Protective Effects of Dipeptidyl Peptidase-4 (DPP-4) Inhibitor against Increased β Cell Apoptosis Induced by Dietary Sucrose and Linoleic Acid in Mice with Diabetes*

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Chronic exposure to high glucose and fatty acid levels caused by dietary sugar and fat intake induces β cell apoptosis, leading to the exacerbation of type 2 diabetes. Oleic acid and linoleic acid are two major dietary fatty acids, but their effects in diabetes are unclear. We challenged mice to the exacerbation of type 2 diabetes. Oleic acid and linoleic acid are the most abundant fatty acids among the total plasma fatty acids, plasma non-esterified fatty acids, plasma triacylglycerol, phospholipids, and plasma cholesterol esters (3). Palmitic acid is a well known inducer of β cell lipotoxicity. However, the effects of oleic acid and linoleic acid, two major unsaturated fatty acids, on β cells remain obscure. We therefore selected two diet protocols, namely SO3 and SL, to examine the effects of oleic acid and linoleic acid on pancreatic islets (4). The main components of the SO diet are sucrose and oleic acid, whereas those of the SL diet are sucrose and linoleic acid. Both of these diets contain similar amounts of palmitic acids (4).

Desfluorositagliptin (DFS), a dipeptidyl peptidase-4 (DPP-4) inhibitor, acts by inhibiting the breakdown of many regulatory peptides including glucagon-like peptide-1 (GLP-1) (5). The clinically beneficial effects of DPP-4 inhibition on β cells cannot be fully explained by the increase in insulin release alone, and other mechanisms are thought to affect β cell mass and β cell apoptosis (6–9). Treatment with DFS increases the number of insulin-positive β cells in the islets, leading to the normalization of the β cell mass and the β cell-to-α/β cell ratio in mice with high fat diet-streptozotocin-induced diabetes, a mouse model with defects in insulin sensitivity and secretion (10). However, the beneficial effect of DPP-4 inhibition on nutrient-induced β cell apoptosis remains poorly understood.

Haploinsufficiency of β cell-specific glucokinase (Gck+/−) causes impaired insulin secretion in response to glucose in mice fed a standard chow diet despite the mice having a normal β cell mass (11, 12). In this study, we used Gck+/− mice to evaluate the impact of diet on β cell damage in the diabetic state and inves-

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2 The abbreviations used are: SO, sucrose and oleic acid; SL, sucrose and linoleic acid; ATF4, activating transcription factor 4; Bip, immunoglobulin heavy chain-binding protein; C/EPP, CCAAT/enhancer-binding protein; CHOP, C/EPP-homologous protein; DFS, desflurositagliptin; DPP-4, dipeptidyl peptidase-4; ER, endoplasmic reticulum; Gck, glucokinase; GLP-1, glucagon-like peptide-1; IRS, insulin receptor substrate; elf2α, eukaryotic initiation factor 2α; SREBP-1c,sterol regulatory element-binding protein-1c.

3 The on-line version of this article (available at http://www.jbc.org) contains supplemental Figs. 1–6.

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6 The online version of this article (available at http://www.jbc.org) contains supplemental Figs. 1–6.
tigated whether DFS prevented nutrient-induced β cell apoptosis.

**EXPERIMENTAL PROCEDURES**

**Animals and Animal Care**—We backcrossed Gck
\(^{+/−}\)
mice, which were generated by disrupting the β cell- and brain-specific exon (11), with C57Bl/6 mice more than 10 times. Both the wild-type (WT) and Gck
\(^{+/−}\)
mice were fed a standard chow diet (MF, Oriental Yeast, Tokyo, Japan) until 8 weeks of age and then were given free access to the experimental diets. All the experiments were conducted on male littermates. All the animal procedures were performed in accordance with the guidelines of the Animal Care Committee of Yokohama City University. Animal housing rooms were maintained at a constant temperature (25 °C) and a 12-h light (7:00 a.m.)/dark (7:00 p.m.) cycle.

