activity sensitized cells to the apoptosis inducer LCL-161. LCL-161 is a small molecule second mitochondrial activator of Caspase (SMAC) mimetic (30). LCL-161 induces apoptosis by promoting the degradation of inhibitor of apoptosis proteins (IAPs) that bind procaspases and inhibit their processing/activation (30). However, the action of LCL-161 alone is insufficient to kill cancer cells and a second stimulator of apoptosis is required for it to induce cancer cell death (31). Indeed, we found that treatment of Huh-7 with LCL-161 caused the depletion of cIAP-1 (Figure 2B), but did not reduce the viability of Huh-7 (Figure 7A). Yet, the depletion of JB12 from Huh-7 sensitized them to LCL-161-induced death (Figure 7A).

Consistent with the shRNA depletion of JB12 causing the accumulation of proapoptotic BOK (Figure 6A), reduction of JB12 permitted LCL-161 to drive a dose dependent increase in Caspase activation (Figure 7B). These findings indicate that JB12 is required to protect Huh-7 from proteotoxic stress via a mechanism that involves the suppression of BOK accumulation. The loss of JB12 function in Huh-7 contributes to the accumulation of BOK, activation of proCaspase processing, and induction of ER stress-induced apoptosis.

Discussion

Hsp70 and Hsp40s are constitutively expressed and a subset of them are induced to protect cells from proteotoxicity (1,32). JB12 recruits Hsp70 to the ER surface to facilitate protein folding and protein triage and has unique features amongst other members of the Hsp40 family because it is destabilized by ER stress. Destabilized JB12 is degraded via an ERAD pathway that utilizes HERP, the E3 ligase gp78, and the ERAD substrate selector Sel1L. Alteration of C363 in the ER luminal DUF1977 domain prevents JB12 from adopting a stress sensitive conformation. Proper folding/assembly of JB12 appears to be required for it to adopt a stress sensitive conformation.

JB12 was found to function in association with gp78 to mediate the constitutive degradation of BOK, a stress-sensitive and short lived BCL-2 family member that triggers ER stress-induced apoptosis (Figure 8). Stress-dependent degradation of JB12 is associated with increased BOK, induction of Caspase processing, and sensitization of Huh-7 liver cancer cells to proteotoxic agents and a chemotherapeutic. JB12 has features of an ER stress sensor whose inactivation during acute stress permits the accumulation of BOK, and thereby triggers the initiation of ER stress-induced apoptosis.

JB12 and JB14 perform overlapping functions in ER homeostasis, but are not functionally redundant, as each respective Hsp40 can facilitate a unique aspect of ER protein homeostasis that is not facilitated by the other (8,10,15). The observation that JB12, but not JB14, stability is sensitive to acute stress is therefore consistent with JB12 and JB14 having unique roles in maintenance of protein homeostasis. The mechanism for the differential sensitivity of JB12 and JB14 to stress is not clear, but the DUF1977 of both JB12 and JB14 is highly charged, though not identical, and monomeric forms are predicted to be dynamically unfolded (7). Therefore, differences in the dynamic

conformational states of JB12 and JB14 may permit JB12 to populate a stress-sensitive state that is not populated by JB14. The biogenesis of JB12 is sensitive to mutation of C363 in the DUF1977 of JB12 because C363A JB12 accumulated at low levels and was insensitive to DTT. JB12 also contains C329, but C329A JB12 remains sensitive to DTT, so destabilization of JB12 by DTT does not appear to result from disruption of intrachain disulfide bonds. However, Hsp40s function as homo- and heterodimers (33,34), so assembly of JB12 into a stable oligomeric complex may require disulfide bond formation that involves C363. The disruption of JB12's conformation or assembly with other proteins during ER stress could provide a mechanism for its destabilization.

JB12 facilitates aspects of ERAD that involve the selection of misfolded membrane proteins for ubiquitination by E3 ligase complexes that contain RMA1/RNF5, gp78, Hrd1 and possibly others (7,35-37). When JB12 associates with misfolded ER proteins that are inefficiently retrotranslocated out of the ER, it can also mediate the transfer of its ERAD-resistant clients to the ERQC autophagy pathway (8). JB12 therefore functions as a protein triage factor, and loss of JB12 should cause serious negative side effects on cell biology. Indeed, COS-7 and HEK293 cells that lack JB12 die from apoptosis when challenged with large pools of misfolded membrane proteins (8). Consistently, Huh-7 liver cancer cells that lack JB12 were reported herein to be hypersensitive to challenge with a proteasome inhibitor, DTT, and the apoptosis inducer LCL-161.

Strikingly, reduction of JB12 in Huh-7 is associated a modest elevation of BiP, a strong increase in the accumulation of BOK and robust activation of Caspase processing. These data suggest that the inability of JB12 to function in protein triage triggers early steps in ER stress-induced apoptosis. Such a scenario may occur during proteotoxic stress encountered by cells, and may also occur in disease states where mutant forms of membrane proteins misfold and aggregate in the ER bilayer (5-8). Aggregated membrane proteins are resistant to degradation by ERAD or ERQC-autophagy (8), so they have the potential to sequester JB12 and inhibit its function in ER homeostasis. Sequestration of JB12 would decrease its availability to facilitate degradation of BOK and set the stage for induction of ER stress-induced apoptosis.