**Diets**—The compositions of the SO and SL diets are described in Table 1. The fat component of the SO and SL diets was derived from safflower oil and high oleic sunflower oil blended with perilla oil, respectively. The two diets were identical except for the type of fat used: oleic acid in the SO diet and linoleic acid in the SL diet. Both diets contained similar amounts of palmitic acids. The experimental diets were freshly prepared weekly. DFS was administrated orally by premixing with SL to a concentration of 1.1% (13).

**In Vivo Physiological Studies**—Plasma glucose levels and blood insulin levels were determined using Glutest Neo Super (Sanwa Chemical Co., Kanagawa, Japan) and an insulin kit (Morinaga Institute of Biological Science, Yokohama, Japan), respectively. Active GLP-1 was assayed using a glucagon-like peptide-1 (active) ELISA kit (Millipore). Glucose-stimulated insulin secretion by islets—Islets were isolated as described elsewhere (12). Isolated islets were cultured overnight in RPMI 1640 medium containing 11 mM glucose supplemented with 10% fetal calf serum (FCS), 100 units/ml penicillin, and 100 µg/ml streptomycin. Total RNA was isolated from pancreatic islets using an RNase-free DNase and RNaseasy kit (Qiagen). cDNA was prepared using the TaqMan reverse transcriptase kit (Applied Biosystems) and subjected to quantitative PCR by performing TaqMan Gene Expression Assays with Universal Master Mix (7500 real time PCR system, Applied Biosystems). All the probes were purchased from Applied Biosystems. Each quantitative reaction was carried out in duplicate. Data were normalized according to the β-actin level.

**Immunoblotting**—For immunoblotting, isolated islets (100 islets) or MIN6 cells were lysed in ice-cold radioimmunoprecipitation assay buffer (Cell Signaling Technology) with complete protease inhibitor mixture (Roche Diagnostics). After centrifugation, the extracts were subjected to immunoblotting with antibodies to CHOP, activating transcription factor 4 (ATF4), C/EBP-β, phosphorylated eIF2α (Santa Cruz Biotechnology), Bcl-2, Bax, cleaved caspase-3 (Cell Signaling Technology), and β-actin (Sigma-Aldrich). Densitometry was performed using NIH Image software.

**Glucose-stimulated Insulin Secretion by Islets**—Islets were isolated with Liberase RI (Roche Diagnostics). Immediately or after culturing for 12 h in RPMI 1640 medium containing 11 mM glucose supplemented with 10% fetal calf serum, 100 units/ml penicillin, and 100 µg/ml streptomycin (Sigma), 10 islets were incubated at 37 °C for 1.5 h in Krebs-Ringer bicarbonate buffer containing 2.8, 8.3, or 22.2 mM glucose with or without 100 nM exendin-4 (Sigma). The insulin concentration of the assay buffer was measured using an insulin ELISA kit (Morinaga Institute of Biological Science).

**Treatments of Islets or MIN6 Cells with Fatty Acid and Other Chemicals**—Oleic acid, linoleic acid, and palmitic acid were purchased from Sigma. Isolated islets were incubated in RPMI 1640 medium (1000 or 2500 mg/liter glucose) with 0.25 mmol/liter fatty acid plus 0.5% (w/v) BSA or 0.5% (w/v) BSA alone as a
control. MIN6 cells were incubated in DMEM (4500 or 1000 mg/liter glucose; Sigma) with 0.25 or 0.5 mmol/liter fatty acid plus 0.5% (w/v) BSA. The preparation of the free fatty acid media has been described elsewhere (14). Drugs were added to the regular growth medium. Exendin-4 was dissolved in PBS, and tunicamycin (Sigma) was dissolved in DMSO.

**Analysis of Fatty Acid Composition**—Total lipid was extracted from isolated islets (400 islets/sample) or fatty acid-treated MIN6 cells with H2O:chloroform:methanol (0.7:1:1, v/v) containing butylated hydroxytoluene as an antioxidant according to the method of Bligh and Dyer (15). The phospholipids were separated by preparative layer chromatography (silica gel 60F254, 2-mm thickness, Merck) and developed with a solvent of hexane:isopropanol: methanol (9:1:1, v/v). The phospholipid fractions were transmethylated with HCl/methanol at 100 °C for 2 h. The fatty acid methyl esters were separated in a gas-liquid chromatograph (GC-18A, Shimadzu, Kyoto, Japan) with a capillary column (SP2330, Supelco, Bellefonte, PA). Individual fatty acids were identified by comparing the retention time of each peak with those of external standards.