Cancer cells suffer from constitutive ER stress due to dysregulation of normal growth control and heavy flux through pathways for oxidative protein folding in the ER lumen (3,38). Recent studies have demonstrated the importance of the Hsp70 and

Hsp90 systems in promoting the cancerous growth of cells in tumors (39). Data presented suggest that one function of Hsp70 in fostering cancerous cell growth is limiting the accumulation of the cell death inducer BOK. Hence, the inhibitors of Hsp70 are under development as drugs with chemotherapeutic potential (40). However, the Hsp70 system is essential for cell viability, so Hsp70 inhibitors are likely to have broad side effects that limit their utility as drugs. Yet, Hsp40s such as JB12 are expressed at substoichiometric levels to Hsp70 and direct specific Hsp70 family members to facilitate distinct subsets of reactions required for protein homeostasis. Therefore, targeting a specific Hsp40, such as JB12, for inactivation to promote BOK accumulation has the potential to reduce the side effects of current drugs designed to modulate Hsp70 that globally antagonize Hsp70 function (41). Since loss of JB12 primes Huh-7 cancer cells for death by causing BOK accumulation for killing by LCL-161, it is possible that a compound that selectively drives the degradation of JB12 could be used in the clinic to induce apoptotic death of cancer cells.

Experimental Procedures

Cell Culture and Transfection

African green monkey kidney (COS-7), human hepatocellular carcinoma (Huh-7), and human embryonic kidney (HEK293) cells were cultured in Dulbecco's eagle minimum enriched medium (DMEM) at 37°C, 5% CO₂. Cells were seeded in 6-well plates at a density of 3x10⁵ cells/well. Plasmid transfections into COS-7 cells were performed by using Effectene (QIAGEN, Hilden, Germany) at 20-hr post-seeding. Expression plasmids included pcDNA3.1-DNAJB12, pcDNA3.1-DNAJB12-FLAG, pcDNA3.1-Sel1L-myc, pCMV-HERP-myc-DDK (Cat.RC217811, Origene), and pcDNA3.1-SubtilaseA (Plasmid 35948, Addgene). (see Supplemental Materials and Methods for additional plasmid information). Cell were cultured for 48 hrs before being challenged with stress-inducing chemicals (see Supplemental Table 1-3 for list of chemicals and tools utilized in this study). Supplemental Table 4 contains the molecular weights of the proteins that were detected by western blot.

For knockdown experiments, all siRNAs were purchased from Thermo Fischer Scientific. For some target genes, two siRNAs to the same target gene were used to increase knockdown efficiency. COS-7 cells that were seeded into 6-well plate at 3x10⁵ cells/well for 20 hrs were transfected at 5 pmole per well of siRNA with Lipofectamine2000® (Thermo

Fischer Scientific, Waltham, MA, USA). Twenty hrs post-transfection, cells were split at 1:2 ratios and allowed to recover for 48 hrs. JB12 knockdown in Huh-7 cells was achieved by using Lentiviral shJB12 expressed from pLKO.1 (TRCN0000022297, Open Biosystems, Huntsville, AL, USA). Huh-7 cells were grown for 48 hrs and then incubated with 10 μ L of raw lentiviral stock per ml of serum-free DMEM for 24 hrs. The infected Huh-7 cells were then grown an additional 24 hrs in complete media and then split 1:2 into a new plate and grown for 48 hrs prior to analysis.

Lysate Preparation for Western blotting

COS-7 cells or Huh-7 cells were collected and then solubilized in 100 μ L NP-40 lysis solution (1% (v/v) NP-40, 50 mM Tris-HCl (pH 7.5), 150 mM NaCl and 0.05% (w/v) SDS) supplemented with 1 mM PMSF, 1x complete protease inhibitor cocktail (Roche, Basel, Switzerland), 1x PHOSstop phosphatase inhibitor cocktail (Roche). The lysates were then homogenized by digital sonicator at 26% intensity for 7 sec. The homogenized lysates were diluted to 1 μ g/ μ L total protein in 1x SDS-PAGE sample buffer containing 20mM DTT. The mixed protein samples were incubated at 37°C for 30 min. before resolving by SDS-PAGE. The material in resolved SDS-PAGE gels was transferred onto 0.22 μ M nitrocellulose membranes (Bio-Rad, Hercules, USA). Membranes were analyzed by western detection with indicated antibodies (see Supplemental Table 3 for antibodies used in this study and Supplemental Table 4 for the molecular weight of the respective proteins).

Protein signal detection was performed by using Clarity™ western ECL substrate (Bio-Rad, Hercules, USA) and visualized by LAS4000 imager (GE Life Sciences, Pittsburgh, USA). Image data was analyzed by ImageJ (NIH, USA) and quantitated with Imagequant (GE Life Sciences, Pittsburgh, USA).

Analysis of PERK phosphorylation was enhanced by used of the Phos-Tag™ reagent (Wako, Osaka, Japan). This required cell lysate preparation in NP-40 lysis buffer that was supplemented with EDTA-free 1x complete protease inhibitor cocktail (Roche). Electrophoresis was performed with use of EDTA-free 6% SDS-PAGE that contained 50 μ M MnCl₂ and 3.5 μ M Phos-Tag™ (Wako, Osaka, Japan) and proteins were resolved at a constant current of 10 mA.

Immunoprecipitation of Proteins from Cell Extracts Prepared with Non-Ionic Detergents

Transfected HEK293 or COS-7 cells were harvested from plates with ice-cold Citric Saline (135mM KCl, 15 mM Sodium Citrate), and then lysed in NP-40 buffer that was supplemented with 1 mM PMSF. Insoluble material was removed by centrifugation at 14,000 rpm (use x g instead of rpm?), 4°C for 5 min. Antibodies against bait proteins were added to the cleared lysates and incubated for 30 min. at 4°C. The target protein-antibody complex was isolated with Protein G-Agarose (Roche) during an incubation at 4°C for 30 min. Material bound to agarose beads was washed three times with NP-40 lysis buffer was then recovered from beads via incubation with 2x SDS-PAGE sample buffer at 37°C for 30 min. To pull down FLAG-containing proteins, anti-FLAG M2 beads (Sigma-Aldrich, St. Louis, USA) were used instead of Protein G-Agarose. Material that was bound to FLAG beads was then eluted with FLAG peptide and the liberated materials were probed by western blot.

In studies of the effect of HERP on interaction between DNAJB12 and Sel1L, COS-7 cells grown for 24 hrs in a 6-well plate were transfected with siHERP. After 24 hrs, cells were split 1:2, allowed to grow for another 20 hrs, then transfected with pcDNA3.1-Sel1L-myc and pcDNA3.1-DNAJB12. After a period of 24 hrs cells were harvested and immunoprecipitations and Western blot analysis was conducted.

Assay for Changes in Cell Viability

COS-7 or Huh-7 cells were seeded at 1x10⁵ cells/well in a 96-well plate and grown for 48 hrs. Media was then replaced with a new one containing ER stressors or anti-cancer drug LCL-16. Cell viability was indirectly measured by changes in cellular ATP levels using the CellTiter-Glo Luminescent Cell Viability Assay (Promega, Madison, WI, USA), following manufacturer's protocol.

Acknowledgements

This work is supported by NIH grant 2RO1 GM56981. Thanks to Dr. Lihua He and Ali Cyr-Scully for assistance with editing the manuscript.

Conflicts of Interest

There are not conflicts of interest.

Author Contributions

SP, DG, HYR designed and carried out experiments, and also prepared the figures and text of the manuscript. DMC designed the study and assisted with manuscript preparation.

Literature Cited

1. Balchin, D., Hayer-Hartl, M., and Hartl, F. U. (2016) In vivo aspects of protein folding and quality control. *Science* **353**, aac4354
2. Yamamoto, K., Sato, T., Matsui, T., Sato, M., Okada, T., Yoshida, H., Harada, A., and Mori, K. (2007) Transcriptional induction of mammalian ER quality control proteins is mediated by single or combined action of ATF6alpha and XBP1. *Dev Cell* **13**, 365-376
3. Walter, P., and Ron, D. (2011) The unfolded protein response: from stress pathway to homeostatic regulation. *Science* **334**, 1081-1086
4. Hetz, C., Chevet, E., and Oakes, S. A. (2015) Proteostasis control by the unfolded protein response. *Nat Cell Biol* **17**, 829-838
5. Meacham, G. C., Patterson, C., Zhang, W., Younger, J. M., and Cyr, D. M. (2001) The Hsc70 co-chaperone CHIP targets immature CFTR for proteasomal degradation. *Nat Cell Biol* **3**, 100-105
6. Meacham, G. C., Lu, Z., King, S., Sorscher, E., Tousson, A., and Cyr, D. M. (1999) The Hdj-2/Hsc70 chaperone pair facilitates early steps in CFTR biogenesis. *Embo J* **18**, 1492-1505
7. Grove, D. E., Fan, C. Y., Ren, H. Y., and Cyr, D. M. (2011) The endoplasmic reticulum-associated Hsp40 DNAJB12 and Hsc70 cooperate to facilitate RMA1 E3-dependent degradation of nascent CFTRDeltaF508. *Mol Biol Cell* **22**, 301-314
8. Houck, S. A., Ren, H. Y., Madden, V. J., Bonner, J. N., Conlin, M. P., Janovick, J. A., Conn, P. M., and Cyr, D. M. (2014) Quality control autophagy degrades soluble ERAD-resistant conformers of the misfolded membrane protein GnRHR. *Mol Cell* **54**, 166-179
9. Yamamoto, Y. H., Kimura, T., Momohara, S., Takeuchi, M., Tani, T., Kimata, Y., Kadokura, H., and Kohno, K. A novel ER J-protein DNAJB12 accelerates ER-associated degradation of membrane proteins including CFTR. *Cell Struct Funct* **35**, 107-116
10. Sopha, P., Kadokura, H., Yamamoto, Y. H., Takeuchi, M., Saito, M., Tsuru, A., and Kohno, K. (2012) A novel mammalian ER-located J-protein, DNAJB14, can accelerate ERAD of misfolded membrane proteins. *Cell Struct Funct* **37**, 177-187
11. Liberek, K., Marszalek, J., Ang, D., Georgopoulos, C., and Zylicz, M. (1991) Escherichia coli DnaJ and GrpE heat shock proteins jointly stimulate ATPase activity of DnaK. *Proc Natl Acad Sci U S A* **88**, 2874-2878
12. Cyr, D. M., Lu, X., and Douglas, M. G. (1992) Regulation of Hsp70 function by a eukaryotic DnaJ homolog. *J Biol Chem* **267**, 20927-20931
13. Langer, T., Lu, C., Echols, H., Flanagan, J., Hayer, M. K., and Hartl, F. U. (1992) Successive action of DnaK, DnaJ and GroEL along the pathway of chaperone-mediated protein folding. *Nature* **356**, 683-689
14. Houck, S. A., and Cyr, D. M. (2012) Mechanisms for quality control of misfolded transmembrane proteins. *Biochimica Et Biophysica Acta-Biomembranes* **1818**, 1108-1114
15. Li, K., Jiang, Q., Bai, X., Yang, Y. F., Ruan, M. Y., and Cai, S. Q. (2017) Tetrameric Assembly of K+ Channels Requires ER-Located Chaperone Proteins. *Mol Cell* **65**, 52-65
16. Llambi, F., Wang, Y. M., Victor, B., Yang, M., Schneider, D. M., Gingras, S., Parsons, M. J., Zheng, J. H., Brown, S. A., Pelletier, S., Moldoveanu, T., Chen, T., and Green, D. R. (2016) BOK Is a Non-canonical BCL-2 Family Effector of Apoptosis Regulated by ER-Associated Degradation. *Cell* **165**, 421-433
17. Chipuk, J. E., and Luna-Vargas, M. P. (2016) Cell Biology: ERADicating Survival with BOK. *Curr Biol* **26**, R473-476
18. Fernandez-Marrero, Y., Ke, F., Echeverry, N., Bouillet, P., Bachmann, D., Strasser, A., and Kaufmann, T. (2016) Is BOK required for apoptosis induced by endoplasmic reticulum stress? *Proc Natl Acad Sci U S A* **113**, E492-493
19. Younger, J. M., Chen, L., Ren, H. Y., Rosser, M. F., Turnbull, E. L., Fan, C. Y., Patterson, C., and Cyr, D. M. (2006) Sequential quality-control checkpoints triage misfolded cystic fibrosis transmembrane conductance regulator. *Cell* **126**, 571-582
20. Kampinga, H. H., and Craig, E. A. (2010) The HSP70 chaperone machinery: J proteins as drivers of functional specificity. *Nat Rev Mol Cell Biol* **11**, 579-592
21. Nakajima, S., Hiramatsu, N., Hayakawa, K., Saito, Y., Kato, H., Huang, T., Yao, J., Paton, A. W., Paton, J. C., and Kitamura, M. (2011) Selective abrogation of BiP/GRP78 blunts activation of NF-kappaB