**Propidium Iodide/DAPI and Annexin V Cell Death Assays**—MIN6 cells were stimulated for 24 h with vehicle alone or with fatty acids at the indicated concentrations. For the last hour of incubation, 10 μg/ml propidium iodide was added directly to the medium. After the incubation, the cells were fixed and mounted on glass slides with VECTASHIELD with DAPI (Vector Laboratories). The cell images were acquired using a BZ-9000 microscope (Keyence). The percentage of cells that stained was performed using an Annexin V-FITC Apoptosis Detection kit (Sigma) and analyzed using FACSCanto II (BD Biosciences). The proportions of Annexin V-FITC-positive and propidium iodide-negative apoptotic cell fractions were calculated using FACSDiva software (BD Biosciences).

**Statistical Analyses**—All the data are reported as the mean ± S.E. and were analyzed using the Student t test or analysis of variance. Differences were considered significant if the p value was <0.05.

**RESULTS**

Sucrose and Linoleic Acid Diet-induced β Cell Endoplasmic Reticulum (ER) Stress and Apoptosis in Gck+/– Mice—WT mice and Gck+/– mice fed an SO diet or an isocaloric SL diet for 25 weeks were evaluated. No significant differences in body weight gain, fasting blood glucose level, or food intake were observed between the SO group and the SL group of either genotype (16). After 25 weeks, the β cell mass (Fig. 1A) and relative β cell mass as a proportion of the total α cell plus β cell mass (Fig. 1B) were significantly lower in the Gck+/– mice fed the SL diet than in the Gck+/– mice fed the SO diet. The relative loss of β cells was associated with the abnormal distribution of α cells in the islets of the SL-fed Gck+/– mice (supplemental Fig. 1A). Of note, both alterations occurred despite a normal body weight and no exacerbation of insulin sensitivity in the Gck+/– mice (16), and such alterations were not seen in Gck+/– mice fed a high fat diet (HFD32, Clea Japan, Inc.) (12). A similar β cell mass and number of β cells as a proportion of all the islet cells were observed between the SO diet group and SL diet group of WT mice or insulin receptor substrate-1-deficient (IRS-1−/−) mice (Fig. 1, A and B, and data not shown). These results raise the possibility that hyperglycemia as well as fatty acid-induced lipotoxicity is required for β cell abnormalities to develop.
...indicate S.E.*, the indicated molecules in the islets were determined using real time quantitative RT-PCR and were normalized to the level of staining pancreatic (18, 19). We assessed ER stress and apoptosis by immunostaining. More CHOP-positive nuclei and TUNEL-positive apoptotic nuclei were detected in Gck+/− mice fed the SL diet than in mice fed the SO diet (Fig. 1, C and D, and supplemental Fig. 1, B and C). To assess β cell proliferation, we counted the number of Ki67-positive cells; no significant differences in the number of Ki67-positive β cells were observed between the SO group and the SL group among either the WT or Gck−/− mice (Fig. 1, E and supplemental Fig. 2, B and C). The proportion of Ki67-positive proliferating cells is shown as a percentage of the total number of insulin-positive cells in the sections (n = 5). D and E, more than 100 islets were counted in each mouse. F, the mRNA expression levels of the indicated molecules in the islets were determined using real time quantitative RT-PCR and were normalized to the level of β-actin mRNA (n = 5). The experiments were performed in WT mice and Gck−/− mice after 20 weeks on the SL diet or the SL plus DFS diet. Black bars, SL; hatched bars, SL + DFS. Error bars indicate S.E. *, p < 0.05.

DFS Protected against Sucrose- and Linoleic Acid-induced β Cell ER Stress and Apoptosis in Gck+−/− Mice—Not only does GLP-1 receptor signaling preserve the β cell mass in experimental diabetic states, it also directly modulates the ER stress response, promoting β cell adaptation and survival (21–23). To evaluate DFS, a DPP-4 inhibitor, as a treatment for diet-induced β cell damage, we performed a 20-week study of SL or SL plus 1.1% DFS diet-fed WT and Gck−/− mice. At the end of the 20-week DFS treatment period, glucose tolerance and insulin secretion after glucose loading were significantly improved by DFS monotherapy in Gck+−/− mice (16).