- through the ATF6 branch of the UPR: involvement of C/EBP β and mTOR-dependent dephosphorylation of Akt. *Mol Cell Biol* **31**, 1710-1718
22. Christianson, J. C., Shaler, T. A., Tyler, R. E., and Kopito, R. R. (2008) OS-9 and GRP94 deliver mutant α 1-antitrypsin to the Hrd1-SEL1L ubiquitin ligase complex for ERAD. *Nat Cell Biol* **10**, 272-282
 23. Kanehara, K., Xie, W., and Ng, D. T. (2010) Modularity of the Hrd1 ERAD complex underlies its diverse client range. *J Cell Biol* **188**, 707-716
 24. Horn, S. C., Hanna, J., Hirsch, C., Volkwein, C., Schutz, A., Heinemann, U., Sommer, T., and Jarosch, E. (2009) Usa1 functions as a scaffold of the HRD-ubiquitin ligase. *Mol Cell* **36**, 782-793
 25. Schulze, A., Standera, S., Buerger, E., Kikkert, M., van Voorden, S., Wiertz, E., Koning, F., Kloetzel, P. M., and Seeger, M. (2005) The ubiquitin-domain protein HERP forms a complex with components of the endoplasmic reticulum associated degradation pathway. *J Mol Biol* **354**, 1021-1027
 26. Zhang, T., Xu, Y., Liu, Y., and Ye, Y. (2015) gp78 functions downstream of Hrd1 to promote degradation of misfolded proteins of the endoplasmic reticulum. *Mol Biol Cell* **26**, 4438-4450
 27. Bhat, T. A., Chaudhary, A. K., Kumar, S., O'Malley, J., Inigo, J. R., Kumar, R., Yadav, N., and Chandra, D. (2017) Endoplasmic reticulum-mediated unfolded protein response and mitochondrial apoptosis in cancer. *Biochim Biophys Acta* **1867**, 58-66
 28. Hazari, Y. M., Bashir, A., Haq, E. U., and Fazili, K. M. (2016) Emerging tale of UPR and cancer: an essentiality for malignancy. *Tumour Biol* **37**, 14381-14390
 29. Kausar, H., Gull, S., Ijaz, B., Ahmad, W., Sarwar, M. T., Iqbal, Z., Nawaz, Z., Riazuddin, S., and Hassan, S. (2011) Huh-7 cell line as an alternative cultural model for the production of human like erythropoietin (EPO). *J Transl Med* **9**, 186
 30. Qin, Q., Zuo, Y., Yang, X., Lu, J., Zhan, L., Xu, L., Zhang, C., Zhu, H., Liu, J., Liu, Z., Tao, G., Dai, S., Zhang, X., Ma, J., Cai, J., and Sun, X. (2014) Smac mimetic compound LCL161 sensitizes esophageal carcinoma cells to radiotherapy by inhibiting the expression of inhibitor of apoptosis protein. *Tumour Biol* **35**, 2565-2574
 31. West, A. C., Martin, B. P., Andrews, D. A., Hogg, S. J., Banerjee, A., Grigoriadis, G., Johnstone, R. W., and Shortt, J. (2016) The SMAC mimetic, LCL-161, reduces survival in aggressive MYC-driven lymphoma while promoting susceptibility to endotoxin shock. *Oncogenesis* **5**, e216
 32. Cyr, D. M., Langer, T., and Douglas, M. G. (1994) DnaJ-like proteins: molecular chaperones and specific regulators of Hsp70. *Trends Biochem Sci* **19**, 176-181
 33. Sha, B., Lee, S., and Cyr, D. M. (2000) The crystal structure of the peptide-binding fragment from the yeast Hsp40 protein Sis1. *Structure Fold Des* **8**, 799-807
 34. Nillegoda, N. B., and Bukau, B. (2015) Metazoan Hsp70-based protein disaggregases: emergence and mechanisms. *Front Mol Biosci* **2**, 57
 35. Han, S., Liu, Y., and Chang, A. (2007) Cytoplasmic Hsp70 promotes ubiquitination for endoplasmic reticulum-associated degradation of a misfolded mutant of the yeast plasma membrane ATPase, PMA1. *J Biol Chem* **282**, 26140-26149
 36. Nakatsukasa, K., Hoyer, G., Michaelis, S., and Brodsky, J. L. (2008) Dissecting the ER-associated degradation of a misfolded polytopic membrane protein. *Cell* **132**, 101-112
 37. Morito, D., Hirao, K., Oda, Y., Hosokawa, N., Tokunaga, F., Cyr, D. M., Tanaka, K., Iwai, K., and Nagata, K. (2008) Gp78 cooperates with RMA1 in endoplasmic reticulum-associated degradation of CFTR Δ F508. *Mol Biol Cell* **19**, 1328-1336
 38. Lee, A. S., and Hendershot, L. M. (2006) ER stress and cancer. *Cancer Biol Ther* **5**, 721-722
 39. Sabnis, A. J., Guerriero, C. J., Olivas, V., Sayana, A., Shue, J., Flanagan, J., Asthana, S., Paton, A. W., Paton, J. C., Gestwicki, J. E., Walter, P., Weissman, J. S., Wipf, P., Brodsky, J. L., and Bivona, T. G. (2016) Combined chemical-genetic approach identifies cytosolic HSP70 dependence in rhabdomyosarcoma. *Proc Natl Acad Sci U S A* **113**, 9015-9020
 40. Assimon, V. A., Gillies, A. T., Rauch, J. N., and Gestwicki, J. E. (2013) Hsp70 protein complexes as drug targets. *Curr Pharm Des* **19**, 404-417
 41. Miyata, Y., Li, X., Lee, H. F., Jinwal, U. K., Srinivasan, S. R., Seguin, S. P., Young, Z. T., Brodsky, J. L., Dickey, C. A., Sun, D., and Gestwicki, J. E. (2013) Synthesis and initial evaluation of YM-08, a blood-brain barrier permeable derivative of the heat shock protein 70 (Hsp70) inhibitor MKT-077, which reduces tau levels. *ACS Chem Neurosci* **4**, 930-939