The treatment of SL-fed Gck−/− mice with DFS for 20 weeks produced a significant increase in the β cell mass (Fig. 2A) and the relative β cell mass as a proportion of the total α cell plus β cell mass (Fig. 2B), and the abnormal distribution of pancreatic α cells was also corrected (supplemental Fig. 2A). DFS significantly reduced the proportion of CHOP-positive nuclei and TUNEL-positive apoptotic nuclei among the β cells of Gck−/− mice (Fig. 2, C and D, and supplemental Fig. 2, B and C). The proportion of single-stranded DNA-positive apoptotic nuclei was also reduced by DFS (data not shown). At week 20, the proportion of Ki67-positive β cells in the islets of Gck−/− mice fed the SL diet plus DFS was much higher than in those fed the SL diet alone (Fig. 2E and supplemental Fig. 2D). Next, we examined the levels of CHOP, Bip, SREBP-1c, IRS-2, and E-cadherin mRNA expression in the islets (Fig. 2F). Lower levels of CHOP, Bip, and SREBP-1c were expressed in the islets of...
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To assess the acute effects of DFS on GLP-1 release, we performed an oral SL plus 1.1% DFS meal tolerance test (12 mg/g of body weight) in standard chow diet-fed WT and Gck+/− mice. The results showed that the meal-induced rise in the blood glucose level is significantly higher in Gck+/− mice than in WT mice and that the corresponding rise in the serum insulin level is significantly lower (Fig. 5, A and B). In addition, there was a meal-induced rise in serum active GLP-1 of a similar magnitude in both WT and Gck+/− mice that was significantly greater in mice treated with DFS compared with untreated mice (Fig. 5C).

To assess the acute effects of DFS on GLP-1 release, we performed an oral SL plus 1.1% DFS meal tolerance test (12 mg/g of body weight) in these mice. The serum insulin level is significantly lower (Fig. 5, A and B). In addition, there was a meal-induced rise in serum active GLP-1 of a similar magnitude in both WT and Gck+/− mice that was significantly greater in mice treated with DFS compared with untreated mice (Fig. 5C).
levels were also determined using these samples. The blood glucose and serum insulin of whole blood to measure the biologically active form of GLP-1, blood was hatched bars SL; SO-fed and the SL-fed groups in both WT and in the serum active GLP-1 concentration at 30 min between the diet for 20 weeks. The results showed no significant differences in the WT mice.

Changes in blood glucose, insulin, and active GLP-1 concentrations at 0 (fasted > 20 h), 30, and 120 min after the oral administration of the SO diet, SL diet, or SL plus DFS diet (12 mg/g of body weight) to 6–8-week-old WT mice and Gck\(^{+/−}\) mice (n = 4–5). To obtain a sufficient amount of whole blood to measure the biologically active form of GLP-1, blood was collected from the inferior vena cava under anesthesia with a DPP-4 inhibitor (Millipore) at the time point indicated. The blood glucose and serum insulin levels were also determined using these samples. White bars, SO; black bars, SL; hatched bars, SL + DFS. Error bars indicate S.E. *, p < 0.05.

To investigate the long term effects of an SO or SL diet on GLP-1 release, we performed an oral SO or SL meal tolerance test in WT and Gck\(^{+/−}\) mice after the mice had been fed either diet for 20 weeks. The results showed no significant differences in the serum active GLP-1 concentration at 30 min between the SO-fed and the SL-fed groups in both WT and Gck\(^{+/−}\) mice (supplemental Fig. 4). Interestingly, after 20 weeks of the SO or SL diet, GLP-1 release in the Gck\(^{+/−}\) mice was higher than that in the WT mice.