FIGURE LEGENDS

Figure 1. JB12 is a protein quality control factor that is selectively depleted from cells in response to ER stress

(A) Western blot analysis of the impact that proteotoxic agents have on levels of cytosolic and ER associated protein quality control factors. Extracts of COS-7 were analyzed by western blot after a 6-hr incubation with 2mM DTT, 3 μ M Tg, 5 μ g/mL Tm or 20 μ M Bort. (B) Reduction of JB12 levels by 6 μ M Tg during a 24 hr time course. (C) Dose-dependent impact of DTT on JB12 levels during a 2-hr incubation. (D) Cycloheximide (CHX; 150 μ g/ml) chase analysis of the impact that challenge with 2 mM DTT has on levels of JB12, Derlin-1 and RMA1.

Figure 2 Reduced Sensitivity of JB12 to ER Stress Upon Alteration of its ER luminal DUF1977 domain

(A) Schematic of the JB12 domain structure and sequence alignment of the JB12 and JB14 DUF1977. The DUF of JB12 is located between amino acid 263 and 375 and in JB14 lies between residues 266 and 379.

(B) Changes in sensitivity of JB12 to DTT (2 mM) during a 3 hr incubation caused by placing an epitope Tag on the JB12 N- or C-terminus or mutation of conserved cysteine residues in DUF1977.

Figure 3. Depletion of BiP from the ER lumen Leads to Depletion of JB12

(A) DTT stimulates the association of JB12 with BiP. COS-7 cells expressing JB12-FLAG were untreated or treated with DTT or Tg for 1 hr. The BiP that co-precipitated with JB12-FLAG was then detected by western blot. (B) BiP depletion induces ER stress and loss of JB12. COS-7 cells were transfected with siBiP for 48 hrs and where indicated were treated with DTT for 6 hrs. (C) Subtilase A-mediated BiP depletion stimulates Caspase processing and decreases JB12. Cells that were transfected with a Subtilase-A expression plasmid were grown for the indicated time period prior to preparation of extracts for use in western blots.

Figure 4. Stress Stimulated Turnover of JB12 Requires the ERQC Factor gp78, Sel1 and HERP

(A) Impact that siRNA knockdown (KD) of ERAD factors on ER stress-dependent degradation of JB12. COS-7 were treated with siRNAs for the indicated proteins as described in the materials and methods and then treated with DTT (2 mM) and CHX (150 μ g/ml) for 2 hrs. Changes in levels of the indicated proteins was then determined by western blot (B) JB12 is present in immunoprecipitable complexes that contain FLAG-HERP. COS-7 cells expressed exogenous and untagged JB12, HERP-FLAG and Sel1L-myc. (C) SiRNA depletion of HERP reduced the association of JB12 with Sel1L-myc. Identification of proteins that co-immunoprecipitated with tagged bait proteins was achieved by western blot.

Figure 5. Depletion of JB12 sensitizes Huh-7 cancer cells to proteotoxic stress and apoptosis

(A) Steady-state levels of ERQC factors and chaperones in Huh-7 as compared to COS-7 and HEK293. (B) Depletion of JB12 via shRNA is associated with the accumulation of cleaved caspases, cCasp 3, cCasp7 and cCasp 9. Normal and JB12 depleted Huh-7 cells were challenged with indicated proteotoxic stressors for 6-hrs. Levels of indicated proteins were determined by western blot.

Figure 6. Depletion of JB12 from Huh-7 is associated with an increase in BOK and induction of Caspase processing

(A) There is an inverse relationship between BOK and JB12 levels in Huh-7 cells that are challenged with DTT. Huh-7 were treated with JB12 shRNA and/or DTT and levels of indicated proteins were determined by western blot. Blots for gp78 and Erlin-2 serve as load controls (B) Expression of JB12-QPD, a dominant negative form of JB12, promotes BOK accumulation. HEK293 cells were transfected with indicated quantities of wild-type JB12 or JB12-QPD and impacts of each on FLAG-BOK accumulation was determined by western blot. (C and D) FLAG-BOK accumulated in immunoprecipitable complexes that contain QPD-JB12 and gp78. Indicated proteins were expressed in HEK293 cells and detected in native precipitates with FLAG-BOK by western blot.