We also evaluated the insulinotropic effect of GLP-1 receptor signaling on islets by analyzing-stimulated insulin secretion in the presence or absence of a potent GLP-1 receptor agonist, exendin-4, at 2.8, 8.3, or 22.2 mM glucose (supplemental Fig. 5). Experiments were performed using two protocols: (a) immediately after islet isolation and (b) after a 12-h culture in medium containing 11 mM glucose after islet isolation. Exendin-4 did not affect glucose-stimulated insulin secretion at 2.8 mM glucose and tended to increase glucose-stimulated insulin secretion at 22.2 mM glucose in both genotypes of islets. The increase in insulin secretion by exendin-4 in response to 8.3 mM glucose was much lower in the Gck\(^{+/−}\) islets than in the WT islets.

Requirement of High Glucose Concentration for Linoleic Acid-induced β Cell ER Stress and Apoptosis in Vitro and Mitigation by GLP-1 Receptor-mediated Signals—Palmitic acid and, to a lesser extent, oleic acid induce ER stress and apoptosis in vitro (17), and GLP-1 receptor signaling activated by exendin-4 prevents ER stress and apoptotic cell death in β cells both in vivo and in vitro (21). We investigated whether linoleic acid induced ER stress and apoptosis and whether GLP-1 receptor signaling regulated fatty acid-induced ER stress and apoptosis in pancreatic islets and β cell lines.

Pancreatic islets from WT mice were exposed to fatty acids for 16 h in the presence of 5.5 or 13.9 mM glucose, and mRNA expression levels were analyzed (Fig. 6). The expressions of CHOP, Bip, and C/EBP-β were increased to a greater degree by fatty acids at 13.9 mM glucose than at 5.5 mM glucose. Linoleic acid significantly increased the expressions of CHOP and ATF4 compared with oleic acid at 13.9 mM glucose. However, the expressions of SREBP-1c and E-cadherin did not change significantly in vitro. The Bax/Bcl-2 ratio was also significantly increased in linoleic acid-treated islets at 13.9 mM glucose. In Gck\(^{+/−}\) islets, the inductions of ER stress-related genes and apoptosis-related genes by fatty acids were similar to those in WT islets (data not shown). These results suggested that linoleic acid and high glucose act synergistically to induce β cell ER stress and apoptosis. Furthermore, exendin-4 significantly decreased the Bax/Bcl-2 ratio and C/EBP-β but did not affect the expressions of CHOP, Bip, and ATF4 (Fig. 6).

MIN6 cells were also exposed to fatty acids for 24 h. In the presence of 25 mM glucose, linoleic acid up-regulated the expressions of CHOP and cleaved caspase-3 more efficiently than oleic acid. However, such differences were not observed in the presence of 5.5 mM glucose (Fig. 7A). These results supported the hypothesis that a high glucose concentration was required for the sufficient induction of ER stress and apoptosis by linoleic acid. The expressions of ATF4 and phosphorylated eukaryotic initiation factor 2α (p-eIF2α) increased to a greater degree in the presence of 0.25 and 0.5 mM linoleic acid than in the presence of the same concentration of oleic acid but to a lesser degree than in the presence of the same concentration of palmitic acid in the presence of 25 mM glucose (Fig. 7B). Tunicamycin markedly induced the expression of CHOP and the phosphorylation of eIF2α. Exendin-4 increased the magnitude of CHOP and ATF4 induction in the presence of each fatty acid; however, the level of phosphorylated eIF2α induced by linoleic acid or palmitic acid was reduced by exendin-4. These findings were consistent with a previous report that exendin-4 potentiated the ATF4−CHOP-GADD34-mediated ER stress pathway and promoted translational recovery from the ER stress-mediated repression of protein synthesis by reducing the levels of phospho-eIF2α (21).

We next investigated fatty acid-induced cell death using propidium iodide staining in the presence and absence of exen-
More cell death was observed after exposure to 0.5 mM linoleic acid than after exposure to 0.5 mM oleic acid, and exendin-4 significantly reduced the linoleic acid-induced cell death (Fig. 7C and supplemental Fig. 6A). Annexin V staining revealed that the rescue from linoleic acid-induced cell death by extendin-4 was at least in part attributable to the inhibition of apoptosis (Fig. 7D and supplemental Fig. 6B). We also analyzed fatty acid composition in MIN6 cells after exposure to fatty acids for 24 h. Arachidonic acid (C20:4) and its precursors 11,14-eicosadienoic acid (C20:2) and 11,14,17-eicosatrienoic acid (C20:3) were significantly increased after exposure to 0.5 mM linoleic acid (C18:2) compared with exposure to 0.5 mM oleic acid (C18:1) or vehicle control in MIN6 cells (Fig. 7E). Treatment with exendin-4 did not affect the compositions of arachidonic acid and its precursors in linoleic acid-exposed MIN6 cells (Fig. 7E).