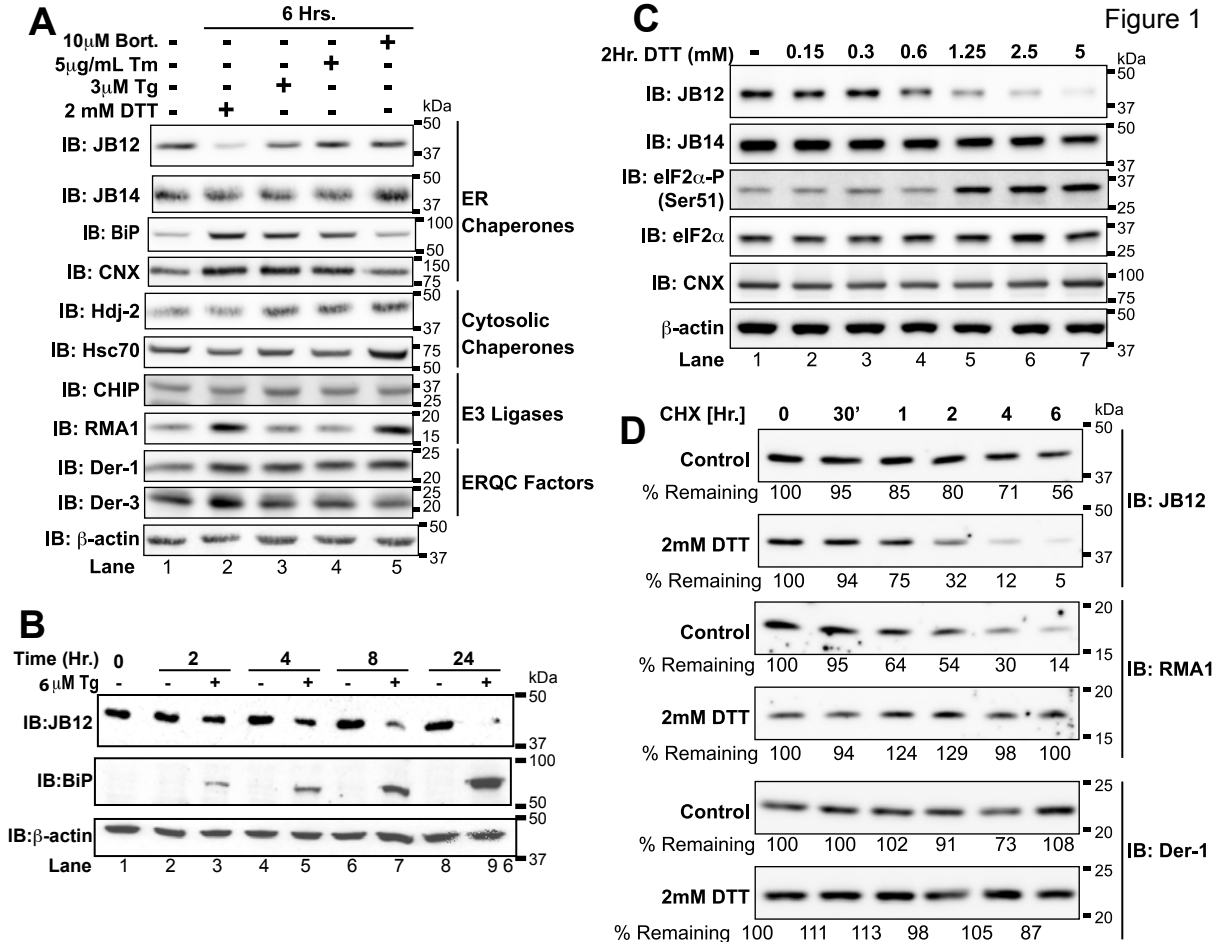
Figure 7. JB12 protects Huh-7 from apoptosis induced by chemical stressors and the chemotherapeutic LCL-161.

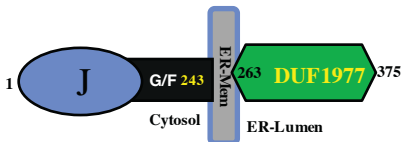
(A) Impact that depletion of JB12 from Huh-7 has on survival from challenge with proteotoxic agents and the apoptosis inducer LCL-161. Changes in cell viability caused by stressors were estimated after 12, 24 and 36 hrs of exposure through assay for the reduction of ATP levels in treated cells (see Materials and Methods for details). Values are normalized as a % of the total ATP in control cells. Statistical significance was evaluated by Student's t-test. A p-value of less than 0.01 was considered statistically significant (**). 2mM DTT 12 Hr. P = 0.0039, 24 Hr. P = <0.0001, 36 Hr. P = <0.0001; 20 uM Bort. 12 Hr. P = < 0.0001; 2 uM LCL-161 12 Hr. P = 0.0026, 24 Hr. P = <0.0001, 36 Hr. P = 0.0078; 20 uM LCL-161 12 Hr. P = <0.0001, 24 Hr. P = <0.0001, 36 Hr. P = <0.0001 (B) The dose dependent impact of LCL-161 on Caspase processing in control and JB12 depleted Huh-7 cells. Shown are western blots of cell extracts where levels of indicated proteins were measured.

Figure 8. Model Depicting how ER stress impacts JB12 and BOK in Normal and Stressed Cells.

During normal growth JB12 cooperates with Hsp70 and gp78 to mediate the constitutive degradation of BOK. Upon inactivation of JB12 during acute and severe ER stress the efficiency of BOK degradation is reduced. The resultant degradation of JB12 and accumulation of BOK then contributes to the induction of ER stress-induced apoptosis.