**DISCUSSION**

In this study, we established a model in which a diet rich in sucrose and linoleic acid induced the apoptosis of β cells in Gck−/− mice. Because considerable and progressive deficits in the β cell mass supposedly occur during the development of human type 2 diabetes before the manifestation of patent hyperglycemia (26, 27), our model should be a good model for the diet-induced exacerbation of diabetes. SL was found to exacerbate β cell ER stress and apoptosis in Gck−/− mice but not in euglycemic WT or IRS-1−/− mice, a finding that was consistent with previous reports that fatty acid-induced β cell ER stress is amplified by high glucose concentrations (28, 29).

We also demonstrated that high glucose concentrations were required for linoleic acid-induced β cell apoptosis in vitro. Because the fatty acid-induced ER stress response is reportedly not modified by high glucose concentrations (30), a factor(s) other than glucose may be involved in linoleic acid-induced β cell ER stress in diabetic Gck−/− mice. Linoleic acid is reportedly converted to arachidonic acid in vivo, and arachidonic acid is converted to prostaglandins, leukotrienes, and other lipid mediators by lipoxygenase or cyclooxygenase (31). In fact, we noted that the content of arachidonic acid in islets was significantly increased in mice fed the SL diet compared with those fed the SO diet. SL diet-induced β cell apoptosis may be mediated in part by the activation of the arachidonic acid cascade. This hypothesis is supported by reports that SREBP-1c-induced calcium-independent phospholipase A2 (iPLA2β) plays a key role in spontaneous ER stress and apoptosis in β cells (32, 33). To clarify the precise mechanisms for SL diet-induced β cell apoptosis, further study using inhibitors of cyclooxygenase or lipoxygenases and mice deficient in phospholipases A2 will be required. A palmitic acid-rich diet may also contribute to...
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**FIGURE 7. Effects of exendin-4 on ER stress and apoptosis induced by combination of high glucose and linoleic acid in MIN6 cells.**

**A.** Upper panel, mouse MIN6 insulinoma cells were treated with vehicle (ctl), linoleic acid (LA), or oleic acid (OA) in the presence of 25 or 5.5 mM glucose for 24 h. Total cell extracts (10 \(\mu\)g) were subjected to immunoblotting for CHOP, cleaved caspase-3, and \(\beta\)-actin as indicated. Lower panel, intensities of the CHOP signal and cleaved caspase-3 signal quantified by densitometry (NIH ImageJ) and corrected by the intensity of the \(\beta\)-actin signal. The results shown are the means of three independent experiments.

**B.** MIN6 cells were treated with the vehicle (ctl), oleic acid, linoleic acid, palmitic acid (PA), or tunicamycin (Tm) in the absence or presence of 50 nM exendin-4 (Ex-4) in the presence of 25 mM glucose for 24 h. The final concentration of tunicamycin was 0.5 \(\mu\)g/ml. Total cell extracts (10 \(\mu\)g) were subjected to immunoblotting and quantified as in A. Results shown are the means of three independent experiments.

**C.** Quantitation of the percentage of propidium iodide-stained nuclei relative to the proportion of DAPI-stained nuclei of MIN6 cells treated with the vehicle alone (ctl) or the 0.5 mM oleic acid or linoleic acid in 25 mM glucose for 24 h in the absence or presence of 50 nM exendin-4.

**D.** Flow cytometric analysis of the proportion of Annexin V-FITC-stained and propidium iodide-negative apoptotic MIN6 cells treated with vehicle alone (ctl) or the 0.5 mM oleic acid or linoleic acid in 25 mM glucose for 24 h in the absence or presence of 50 nM exendin-4.

**E.** Fatty acid content in MIN6 cells treated with vehicle alone (ctl) or the 0.5 mM oleic acid or linoleic acid in 25 mM glucose for 24 h in the absence or presence of 50 nM exendin-4. Error bars indicate S.E., *, \(p < 0.05\).
further elucidation of the mechanism of nutrient-induced β cell lipotoxicity.

The results of this study also showed that DFS protected against SL diet-induced β cell ER stress and apoptosis in Gck \(^{+/−}\) mice. GLP-1 and an analog have been reported to prevent β cell ER stress and apoptosis (21, 22, 28). Despite the massive increase in active GLP-1 induced by a single administration of DFS in both WT and Gck \(^{+/−}\) mice, their serum insulin and blood glucose levels remained largely unchanged. The glucose-dependent insulinotropic effects of GLP-1 may be attenuated by normoglycemia in WT mice. Because glucokinase is the major glucose sensor of β cells, we hypothesized that glucokinase is also a key molecule in the glucose-dependent insulinotropic action of GLP-1 and that the elevated GLP-1 level was still insufficient to stimulate insulin secretion in Gck \(^{+/−}\) mice. Consistent with this hypothesis, a GLP-1 receptor agonist attenuated the effects of incretin on insulin secretion in response to 8.3 mM glucose from Gck \(^{+/−}\) islets compared with WT islets. Because DFS or exendin-4 in mice or MIN6 cells did not affect increased arachidonic acid content induced by linoleic acid, the protective effects of DFS or exendin-4 might be independent of the modulation of the arachidonic acid cascade. Whether an increase in glucose-dependent insulinotropic polypeptide in response to DPP-4 inhibitor contributes to the protection of β cells remains unexplored.

The islet architecture of Gck \(^{+/−}\) mice was restored, and the reduction of E-cadherin expression was recovered in mice fed the SL diet plus DFS. E-cadherin expression on β cells plays an important role in glucose-stimulated insulin secretion (34–36). Because E-cadherin-mediated cell adhesion is controlled by β-catenin and the Wnt signaling pathway (37) and an interaction between GLP-1 signaling and the TCF7L2-dependent Wnt signaling pathway has been reported in pancreatic β cells (38–40), the Wnt/β-catenin network may be involved in the DFS-mediated normalization of islet architecture and E-cadherin expression. SREBP-1c activation caused by ER stress has been implicated in β cell lipotoxicity (41, 42). Although the SREBP-1c mRNA level was significantly increased in the Gck \(^{+/−}\) mice fed the SL diet, the expressions of SREBP-1c and E-cadherin in vitro were not changed by the fatty acids. This discrepancy raised the possibility that the ER stress signaling mechanisms may differ notably between in vivo (chronic reaction) and in vitro (acute reaction) situations. Indeed, a previous study demonstrated that the expression of CHOP was decreased in vivo by treatment with exendin-4 but was increased in vitro under ER stress (21).

We previously reported a reduction in amyloid deposition in Gck \(^{+/−}\) islets (43), suggesting that Gck \(^{+/−}\) islets may be partially protected against glucose toxicity via the glycolytic pathway. Consequently, the high glucose-induced amplification of ER stress and apoptosis in our model might be independent of glucokinase signaling. On the other hand, β cell glucokinase plays a dominant role in the induction of IRS-2 in response to high fat diet-induced insulin resistance, and the antiapoptotic effects of exendin-4 have been implicated in the enhancement of the IRS-2/Akt signaling pathway (12, 44). We also observed an increase in the expression of IRS-2 in response to tumorycin- or fatty acid-induced ER stress in MIN6 cells and isolated islets.\(^3\) Although the SL diet or DFS administration did not affect the mRNA expression of IRS-2 in the Gck \(^{+/−}\) islets in this study, insulin signaling might be involved in the regulation of ER stress and apoptosis via fatty acids or GLP-1 receptor signaling. Thus, the modifications of ER stress by GLP-1 receptor activation in vivo have been controversial. Therefore, further research is needed to clarify the link between DPP-4 inhibition and ER stress in our model.

In summary, we created a model of nutrient-induced β cell apoptosis in a diabetic state and showed that DPP-4 inhibition with DFS ameliorated apoptosis. The results of the current study demonstrate the novel therapeutic potential of DPP-4 inhibitors for the treatment of diabetes.

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