Figure 1



A**JB12 Domain Structure****Figure 2**

Alignment of the DUF1977 from JB12 and JB14

JB12	263	SQLMVSSPPYSLSPRPSVGHIIHRRVTDHLGVVYYVGDTFSEEYTGSS	307
		: : ...: ...: ...: ...: ...: ...: ...:	
JB14	266	SQLMVSNNPPYSLYPRSGTGQTIKMQTENLGVVYYVNKDFKNEYKGML	310
JB12	308	LKTVERNVEDDYIANLRNNCWKEKQQ	334
		:.. : : : : : : : : : : : :	
JB14	311	LQKVEKSVEEDYVTNIRNNCWKERQQ	338
JB12	335	SEGLLYRARYFGDMDYHRAQKMGTPSCSRLSEVQASLHG	375
		...: : ...: : ...: : ...: : ...: :	
JB14	338	KTDMQYAAKVYRDDRLRRKADALSMDNCKELERLTSLYKGG	379

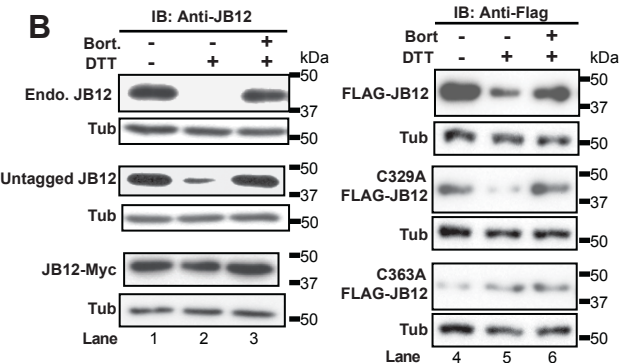
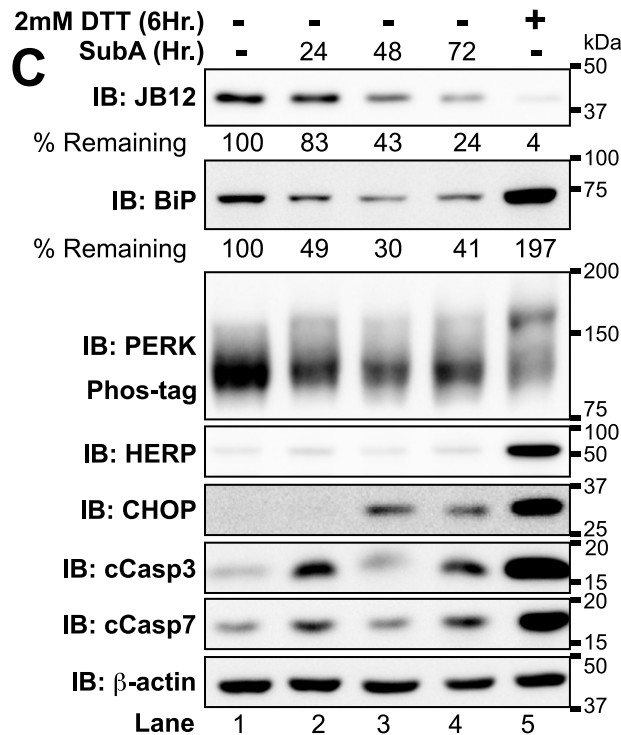
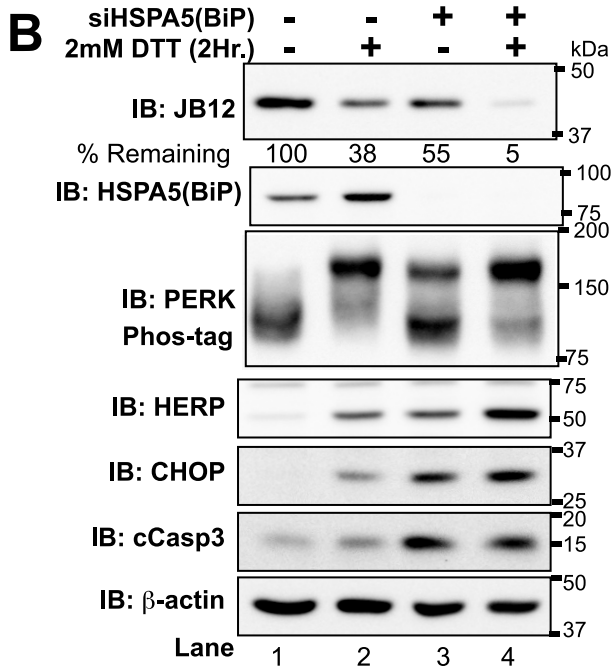
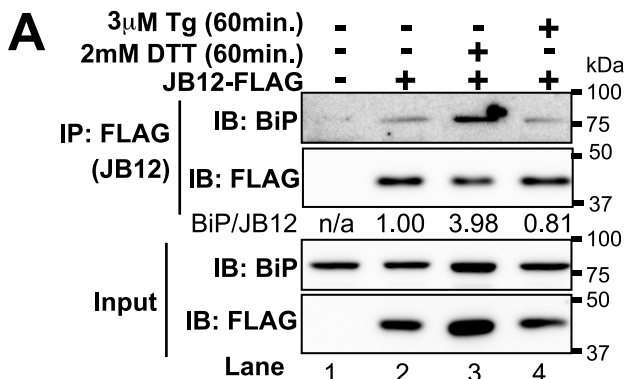
B

Figure 3



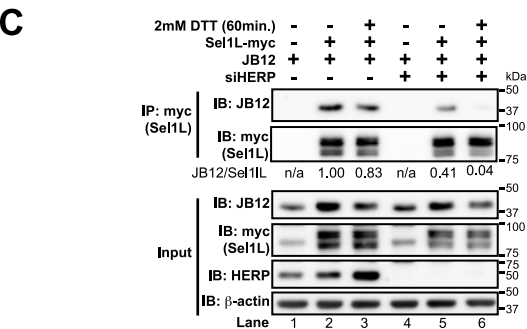
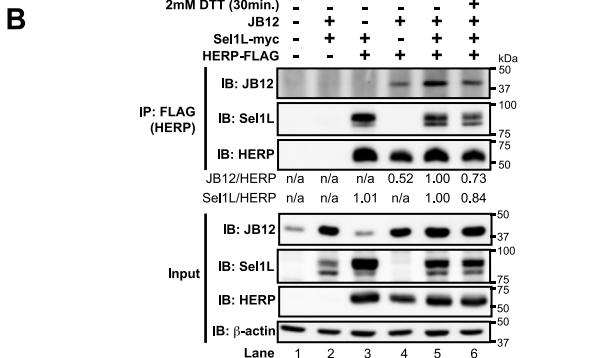
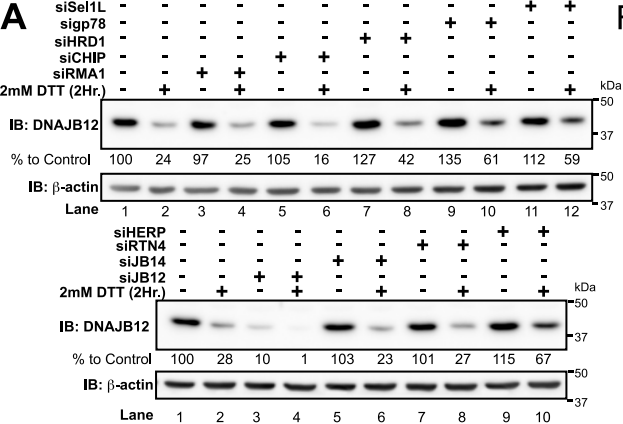
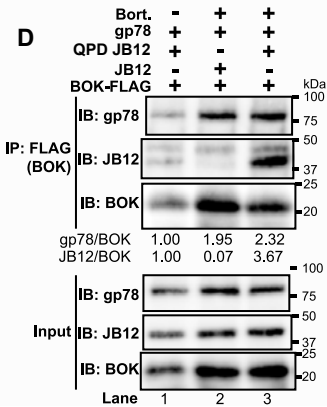
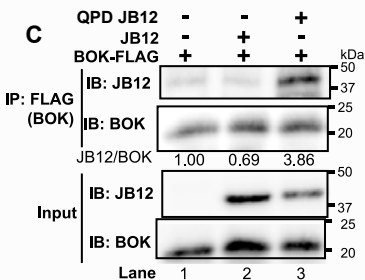
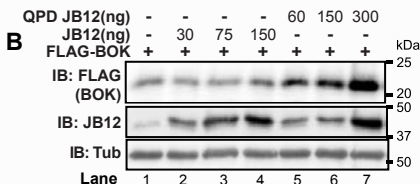
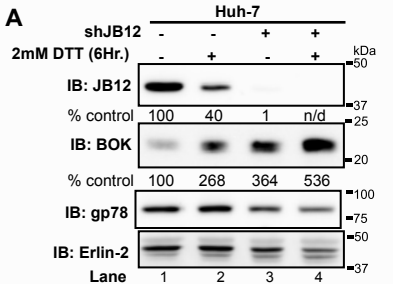
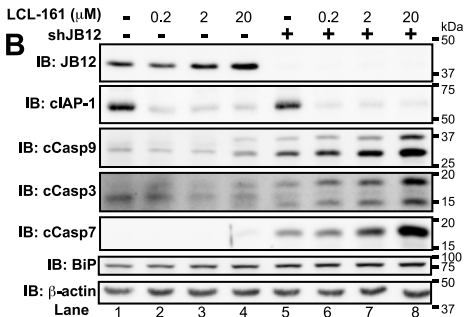
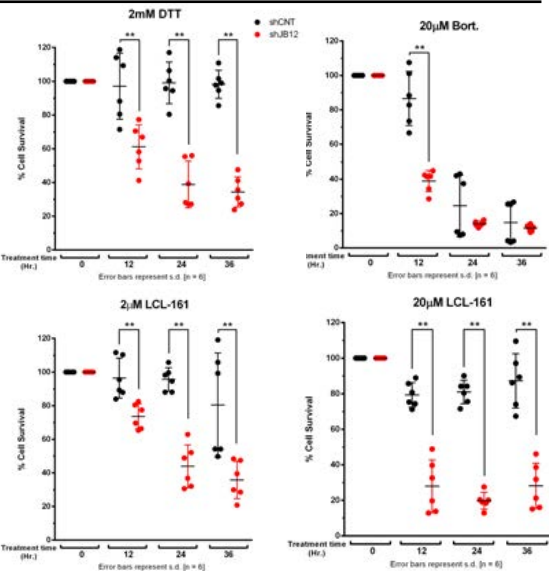
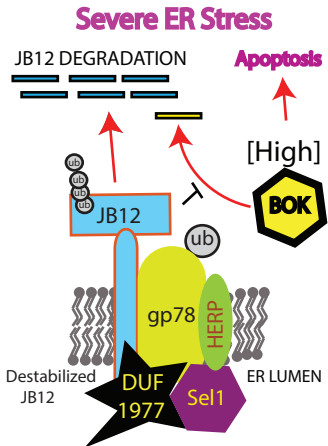
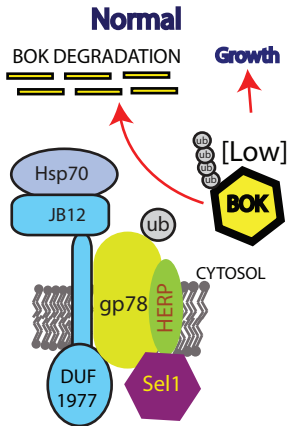


Figure 6



A**Huh-7 Cancer Cells**



**Endoplasmic Reticulum Stress-induced Degradation of DNAJB12 Stimulates BOK
Accumulation and Primes Cancer Cells for Apoptosis**

Pattarawut Sopha, Hong Yu Ren, Diane E. Grove and Douglas M Cyr

J. Biol. Chem. published online May 23, 2017

Access the most updated version of this article at doi: [10.1074/jbc.M117.785113](https://doi.org/10.1074/jbc.M117.785113)

Alerts:

- [When this article is cited](#)
- [When a correction for this article is posted](#)

[Click here](#) to choose from all of JBC's e-mail alerts

Supplemental material:

<http://www.jbc.org/content/suppl/2017/05/23/M117.785113.DC1>

This article cites 0 references, 0 of which can be accessed free at

<http://www.jbc.org/content/early/2017/05/23/jbc.M117.785113.full.html#ref-list-1>